

ABSTRACT BOOK

https://meetanyway.com/events/molecular-origins-of-life-munich-2020/space



MOLECULAR ORIGINS OF LIFE CRC 235 - EMERGENCE OF LIFE CONFERENCE . ONLINE CONFERENCE . 8<sup>™</sup>-10<sup>™</sup> Jul 2020

# IMPRINT

Scientific Organizer : Dieter Braun (LMU Munich) Academic Organizer : Filiz Çivril (Collaborative Research Center 235 - Emergence of Life)

This event is organized by CRC 235 - Emergence of Life management and financially supported by the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation) through Project-ID 364653263 – TRR 235 and under Germany's Excellence Strategy - EXC-2094 - 390783311

CRC 235 : http://www.emergence-of-life.de/ Origins Clusters : https://www.origins-cluster.de/en/

emergenceoflife@lmu.de@emergenceoflife

# CONTACT

### Filiz ÇIVRIL, PhD

CRC 235-Emergence of Life Management Ludwig-Maximilians-Universität München Schellingstr. 4 D- 80799 München, Germany +49 89 - 2180 3514 (Tel) +49 89 - 2180 17303 (Fax) emergenceoflife@lmu.de (Email)

# INDEX

04	WELCOME ADDRESS
05	CODE OF CONDUCT
06	EVENT PLATFORM MAP & GUIDE
08	EVENT PLATFORM FAQS
10	PARTICIPATING IN TALKS (ZOOM W
10	PARTICIPATING IN ZOOM MEETING
12	CONFERENCE SCHEDULE
14	SPEAKERS INDEX
16	SPEAKER ABSTRACTS
40	POSTERS BY TIME ZONE / FLOOR
46	POSTER PRESENTERS INDEX
	POSTER ABSTRACTS
48	FLOOR 3
61	FLOOR 4
73	FLOOR 5
85	FLOOR 6
97	FLOOR 7
109	FLOOR 8
121	FLOOR 9
133	FLOOR 10
145	FLOOR 11
158	FLOOR 12
172	PARTICIPANT DIRECTORY
189	CREDITS





### **WELCOME ADDRESS** by Dieter Braun

Dear Emergence of Life researcher!

Evolution means to adapt to changing boundary conditions. The virtual format of the Molecular Origins of Life, Munich (MOM) conference offers many positive aspects.

This year we will be hosting many more attendees than the previous years. And the time you lose at the airports and for transfers can now be used for scientific discussions.

Besides the short talks and the interactive panel discussion format you know from previous MOMs, this time we will also have 'Meet the Speakers' sessions and what I think will be very productive; parallel poster sessions along with the talks using your browser. So besides listening to the talks over Zoom, you can look around and chat with the poster presenters.

In the evening, we will have a virtual hangout to chat and have a drink and visit more posters.

As we are learning to communicate over screens in a pleasant way, our otherwise distributed collaboration network can use this opportunity to get closer. While in the past it was difficult to hold meetings with more than 2 PIs at one location, meeting with many more across the globe is becoming very efficient.

The field of Emergence of Life has a small number of very specialized experts distributed all over the world. We should take the best out of Corona lockdown; reach out to neighboring experts, trigger collaborations and make more connections. I am optimistic that this difficult situation will have the positive effect of catalyzing the formation of 'Emergence of Life' as a scientific field similar to any other specialization.

Looking forward to meeting you!

Dieter Braun.

# CODE OF CONDUCT

Inappropriate/illegal behavior and/or harassment of any kind will not be tolerated at Molecular Origins of Life, Munich virtual conference.

- This includes, but not limited to, .
- individuals without consent screenshot
- . and/or recordings of scientific content without consent

Participants who refuse to follow this code can face temporary/ permanent ban from the Molecular Origins of Life, Munich virtual conference and other CRC235 - Emergence of Life events.

comments and content that are offensive or inflammatory due to gender, gender identity or expression, race, religion, ethnicity, lifestyle, age, physical appearance or disability inappropriate contact, sexual attention or innuendo, deliberate intimidation, stalking, and screenshots and/or recordings of

# EVENT PLATFORM MAP & GUIDE

### AUDITORIUM (Floor 0)

STAGE Zoom Webinar connection to the Conference Talks & Panel Discussions

### **PUZZLE & BEER OF THE DAY**

Zoom Meeting connection to informal chat after all the talks, each speaker has a bottle of beer and a puzzle of the day to start the conversation

**INFORMATION** Files and contact info

SUPPORT Technical support tables for the platform

MEET THE SPEAKERS (Floor 1) Zoom Meeting connection to chat with the speakers

**NETWORKING (Floor 2)** Tables to meet the other conference participants either open networking or chats on # topics

FLOORS 3-12 (POSTER PRESENTATION ROOMS)

### FLOOR 3

EF D'É

Presenters from time zones UTC + 04:30 to 12:00 - Visit at early hours of the event. The presenters will probably not be available during the poster session.

FLOOR 4 - 10 Presenters from central Europe - Aligns with meeting schedule

Floor 11 & 12 Presenters from time zones UTC -04:00 to -07:00 - Visit at late hours of the event or during poster session

Shall you have problems to connect to the event space: Email: erich@meetanyway.com Chat: https://nextcloud.christofmast.de/call/n88vjxtn (Password for chat: 171766)

### AUDITORIUM

Stage (Zoom Webinar)

### Room 1 / Date Speaker Names

MEET THE SPEAKERS

2:

=

==

( = )

Room 2 / Date

Speaker Names

Room 3 / Date **Speaker Names** 

Room 4 / Date **Speaker Names** 

### NETWORKING

**Open Networking** Just grab a seat somewhere and interact.

#astrophysical origins Just grab a seat somewhere and interact.

# early earth Just grab a seat somewhere and interact.

# RNA/DNA world Just grab a seat somewhere and interact.

# emergence of metabolism Just grab a seat somewhere and interact.

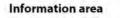
# prebiotic chemistry Just grab a seat somewhere and interact.

# systems chemistry Just grab a seat somewhere and interact.

# synthetic biology Just grab a seat somewhere and interact.



nttps://meetanyway.com/events/molecularrigins-of-life-munich-2020/space



Support



ŏ

11

( ==















**FLOOR 3-12** 

Presenter Name(s) Poster title



Presenter Name(s) Poster title



( 11

41

-

11

( == )

(#

Presenter Name(s) Poster title

# **EVENT PLATFORM FAQS**

1. How do I get to the MOM 2020 event platform?

The event is hosted at MeetAnyway Platform and here is the event space link: https://meetanyway.com/events/molecular-origins-of-life-munich-2020/space

2. Which browser should I use?

Chrome is the best option and Firefox is 2nd. Edge and Internet Explorer are not supported.

3. My camera and microphone do not work? You need to allow your browser to access camera and microphone.

Chrome -> Settings -> Privacy and security -> Camera or Microphone -> Turn on 'Ask before accessing on or off' and allow MeetAnyway to access when you open the event platform

Firefox -> Options -> Privacy and security -> Camera or Microphone Setting -> Click off 'Block new requests asking to access your camera/microphone' and allow MeetAnyway to access when you open the event platform

- 4. I cannot connect to the platform. What is the problem? Please deactivate any Addons such as WebRTC Control, WebRTC Leak, etc.
- 5. How do I get login details?

You must have received an email with the login details from CRC 235 Emergence of Life management (emergenceoflife@ Imu.de). Contact them if you cannot locate the email.

- 6. How can I change my password? Please use the 'Forgot Password?' link at the Login page to reset your password!
- 7. How can I change my profile at the event platform?

Please click on the '...' next to your name then on 'Profile Settings'. You can upload a photo, change your name and time zone

- 8. How do I get to the talks? Please scroll to Auditorium Floor on the event platform and click to 'View Stage' and open the link with 'Zoom'.
- 9. Can I join the event directly from Zoom?

We strongly suggest the attendees to use the platform for ease of access to the posters and to 'Meet the Speaker' sessions. Technically it is possible to directly connect to the Zoom sessions but we will not be providing the info.

#### 10. Why do I have to change room for 'Meet the Speaker' sessions?

The lectures will be hosted as Zoom Webinar due to high number of attendees. The Webinar add on allows only text interaction with the speakers and the panelists.

The 'Meet the Speaker' sessions will be hosted as 'Zoom Meeting' so all the attendees can verbally and with image/video interact with the speaker.

#### 11. Do I have to be present at my poster room at all times?

We suggest you to be present at your poster room as much as possible when you are not attending the talks.

#### 12. Where do I upload my poster?

You will not upload your poster. Once you are in your chat room, click to share your screen to show your research to the others.

13. How do I reach to the person whose poster I would like to see?

The platform provides a search option to find the location of the person. Once you are at the same location then you can chat with the presenter directly to schedule a visit. At the end of the abstract book there is contact details of all the attendees who allowed us to share information. So you can also send an email to the poster presenter to schedule a visit.

#### 14. Is there a possibility to meet the other attendees other than poster sessions?

- Yes, the Networking Floor provides several tables to hang around and meet the others. We have also created some topic based tables if you would like to speak about a certain topic: # astrophysical origins
- # early earth
- # RNA/DNA world
- # emergence of metabolism
- # prebiotic chemistry
- # systems chemistry
- # synthetic biology
- 15. Is there a logical order/grouping for poster floors?

We have grouped the presenters from similar time zones, floor number increases from east to west. It is more likely that you can reach the poster presenters at the lower floors earlier in the day and the higher floors later in the day. Floors 4 to 10 are presenters from central Europe so compatible with meeting schedule.

### 16. How can I delete my account from the MeetAnyway Platform?

Technical Support Contact for MeetAnyway Event Platform Erich Lehmann erich@meetanyway.com 0176 209 566 86

Please send an email to erich@meetanyway.com asking removal of your account from the platform after the event!

# PARTICIPATING IN TALKS (ZOOM WEBINAR)

### A. Join the Webinar via URL

- Install Zoom Desktop Client: https://zoom.us/client/latest/ZoomInstaller.exe 1
- 2. Go to the event space (Chrome): https://meetanyway.com/events/molecular-origins-of-lifemunich-2020/space
- 3 Sign in & join the Stage on the Auditorium Floor
- 4 Run the file with Zoom application on your computer

### B. Interacting in the Webinar

In a Zoom Webinar only host and panelists can talk. You can interact with the host, co-host and panelists by raising your hand, typing in chat or typing in Q&A.

#### I. Raise Your Hand:

- 1 Click the Raise Hand button at any time to indicate to the host and panelists that you have a question.
- 2 To lower your hand, click Lower Hand button.

### II. Send Messages in Chat:

The feature is controlled by the host and might be limited during the event.

- 1. Click the Chat button to open the chat panel.
- Type your message in the Text box at the bottom of the panel. 2
- 3 Press Enter to send you message.

#### III. Ask Questions in Q&A:

- Click the Q&A button to open the window.
- 2. Type your question in the text field.
- 3 If you want your question to be anonymous, check the Send Anonymously checkbox.
- Click Send

### C. Leave the Webinar

- Click the X at the upper-right corner of the window to exit the webinar or
- Click the Leave Meeting button in the dialog box.

# PARTICIPATING IN ZOOM MEETINGS

### A. Join the Zoom meeting via URL

- 1. Install Zoom Link : https://zoom.us/client/latest/ZoomInstaller.exe
  - Join the video call of the room on MeetAnyway MOM2020 Platform
  - Run the file with Zoom application on your computer

Note: A menu bar containing participant tools appears at the bottom of the Zoom meeting. This menu bar will appear and disappear as you roll your mouse over the area.

### **B. Audio Controls**

2

3

Using audio in a Zoom meeting requires you to have access to a microphone. Please be aware the host can mute and unmute you at any time. Check the icons in the menu bar and the participants panel to determine your current audio setting.

> To unmute yourself and begin talking, click the Unmute button (microphone) in the bottom-left corner of the meeting window.

- 2. indicating that your audio is now off.
- 3. To test your computer microphone and speakers, click the up arrow to the right of the Microphone icon and select Audio settings.

Note: You can switch to a different audio input device using the Audio Options button.

#### C. Video Controls

- Click the Start Video button in the menu bar at the bottom to begin your video. 1.
- 2. Click the Stop Video button to stop sharing your video stream.
- 3
- icon and select Video Settings.

Note: When video is activated, display options are available in the upper right of the screen and in the upper right of each participant's window in both Speaker View and Gallery View. Note: If you choose not to turn on your webcam in the meeting and video has been activated, your video window will contain either your name, email address, or a photo, depending on how your profile is set up.

#### D. Send Messages in Chat

You can send a chat message to all participants in the meeting or privately to specific individuals. Note: Messages posted in chat prior to you joining the meeting are not visible to you in the chat panel.

- 1. Click the Chat button to open the chat panel.
- Type your message in the Text box at the bottom of the panel. 2.
- 3. Press Enter to send you message.

Note: You can send a private message to a single person by clicking the down arrow in the To: field and selecting the person's name from the list. The person's name will stay selected until you click the down arrow again and select Everyone.

### E. Nonverbal Feedback

Nonverbal feedback icons include a thumbs up & down, clap, away, etc. allowing you to let the host know what you are thinking without interrupting the meeting.

- 1. Click the Participants button in the menu bar to open the Participants panel.
- 2 Click the '...more' button to display the icons and click the icon.

Note: Only a single icon is visible at any time. After clicking an icon, clicking a new icon will overwrite the first icon. Note: Clap and thumbs up icons can be found also at the Reactions button in the menu bar.

#### F. Share Your Screen

Both hosts and participants can share their screen in Zoom. However, participants cannot share if the host is already sharing, or if the host has disabled this feature for participants.

- 1. Click the Share Screen button on the menu bar.
- 2. Selecting Desktop will allow you to share everything on your desktop.
- Click the Share Screen. 3
- 4. message, click the More button and select Chat from the list.
- 5 participants' attention to an area of your screen or use the text tool to type notes on the screen.
  - Click the X in the upper right of the annotation menu to close the annotation menu.
  - Click the More icon to view additional options.
  - Click Stop Share in the small menu bar to stop sharing your screen.

Note: When sharing your screen, the menu bar moves to the top of your screen and disappears until your oll your mouse over the area. Additional tools, such as Chat, Remote Control, and Audio options are located under the More button. To reposition the menu bar, click and drag it to another location on your computer.

#### G. Leave the Webinar

6.

7.

8.

- 1. Click the Leave Meeting option in the menu bar to exit the meeting or
- 2. Click the Leave Meeting button in the dialog box.

2

To mute yourself, click the Mute button (microphone). A red slash will appear over the microphone icon

To choose a different webcam or adjust your video settings, click the up arrow to the right of the Video

Select the desktop or application you would like to share or select whiteboard to share a whiteboard

If you receive a chat message while you are screen sharing, the More button will blink. To view the chat

Click the Annotate button to open the annotation menu. Use the draw tools (arrows, shapes) to direct

# TIME' BTH JULY . WEDNESDAY

10:45-11:00	Welcome & Info Session by Dieter Braun
SESSION I	Chair : Petra Schwille
11:00-11:25	Tetsuya Yomo East China Normal Uni.
	Construction and Evolution of a Synthetic Cell
11:25-11:50	Marileen Dogterom TU Delft
	Reconstituting Cytoskeletal Systems in Artificial Cells
11:50-12:00	Panel Discussion
SESSION II	Chair : William Orsi
12:00-12:25	Uwe Meierhenrich Université Côte d'Azur
	3D Presentation Rosetta-Philae - The Detection of Organic Molecules on the Surface of a Cometary Nucleus
12:25-12:50	Joachim Reitner Georg-August-Uni. of Göttingen
	Traces of Life in Very Old Rocks – What is Convincing and What Not?
12:50-13:00	Panel Discussion
12.50-13.00	Panel Discussion
13:00-13:45	Meet the Speakers <sup>2</sup> of Session I & II
13:45-14:45	Lunch Break
14:45-15:00	Info Session by Dieter Braun
14.40-10.00	into desision by Dicter Diddi
SESSION III	Chair : Paola Caselli
SESSION III	Chair : Paola Caselli
SESSION III	Chair : <b>Paola Caselli</b> <b>Thomas Henning</b> MPI for Astronomy <i>Chemistry in Protoplanetary Disks</i> <b>Ralph Pudritz</b> McMaster University
SESSION III 15:00-15:25	Chair : <b>Paola Caselli</b> <b>Thomas Henning</b> MPI for Astronomy <i>Chemistry in Protoplanetary Disks</i> <b>Ralph Pudritz</b> McMaster University <i>RNA Polymerization in Pre-Biotic Environments: Experiments</i>
SESSION III 15:00-15:25	Chair : <b>Paola Caselli</b> <b>Thomas Henning</b> MPI for Astronomy <i>Chemistry in Protoplanetary Disks</i> <b>Ralph Pudritz</b> McMaster University
SESSION III 15:00-15:25	Chair : <b>Paola Caselli</b> <b>Thomas Henning</b> MPI for Astronomy <i>Chemistry in Protoplanetary Disks</i> <b>Ralph Pudritz</b> McMaster University <i>RNA Polymerization in Pre-Biotic Environments: Experiments</i> with the Planet Simulator McMaster's Origins of Life
SESSION III 15:00-15:25 15:25-15:50	Chair : Paola Caselli Thomas Henning MPI for Astronomy Chemistry in Protoplanetary Disks Ralph Pudritz McMaster University RNA Polymerization in Pre-Biotic Environments: Experiments with the Planet Simulator McMaster's Origins of Life Laboratory
SESSION III 15:00-15:25 15:25-15:50 15:50-16:00	Chair : Paola Caselli Thomas Henning MPI for Astronomy Chemistry in Protoplanetary Disks Ralph Pudritz McMaster University RNA Polymerization in Pre-Biotic Environments: Experiments with the Planet Simulator McMaster's Origins of Life Laboratory Panel Discussion
SESSION III 15:00-15:25 15:25-15:50 15:50-16:00 SESSION IV	Chair : Paola Caselli Thomas Henning MPI for Astronomy Chemistry in Protoplanetary Disks Ralph Pudritz McMaster University RNA Polymerization in Pre-Biotic Environments: Experiments with the Planet Simulator McMaster's Origins of Life Laboratory Panel Discussion Chair : Philippe Schmitt-Kopplin John Eiler California Institute of Technology Reconstructing Prebiotic and Biotic Chemistry Using
SESSION III 15:00-15:25 15:25-15:50 15:50-16:00 SESSION IV	Chair : Paola Caselli Thomas Henning MPI for Astronomy Chemistry in Protoplanetary Disks Ralph Pudritz McMaster University RNA Polymerization in Pre-Biotic Environments: Experiments with the Planet Simulator McMaster's Origins of Life Laboratory Panel Discussion Chair : Philippe Schmitt-Kopplin John Eiler California Institute of Technology Reconstructing Prebiotic and Biotic Chemistry Using Molecular Isotopic Structures
SESSION III 15:00-15:25 15:25-15:50 15:50-16:00 SESSION IV	Chair : Paola Caselli Thomas Henning MPI for Astronomy Chemistry in Protoplanetary Disks Ralph Pudritz McMaster University RNA Polymerization in Pre-Biotic Environments: Experiments with the Planet Simulator McMaster's Origins of Life Laboratory Panel Discussion Chair : Philippe Schmitt-Kopplin John Eiler California Institute of Technology Reconstructing Prebiotic and Biotic Chemistry Using Molecular Isotopic Structures Elizabeth Bell UC Los Angeles
SESSION III 15:00-15:25 15:25-15:50 15:50-16:00 SESSION IV 16:00-16:25	Chair : Paola Caselli Thomas Henning MPI for Astronomy Chemistry in Protoplanetary Disks Ralph Pudritz McMaster University RNA Polymerization in Pre-Biotic Environments: Experiments with the Planet Simulator McMaster's Origins of Life Laboratory Panel Discussion Chair : Philippe Schmitt-Kopplin John Eiler California Institute of Technology Reconstructing Prebiotic and Biotic Chemistry Using Molecular Isotopic Structures
SESSION III 15:00-15:25 15:25-15:50 15:50-16:00 SESSION IV 16:00-16:25	Chair : Paola Caselli Thomas Henning MPI for Astronomy Chemistry in Protoplanetary Disks Ralph Pudritz McMaster University RNA Polymerization in Pre-Biotic Environments: Experiments with the Planet Simulator McMaster's Origins of Life Laboratory Panel Discussion Chair : Philippe Schmitt-Kopplin John Eiler California Institute of Technology Reconstructing Prebiotic and Biotic Chemistry Using Molecular Isotopic Structures Elizabeth Bell UC Los Angeles The Role of Zircon in the Search for Earth's Earliest
SESSION III     15:00-15:25     15:25-15:50     15:50-16:00     SESSION IV     16:00-16:25     16:25-16:50	Chair : Paola Caselli Thomas Henning MPI for Astronomy Chemistry in Protoplanetary Disks Ralph Pudritz McMaster University RNA Polymerization in Pre-Biotic Environments: Experiments with the Planet Simulator McMaster's Origins of Life Laboratory Panel Discussion Chair : Philippe Schmitt-Kopplin John Eiler California Institute of Technology Reconstructing Prebiotic and Biotic Chemistry Using Molecular Isotopic Structures Elizabeth Bell UC Los Angeles The Role of Zircon in the Search for Earth's Earliest Biosphere
SESSION III     15:00-15:25     15:25-15:50     15:50-16:00     SESSION IV     16:00-16:25     16:25-16:50     16:50-17:00	Chair : Paola Caselli Thomas Henning MPI for Astronomy Chemistry in Protoplanetary Disks Ralph Pudritz McMaster University RNA Polymerization in Pre-Biotic Environments: Experiments with the Planet Simulator McMaster's Origins of Life Laboratory Panel Discussion Chair : Philippe Schmitt-Kopplin John Eiler California Institute of Technology Reconstructing Prebiotic and Biotic Chemistry Using Molecular Isotopic Structures Elizabeth Bell UC Los Angeles The Role of Zircon in the Search for Earth's Earliest Biosphere Panel Discussion

# BTH JULY . THURSDAY

10:45-11:00	Welcome & Info Session by Dieter Braun	10:45-11:00	
SESSION V	Chair : Job Boekhoven	SESSION IX	
11:00-11:25	Jan van Esch TU Delft	11:00-11:25	
	Approaching Biological Complexity: Beyond Self-Assembly		
11:25-11:50	Stephen Mann University of Bristol		
	Coacervate Dynamics and the Origin of Life	11:25-11:50	
11:50-12:00	Panel Discussion		
SESSION VI	Chair : Andres Jäschke	11:50-12:00	
12:00-12:25	Longfei Wu MRC LMB		
	Harnessing Chemical Energy for Activation and Joining of Prebiotic Building Blocks	SESSION X	
12:25-12:50	Rafal Szabla University of Edinburgh	12:00-12:25	
	Shedding UV Light on the Common Origins of RNA and		
	DNA	12:25-12:50	
12:50-13:00	Panel Discussion		
13:00-13:45	Meet the Speakers <sup>2</sup> of Session V & VI	12:50-13:00	
13:45-14:45	Lunch Break	13:00-13:45	
CTIIN VIII		13:45-14:45	
14:45-15:00	Info Session by <b>Dieter Braun</b>	CHENDERD	11.25
session vii	Chair : Clemens Richert	14:45-15:00	
15:00-15:25	Daniel Duzdevich HHMI	SESSION XI	
	The Sequence Space of Non-enzymatic RNA Copying	15:00-15:25	
15:25-15:50	Nick Hud Georgia Institute of Technology		
	A Self-Assembly Approach to Uncovering Possible Ancestors of RNA	15:25-15:50	
15:50-16:00	Panel Discussion		
SESSION VIII	Chair : <b>Oliver Trapp</b>	15:50-16:00	
16:00-16:25	Lijun Zhou HHMI	SESSION XII	
	Assembly of a Functional Ribozyme from Short Oligomers	16:00-16:25	
	by Enhanced Non-enzymatic Ligation	10.00-10.25	
16:25-16:50	Donna Blackmond Scripps Research	16:25-16:50	
	Asymmetric Amplification in Peptide-Catalyzed Formation of C4 Sugars from Nearly Racemic Amino Acids		
16:50-17:00	Panel Discussion	16:50-17:00	
17:00-17:45	Meet the Speakers <sup>2</sup> of Session VII & VIII	17:00-17:45	
			<b>—</b>

1 UTC+02:00, CEST, CEDT, MEST

CONFERENCE SCHEDU

2 Each Speaker will be hosted in a separate chat room that is moderated!

3 Each speaker will have a puzzle and a beer for each day to start the informal evening session in parallel to Poster Session! 4 Posters can be visited at any time, please contact the presenter!

# TIME' **10 TH JULY**. FRIDAY

### Welcome & Info Session by Dieter Braun Chair : Don Dingwell Allen Nutman University of Wollongong How Long Ago was the Beginning? Looking for Life Signatures in ≥3.7 Billions-of-years-old Greenland Rocks William Orsi LMU Munich Quantifying the Effects of Abiotic H2 Production on Carbon Metabolism in Serpentinization Systems **Panel Discussion** Chair : Friedrich Simmel Sudha Rajamani Indian Ins. of Sci. Edu. Res. Prebiotic Selection Pressures Shape the Evolution of Protocells Christophe Danelon TU Delft Roadmap to Building a Cell **Panel Discussion** Meet the Speakers<sup>2</sup> of Session IX & X Lunch Break Info Session by Dieter Braun Chair : Hannes Mutschler Klara Hlouchova Charles University in Prague Searching for Early Proteins in Randomness Roy Black University of Washington Prebiotic Membranes Bind Protocell Building Blocks and Catalyze Formation of Biopolymers Panel Discussion Chair : Erwin Frey Christoph Weber MPI PKS

Selection via Phase Separation

Sergei Maslov Uni. Illinois Urbana-Champaign

Onset of Natural Selection in Populations of Autocatalytic Heteropolymers

**Panel Discussion** 

Meet the Speakers<sup>2</sup> of Session XI & XII

Puzzle & Beer of the Day<sup>3</sup> + Poster Session<sup>4</sup>

Date & Time		Speakers	Titles					
	11:00	TETSUYA YOMO	Construction and Evolution of a Synthetic Cell					
Ś	11:50	MARILEEN DOGTEROM	Reconstituting Cytoskeletal Systems in Artificial Cells					
=	12:00	UWE MEIERHENRICH	3D Presentation Rosetta-Philae - The Detection of Organic Molecules on the Surface of a Cometary Nucleus					
Ś	12:25	JOACHIM REITNER	Traces of Life in Very Old Rocks – What is Convincing and What Not?					
≡.	15:00	THOMAS HENNING	Chemistry in Protoplanetary Disks					
ഗ്	15:25	RALPH PUDRITZ	RNA Polymerization in Pre-Biotic Environments: Experiments with the Planet Simulator McMaster's Origins of Life Laboratory					
≥.	16:00	JOHN EILER	Reconstructing Prebiotic and Biotic Chemistry Using Molecular Isotopic Structures					
Ś	16:25	ELIZABETH BELL	The Role of Zircon in the Search for Earth's Earliest Biosphere					
S. V	11:00	JAN VAN ESCH	Approaching Biological Complexity: Beyond Self-Assembly					
S	11:25	STEPHEN MANN	Coacervate Dynamics and the Origin of Life					
S. VI	12:00	LONGFEI WU	Harnessing Chemical Energy for Activation and Joining of Prebiotic Building Blocks					
S	12:25	RAFAL SZABLA	Shedding UV Light on the Common Origins of RNA and DNA					
IN.	15:00	DANIEL DUZDEVICH	The Sequence Space of Non-enzymatic RNA Copying					
ŝ	15:25	NICHOLAS HUD	A Self-Assembly Approach to Uncovering Possible Ancestors of RNA					
III	16:00	LIJUN ZHOU	Assembly of a Functional Ribozyme from Short Oligomers by Enhanced Non-enzymatic Ligation					
Ś	16:25	DONNA BLACKMOND	Asymmetric Amplification in Peptide-Catalyzed Formation of C4 Sugars from Nearly Racemic Amino Acids					
×	11:00	ALLEN NUTMAN	How Long Ago was the Beginning? Looking for Life Signatures in ≥3.7 Billions-of-Years-Old Greenland Rocks ————————————					
ŝ	11:25	WILLIAM ORSI	Quantifying the Effects of Abiotic H2 Production on Carbon Metabolism in Serpentinization Systems					
s. x	12:00	SUDHA RAJAMANI	Why Composite Prebiotic Membranes are CooL?					
	12:25	CHRISTOPHE DANELON	Roadmap to Building a Cell					
: XI	15:00	KLARA HLOUCHOVA	Searching for Early Proteins in Randomness					
S	15:25	ROY BLACK	Prebiotic Membranes Bind Protocell Building Blocks and Catalyze Formation of Biopolymers					
IIX	16:00	CHRISTOPH WEBER	Selection via Phase Separation					
ŝ	16:25	SERGEI MASLOV	Onset of Natural Selection in Populations of Autocatalytic Heteropolymers					

SPEAKERS INDEX

1

### **SESSION I**

### SESSION I

### CONSTRUCTION AND EVOLUTION OF A SYNTHETIC CELL



Tetsuya Yomo Laboratory of Biology and Information Science, ECNU

The ability to evolve is a key characteristic that distinguishes living things from non-living chemical compounds. The construction of an evolvable cell-like system entirely from non-living molecules has been a major challenge. Here we constructed an evolvable synthetic cell from an assembly of biochemical molecules. The biochemical molecules assembled into micro-scaled lipid vesicles or water-droplets in oil are the artificial genomic RNA, all the molecules required for protein synthesis and RNA replication. In the micro-compartments, the genetic information on the genomic RNA is translated into RNA replicase, which in turn replicates the original genomic RNA.

After inner replication reaction of RNA, exhausting the nutrient molecules such as nucleic acids and so on, the liposomes containing the replicated RNA were subjected to the fusion with the other liposomes containing the nutrients and the division for proliferation in order to get ready for the next RNA replication reaction<sup>1</sup>.

Using the translation-coupled RNA replication system, we performed a long-term (600-generation) replication experiment, in which mutations were spontaneously introduced by the translated replicase into its genetic information, and highly replicable mutant RNAs dominated the population according to Darwinian principles. At the beginning of the evolution, the replicated RNA accumulated to form the double strand, a dead-end product for the translation while a small parasitic RNA evolved by a deletion mutation on the original RNA genome to dominate by stealing the replicase translated from the original RNA genome. However, during the experimental evolution, the genomic RNA gradually reinforced its interaction with the translated replicase, thereby acquiring competitiveness against the parasitic RNA. This study provides the first experimental evidence that a simple assembly of biomolecules in a cell-like compartment can autonomously develop their genetic code through Darwinian evolution<sup>2</sup>.

#### References

1Tsuji G., Fujii S., Sunami T. and Yomo T. (2016) Proceeding of National Academy of Science USA 113(3) 590-595 2 Ichihashi N, Usui K, Kazuta Y, Sunami T, Matsuura T, Yomo T (2013) Nature Communication. 4(2494):1-7 doi:10.1038/ncomms3494.

### RECONSTITUTING CYTOSKELETAL SYSTEMS IN ARTIFICIAL CELLS

Marileen Dogterom

Kavli Institute of Nanoscience at Delft University of Technology

In my group we are interested in understanding how dynamic and force-generating properties of the cytoskeleton contribute to the spatial organization of cells. I will highlight recent advances (and challenges) in our efforts to reconstitute minimal, functional cytoskeletal systems in artificial confinement. An example is the reconstitution of basic mitotic spindles in microfluidic droplets. These efforts fit in a long-term ambition to build, in collaboration with others, a minimal synthetic cell from scratch.

### 🕞 Wednesday, 08 July, 11:25 CEST



### **SESSION II**

### **3D**\* PRESENTATION ROSETTA-PHILAE -THE DETECTION OF ORGANIC MOLECULES ON THE SURFACE OF A COMETARY NUCLEUS



Uwe Meierhenrich Université Côte d'Azur, Nice, France

ESA's Rosetta mission had made spectators from all over the world dream: On Wednesday, 12 November 2014, the Rosetta mission tried to pose the little robot Philae on the nucleus of comet 67P/Churyumov-Gerasimenko. The Rosetta Space Probe aimed to collect information about the composition of the comet nucleus during its spectacular approach to the sun<sup>1</sup>. Rosetta is the first probe to place itself in orbit around the comet and to place a lander on the surface of a cometary nucleus. The Rosetta probe carried 11 scientific instruments and a Philae lander which itself comprises 10 additional instruments. After 10 years of travel, the separation of the Philae lander from the Rosetta orbiter was carried out on November 12, 2014. The cometary sampling and composition (COSAC) instrument, a device onboard Philae, which we developed in an international partnership lead by the Max Planck Institute for Solar System Research, is a gas chromatograph using eight stationary phases coupled with a mass spectrometer time of flight type. 25 minutes after Philae's landing and bouncing on the cometary nucleus, COSAC successfully performed the first chemical analysis of cometary surface material that cannot be analyzed from the Earth. 16 organic molecules were identified in the cometary sample by using COSAC's MS-only mode<sup>2</sup>. After two additional bouncing events Philae finally landed on the cometary surface and operated for 60 h. During this time the COSAC instrument recorded 420 mass spectra in the enantioselective GC-MS mode. The identification of organic species in these mass spectra remains difficult because of the unexpected 'vertical' landing of Philae and the unexpected low amount of sample that was filled into the oven of COSAC's sample injector system. The first results of the analysis of the cometary nucleus surface by the COSAC instrument will be presented. These in situ cometary results will be interpreted in relation to laboratory experiments that allow for the simulation of cometary ices by condensing volatile molecules such as H<sub>2</sub>O, NH<sub>2</sub>, CO, CO<sub>2</sub>, and CH<sub>2</sub>OH in an ultrahigh vacuum from the gas phase onto a cooled surface of T = 12 K. The room temperature residues of the cometary ice analogues were shown to contain amino acids<sup>3</sup>, aldehydes<sup>4</sup> and ribose<sup>5</sup> as produced in form of simulated cometary ices in the laboratory<sup>6</sup>. The laboratory simulation experiments thereby confirm data on the chemical composition obtained by the Rosetta-Philae cometary probe.

#### References

1 Meierhenrich: Comets and their Origin, Wiley-VCH, Weinheim 2015. 2 Goesmann, Meierhenrich et al.: Science 349, 2015, 497. 3 Munoz Caro, Meierhenrich et al.: Nature 412, 2004, 403-406. 4 De Marcellus, Meierhenrich et al.: Proc. Natl. Acad. Sci. USA 112, 2015, 965–970. 5 Meinert, Meierhenrich et al.: Science 352, 2016, 208–212. 6 Myrgorodaka, Meierhenrich et al.: Angew. Chem. Int. Ed. 54, 2015, 1402–1412

\* Use of 3D glasses is recommended!

### TRACES OF LIFE IN VERY OLD ROCKS - WHAT IS CONVINCING AND WHAT NOT?

Joachim Reitner<sup>1,2</sup>, J.-P. Duda<sup>1,2</sup>, M.Van Zuilen<sup>3</sup>, M.Reinhardt<sup>4</sup>, V.Thiel<sup>1</sup>, H.Mißbach<sup>1</sup> & M.Hoppert<sup>1</sup> <sup>1</sup> Georg-August-Universität Göttingen, Göttingen, Germany <sup>2</sup> Göttingen Academy of Sciences and Humanities, Göttingen, Germany <sup>3</sup> Institut de Physique du Globe de Paris, France

<sup>4</sup> Linnaeus University, Kalmar, Sweden

SESSION II

Life emerged >3.9 billion years (Gy) ago. However, it is very difficult to track this development in the geological rock record because most live traces have been wiped out by later metamorphic processes<sup>1,2</sup>. Organic carbon (Corg) preserved in late Hadean-early Archean metamorphic rocks might evidences the presence of life<sup>1</sup>. However, the Corg is not necessarily syngenetic with the formation of the host rock and could also be derived from abiotic sources (e.g., Fischer-Tropschtype-synthesis, meteoric delivery)1-4. The discrimination of biological and abiotic contributions is challenging because the preserved Corg is recalcitrant and usually does not contain specific organic molecules. Moreover, δ<sup>13</sup>C signatures of biological an abiotic Corg can interfere. Therefore, additional lines of evidence are required, including field observations, petrographic characteristics (e.g., microbial structures, mineral associations) and geochemistry (e.g., stable isotope signatures of inorganic metabolic products such as carbonates)<sup>3,5,6</sup>. In this talk, I will discuss potential pitfalls in the search for earliest life (e.g., 3.7 Gy old so-called "stromatolites" of Greenland)<sup>1</sup> and then demonstrate how we can verify and understand fingerprints of early live (e.g., in 3.5-3.4 Gy old primary fluid inclusions and microbialites)<sup>3,5</sup>. Our investigations help to validate putative traces of life in very old rocks and to develop a solid understanding of how life emerged on our planet.

#### References

1 Reitner et al. (2020) Verlag Kurt Roessler Bornheim, 35–45 (ISBN: 978-3-935369-48-0) 2 Duda & Reitner (2016) Int. J. Astrobiol. 15, 161–163 (10.1017/S1473550416000276) 3 Duda et al. (2018) Biogeosci. 15(5), 1535–1548 (10.5194/bg-15-1535-2018) 4 Mißbach et al. (2018) Org. Geochem. 119, 110-121 (10.1016/j.orggeochem.2018.02.012) 5 Duda et al. (2016) PloS One 11(1), e0147629 (10.1371/journal.pone.0147629) 6 Rincón-Tomás et al. (2016) Int. J. Astrobiol. 15(3), 219-229. (10.1017/S1473550416000264)

wwe.meierhenrich@unice.fr

### Wednesday, 08 July, 12:25 CEST



### **SESSION III**

### SESSION III

### CHEMISTRY IN PROTOPLANETARY DISKS

#### Thomas Henning

Max Planck Institute for Astronomy, Heidelberg, Germany

Protoplanetary disks are circumstellar structures around young stars resembling the solar nebula at the time our planetary system formed. The disks are composed of gas - mostly molecular hydrogen - and a population of solid particles which are important in regulating the radiation level and thermal structure of these objects. Temperatures in the disks range from several thousand Kelvin close to the star to very low temperature of several Kelvin in the outer disk mid-plane. The disk surfaces are exposed to cosmic rays and stellar UV and Xray irradiation. The various regions of protoplanetary disks provide a very diverse environment for chemical processes with surface chemistry and molecular freeze-out playing an important role.

The talk will summarize the chemical properties of protoplanetary disks and will highlight what we know observationally about the molecular content of these objects. The observational basis for what we know about molecules in disks is provided by infrared spectroscopy from space and ground and (sub)millimeter molecular spectroscopy by facilities such as ALMA. The talk will also discuss potential routes to molecular complexity in disks which is connected to the formation of pre-biotic molecules.



### RNA POLYMERIZATION IN PRE-BIOTIC ENVIRONMENTS: EXPERIMENTS WITH THE PLANET SIMULATOR MCMASTER'S ORIGINS OF LIFE LABORATORY

Ralph E. Pudritz, Maikel Rheinstadter, Alix Dujardin, Renée-Claude Bider, and Sebastian Himbert McMaster University, Dept of Physics and Astronomy, and Origins Institute

How did life originate on the Earth, and is it possible that it emerged on other Earth-like planets? It has long been posited that RNA could have been the first genetic materials in protocells<sup>1</sup>. Recent experimental advances have indicated that RNA polymers may grow by means of wet-dry cycles in prebiotic environments<sup>2</sup>. With the intent of performing experiments on RNA polymerization in a wide variety of planetary, pre-biotic conditions, we have designed a planet simulator that is now functioning in McMaster's Origins of Life Laboratory. The simulator allows excellent control and cycling of humidity, temperature, stellar 1rradiation (from IR to UV), gas composition, and pressure (one atmosphere and lower). We describe a number of experiments which act as a first "survey" of the large pre-biotic parameter space. in which various mixtures of nucleotides, salts, lipids, and irradiation conditions are explored for their ability to sustain RNA polymerization. We present our first results that suggest new insights into the role of wet/dry cycle design, salts (especially ammonium chloride) and UV irradiation- on the length of RNA polymers that may form.

References: 1 Gilbert, W. 1986, Nature, 319:618, 2 Da Silva, L., Maurel, M.-C., Maurel, Deamer, D. 2015, J Mol Evol, 80, 86

henning@mpia.de

### Wednesday, 08 July, 15:25 CEST



### **SESSION IV**

John M. Eiler

### **SESSION IV**

### THE ROLE OF ZIRCON IN THE SEARCH FOR EARTH'S EARLIEST BIOSPHERE

#### Elizabeth A.Bell

Dept. of Earth, Planetary, and Space Sciences, UCLA, 595 Charles Young Dr. E, Los Angeles, CA 90095, United States of America

In recent decades, evidence has increasingly pointed to a Hadean (>4 billion years ago) Earth with relatively clement conditions, instead of the hellish and inhospitable environment of its namesake. Although crust older than ca. 4 billion years has not yet been identified, out-of-context crystals of zircon in later sediments have provided evidence for liquid water, a sedimentary cycle, and at least some long-lived, evolved crust within a few hundred million years of planetary formation. This suggests the possibility of planetary habitability far earlier than the oldest yet-confirmed microfossils. Carbonaceous materials from both early Archean metasediments in Greenland and from a Hadean detrital zircon from Western Australia are isotopically lighter than prevailing sources of inorganic carbon in the geologic record and may potentially preserve the signature of a Hadean-Eoarchean biosphere. The search for more such carbonaceous material is ongoing, and its rarity suggests the need for developing alternative lines of evidence for a Hadean biosphere. Available Hadean zircons are dominantly igneous in nature, and the presence of a biosphere is most evident in modern magmas through the assimilation of organic carbon-laden sediments to produce reduced, peraluminous "S-type" granites. Zircon-based proxies for magma composition use certain trace elements and the composition of mineral inclusions in zircon to determine the aluminosity and redox state of their host magmas. Several lines of evidence point to the presence of simultaneously peraluminous and reduced magmas in the Hadean. Further work on the timing of these signals in the zircon record may help to unravel the history of the terrestrial biosphere and may be a helpful companion for interpreting the carbon isotopic record. Further developing geochemical proxies in zircon will also allow us to better understand the composition of the Hadean igneous crust, which would have been the main source of inorganic nutrients to the early environment

### RECONSTRUCTING PREBIOTIC AND BIOTIC CHEMISTRY USING MOLECULAR ISOTOPIC STRUCTURES



Division of Geological and Planetary Sciences, California Institute of Technology, Pasadena, CA, USA

Terrestrial life must have emerged from environments containing prebiotic organic compounds. However, our ability to reconstruct the first organic chemical systems that made these compounds, and to understand their transformation to the earliest life is hampered by several problems: We don't clearly recognize terrestrial organic matter in the rock record that pre-dates life; metamorphism has obscured much of the chemical complexity in our records of early life; attempts to study abiotic organic chemistry in the modern Earth are often contaminated due to the near ubiquity and fecundity of life; and as yet we have only a loose understanding of the chemistry that formed organics in our few samples of extra-terrestrial materials. For these reasons, much of our understanding of prebiotic chemistry rests on laboratory experiments that let us explore possible chemical synthesis pathways and environments and rule out implausible scenarios. Nevertheless, it remains imperative that we find ways to concretely link the chemistry that lab experiments prove is possible to the historical prebiotic chemistry that actually occurred, on the Earth and elsewhere.

Our presentation will explore the use of molecular isotopic structure - differences in isotopic content between non-equivalent atomic sites and proportions of multiply substituted isotopic forms of molecules - as a new means of distinguishing biotic from abiotic organics, of reconstructing relationships between substrates and products in organic reaction networks, and of testing hypotheses regarding the mechanisms and conditions of prebiotic chemistry. This approach has been enabled by a recent revolution in the capabilities of several technologies for analysis of isotopic structures, particularly mass spectrometric methods that are suitable for trace amounts of complex organic molecules. Also key are new theoretical studies of the chemical physics controlling intramolecular partitioning of isotopes. We will discuss these new tools and the ways in which they inform understanding of molecular formation, and we will present the newest findings of the application of this emerging field to the understanding of the synthesis of amino acids, nucleobases and other soluble organics from primitive carbonaceous meteorites. Finally, we will discuss the projected near-future application of these tools to the study of abiogenesis in terrestrial natural settings (e.g., serpentinites) and the study of ancient putative fossil biomass.

### Wednesday, 08 July, 16:25 CEST



### 🕞 Thursday, 09 July, 11:00 CEST

### APPROACHING BIOLOGICAL COMPLEXITY: BEYOND **SELF-ASSEMBLY**



#### Jan van Esch, Rienk Eelkema, Qian Liu, Eduardo Mendes

SESSION V

Department of Chemical Engineering, Delft University of Technology, Delft, The Netherlands

It remains a huge scientific challenge to understand and mimic the utilisation of chemical energy in biological systems to achieve the highly adaptable organisation and sophisticated functions like active transport, motility, selfrepair, replication, and adaptability. The development of biomimetic systems with similar energy consuming organisation and functions requires a radical departure from equilibrium self-assembly approaches, towards out-ofequilibrium systems driven by the continuous input of energy.

In our research we focus on the development of active materials driven by chemicals fuels. First, I will discuss how active materials can result from the transient self-assembly of synthetic molecules, driven by the consumption of a chemical fuel. In these materials, reaction rates and fuel levels, instead of equilibrium composition, determine properties such as lifetime, stiffness, and self- regeneration capability.<sup>1-3</sup> Then, I will discuss our recent steps to achieve temporal and spatial over fuel-driven self-assembly by the development of a chemical reaction network that allow for feedback control. Such systems will form the basis for self-organising systems and for design and construction of energy-consuming dynamic devices and materials.

References

1 J. Boekhoven, A.M. Brizard, K.N. Kowlgi, G.J. Koper, R. Eelkema, J.H. van Esch, Angew. Chem. Int. Ed. 49 DOI: 10.1002/anie.201001511 (2010)

2 J. Boekhoven, W. Hendriksen, G. Koper, R. Eelkema, J.H. van Esch, Science 349, 1075 (2015)

3 B. G. P. van Ravensteijn, W. E. Hendriksen, R. Eelkema, J. H. van Esch, W. K. Kegel, J Am Chem Soc 2017, 139, 9763-9766

4 Y. Wang, R. M. de Kruijff, M. Lovrak, X. Guo, R. Eelkema, J. H. van Esch, Angew. Chem. Int. Ed. 2019, 58, 3800.

Kevwords:

Self-Assembly, Supramolecular Chemistry, Active Materials

### **SESSION V**

### COACERVATE DYNAMICS AND THE ORIGIN OF LIFE

#### Stephen Mann

Max Planck Bristol Centre for Minimal Biology and Centre for Protolife Research, School of Chemistry, University of Bristol Bristol BS8 1TS UK

Many years ago Oparin proposed that liquid-liquid phase separated micro-droplets in the form of complex coacervates were a plausible model for the origin of life1. Although these ideas fell out of favour for many decades, the recent realization that membrane-free molecularly crowded coacervate droplets (condensates) play important and diverse functional roles in extant biology has refocused attention on phase separation as a possible mechanism underlying protobiological organization<sup>2</sup>. Recent studies in my laboratory have shown that coacervate micro-droplets exhibit high levels of molecular enrichment and biochemical activity including in vitro gene expression<sup>3</sup> and photosynthesis<sup>4</sup>, can be transformed into membrane-coated protocells using lipid<sup>5</sup> or inorganic building blocks<sup>6</sup>, and utilized as microscale agents for chemical communication and signalling in reaction-diffusion gradients<sup>7,8</sup>. Coacervate-based protocells are also capable of morphological reorganization in gradients of artificial morphogens<sup>9</sup>, reversible structural transformation into lipid vesicles<sup>10</sup>, light-induced assembly and disassembly for oligonucleotide trafficking<sup>11</sup>, primitive phagocytosis<sup>10</sup> and predator-prey behaviour<sup>12,13</sup>. In this talk I will use the above studies to discuss the dynamical behaviour of coacervate protocell populations and speculate on the possible relevance of these properties in origin of life scenarios.

References

1 Oparin A I. The origin of life, Macmillan, New York 1938.

2 Koga, S., et al., Peptide-nucleotide microdroplets as a step towards a membrane-free protocell model. Nature Chem. 3, 720 (2011). 3 Tang T-Y D., et al., In vitro gene expression within membrane-free coacervate protocells. Chem Comm. 51, 11429 (2015). 4 Kumar P., et al., Chloroplast-containing coacervate micro-droplets as a step towards photosynthetically active membrane-free protocells. Chem. Comm. 54, 3594 (2018).

5 Tang T-Y D, et al., Fatty acid membrane assembly on coacervate micro-droplets as a step towards a hybrid protocell model. Nature Chem. 6, 527 (2014).

6 Gobbo P., et al., Catalytic processing in ruthenium-based polyoxometalate coacervate protocells. Nature Commun. 11, 41 (2020). 7 Tian L., et al., Non-equilibrium spatiotemporal sensing within acoustically patterned two-dimensional protocell arrays. ACS Central Sci. 4, 1551 (2018).

8 Liu J., et al., Hydroael-immobilized coacervate droplets as modular micro-reactor assemblies, Angew, Chem, Int. Ed. 59, 6853 (2020) 9 Tian L., et al., Artificial morphogen-mediated differentiation in synthetic protocell communities. Nature Commun. 10, 3321 (2019). 10 Martin N., et al., Antagonistic chemical coupling in self-reconfigurable host-quest protocells, Nature Commun, 9, 3652 (2018). 11 Martin N., et al., Photo-switchable phase separation and oligonucleotide trafficking in DNA coacervate micro-droplets. Angew. Chem. Int. Ed. 58, 14594 (2019).

12 Qiao Y., et al., Response-retaliation behaviour in synthetic protocell communities. Angew. Chem. Int. Ed. 58, 17758 (2019). 13 Qiao Y., et al. Predatory behaviour in synthetic protocell communities. Nature Chem. 9, 110 (2017).

### Thursday, 09 July, 11:25 CEST



### SESSION VI

### SESSION VI

### SHEDDING UV LIGHT ON THE COMMON ORIGINS OF RNA AND DNA

#### Rafal Szabla

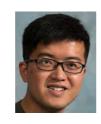
East CHEM, School of Chemistry, University of Edinburgh, Edinburgh, UK

The RNA World hypothesis assumes that ribonucleic acid was the first informational polymer on Earth that was responsible for storing genetic information as well as performing enzymatic activity. However, despite numerous efforts, prebiotic syntheses of RNA nucleotides either suffered from missing elements (e.g. lack of prebiotic sources of pure ribose)1 or given chemical selectivity, were successful for only two out of four of the canonical building blocks<sup>23</sup>. However, recent results suggest that all key components of genetic alphabet could have been delivered prebiotically on Earth as a mixture of RNA pyrimidine and DNA purine nucleosides<sup>4</sup>. In this reaction sequence, UV light offers remarkable selectivity by destroying biologically irrelevant stereoisomers and driving the key chemical transformations. In this talk, I will demonstrate the key aspects of these photochemically driven reactions. I will also show how the oligomers of these building blocks could have protected themselves from the effects of photodamage formation in UV-rich prebiotic environments⁵.

#### References:

1 Becker, S. et al. Science 366, 76-82 (2019). 2 Powner, M. W., Gerland, B.& Sutherland, J. D. Nature 459, 239-242 (2009). 3 Xu, J. et al. Nat. Chem. 9, 303–309 (2017). 4 Xu, J. et al. Nature 582, 60-66 (2020). 5 Szabla, R., Kruse, H., Stadlbauer, P., Šponer, J. & Sobolewski, A. L.Chem. Sci. 9, 3131–3140 (2018)

### HARNESSING CHEMICAL ENERGY FOR ACTIVATION AND JOINING OF PREBIOTIC BUILDING BLOCKS



#### Longfei Wu

MRC Laboratory of Molecular Biology, UK

Life is an out of equilibrium system sustained by a continuous supply of energy. In extant biology, the generation of the primary energy currency, adenosine 5'-triphosphate (ATP), and its use in the synthesis of biomolecules require sophisticated enzymes. Before the emergence of such enzymes, alternative energy sources, perhaps assisted by simple catalysts, must have mediated the activation of carboxylates and phosphates for condensation reactions. I will talk about our recent results that the chemical energy inherent to isonitriles can be harnessed to activate nucleoside phosphates and carboxylic acids through catalysis by acid and 4,5-dicyanoimidazole under mild conditions in aqueous solution. Simultaneous activation of carboxylates and phosphates provides multiple pathways for the generation of reactive intermediates, including mixed carboxylic acid-phosphoric acid anhydrides, for the synthesis of peptidyl-RNAs, peptides, RNA oligomers and primordial phospholipids. Our results indicate that, prior to ATP, the activation and joining of prebiotic building blocks in aqueous solution from a common pool could have been driven by an unified chemical energy.

### Thursday, 09 July, 12:25 CEST



### **SESSION VII**

### **SESSION VII**

### A SELF-ASSEMBLY APPROACH TO UNCOVERING POSSIBLE ANCESTORS OF RNA

#### Nicholas V. Hud

School of Chemistry and Biochemistry, Georgia Institute of Technology, USA

The RNA World hypothesis, which posits that RNA existed before the advent of DNA and proteins, remains a popular and influential hypothesis. However, despite substantial progress in all areas of origins of life research, a robust and plausible prebiotic synthesis for RNA polymers has remained elusive. Persistent challenges with finding such a synthesis include nucleobase selection (from what was likely a complex mixture of molecules on the prebiotic Earth) and nucleotide polymerization (particularly the coupling of mononucleotides without a pre-existing template or chemical activation). These two challenges might share the same solution. We are investigating the possibility that RNA was preceded by a polymer that would have assembled more easily than RNA. This ancestral genetic polymer, or proto-RNA, may have been comprised of different nucleobases and, perhaps, a different backbone. In support of this hypothesis, experiments in our laboratory have revealed that a set of alternative nucleobases, or putative proto-nucleobases, can self-assemble in water and readily form plausible proto-nucleosides with ribose in good yields, two properties not observed with the nucleobases of extant RNA. Moreover, supramolecular assemblies formed by these putative proto-nucleobases exhibit an extraordinarily strong propensity to spontaneously adopt homochiral helical structures, even when not modified by a chiral substituent (e.g., without an attached sugar). Recent x-ray diffraction studies of these assemblies now provide structural details that we plan to use as constraints for determining which of the many proposed proto-RNA backbones would have been compatible with these protonucleobases.

### THE SEQUENCE SPACE OF NONENZYMATIC RNA COPYING



#### **Daniel Duzdevich** Howard Hughes Medical Institute, Department of Molecular Biology, Center for Computational and Integrative

Biology, Massachusetts General Hospital, Boston, MA 02114

An important model system for studying RNA-based pre-biology is nonenzymatic templatedirected primer extension, a posited component of RNA replication. Recent advances are Prompting challenging questions about how environmental contexts and heterogeneity of reaction components affect the sequence space and fidelity of nonenzymatic primer extension. Addressing these questions will require new and versatile methods that can routinely provide detailed information about how this reaction accesses template and product sequences. I will introduce a new deep-sequencing tool for studying primer extension that uses a custom RNA construct, protocol, and analysis pipeline. I will briefly describe how the method was vetted, and its strengths and weaknesses. I will also discuss recent results that relate the established mechanism of primer extension to the patterns observed in the sequencing data, with a view to future experiments.

### Thursday, 09 July, 15:25 CEST



### Thursday, 09 July, 16:00 CEST

### ASSEMBLY OF A FUNCTIONAL RIBOZYME FROM SHORT OLIGOMERS BY ENHANCED NON-ENZYMATIC LIGATION



#### Lijun Zhou

Department of Molecular Biology, Howard Hughes Medical Institute, Massachusetts General Hospital, Boston, United States

The non-enzymatic replication of the primordial genetic material is thought to have enabled the evolution of the first ribozymes, leading to early forms of RNA-based life. However, the replication of oligonucleotides long enough to encode catalytic functions is problematic in many aspects including the low efficiency of template copying. As an alternative to template-directed polymerization of mononucleotides, the template-directed ligation of oligonucleotides could potentially help to assemble long RNAs from shorter oligonucleotides, which would be easier to replicate. However, the reported rate of non-enzymatic RNA ligation is extremely slow. Here we show that the rate of ligation can be greatly enhanced by employing a 3'-amino group at the 3'-end of each oligonucleotide, in combination with an N-alkyl imidazole organocatalyst. These modifications enable the rapid copying of long RNA templates by the multi-step ligation of tetranucleotide building blocks, as well as the assembly of long oligonucleotides using short splint oligonucleotides. We also demonstrate the formation of long oligonucleotides inside model prebiotic vesicles, suggesting a potential route to the assembly of artificial systems capable of evolution. Further, we show that a functional ligase ribozyme can be assembled in this manner. Three approaches of splint templates design enable efficient ligation and avoid the strong strand inhibition afterwards. We suggest that the genomes of primitive protocells may have consisted of relatively easily replicated oligonucleotides as short as 8 to 12 nucleotides in length.

### SESSION VIII

### **ASYMMETRIC AMPLIFICATION IN PEPTIDE-**CATALYZED FORMATION OF CH SUGARS FROM NEARLY RACEMIC AMINO ACIDS

Alexander X. Jones, Lingshan Wen, Luca Legnani, Jason Chen, and Donna G. Blackmond Department of Chemistry Scripps Research, La Jolla CA 92037 USA

Peptides formed from oligomerization of mixtures of amino acids under prebiotically plausible synthetic conditions are screened to identify catalysts for the formation of sugars from glycolaldehyde in buffered aqueous solution. Initial studies of libraries constructed using mixtures of enantiopure amino acids identified a number of peptides capable of inducing enantioenrichment in C4 sugars. Further studies demonstrated that enantioenriched erythrose could be synthesized in a one-pot sequential process starting from nearly racemic amino acids. Several selection levels are at play, combining physical phase behavior via eutectic partitioning with stochastic amplification in peptide formation and asymmetric catalysis of this simplified formose reaction.

### Thursday, 09 July, 16:25 CEST



### Friday, 10 July, 11:00 CEST

### HOW LONG AGO WAS THE BEGINNING? LOOKING FOR LIFE SIGNATURES IN >3.7 BILLIONS-OF-YEARS-OLD GREENLAND ROCKS



#### Allen Nutman

University of Wollongong, Wollongong, NSW, Australia

Because of continual plate tectonic activity, Earth's >3.6 billions-of-years-old (Ga) geological record only survives in about a millionth of the present crust. Most of this consists of metamorphosed granitic rocks, which will be devoid of life signatures. In contrast to the majority of Eoarchean (4.0-3.6 Ga) terranes, the 3.8-3.7 Ga Isua supracrustal (volcanic and sedimentary rock) belt in Greenland contains tectonic slices with maximum metamorphic temperatures below 550°C, as well as rare domains where deformation is low, resulting in the preservation of Eoarchean sedimentary structures. These domains form less than a trillionth of the present geological record. These are the only geological resource to cross-check the molecular clock modelling that places the emergence of life by the Eoarchean.

Since the 1970s, bulk graphite analysis of Isua metamorphosed sedimentary rocks reveals negative  $\delta^{13}C_{verse}$ values, and debate surrounds whether this is a biogenic, or by the reaction 6FeCO<sub>2</sub><->2Fe<sub>2</sub>O<sub>2</sub>+5CO<sub>2</sub>+C, a metamorphic signature. To avoid possible metamorphic graphite, subsequent reduced carbon studies have sought silicate sedimentary rocks devoid of carbonate, and graphite from such sedimentary rocks yielded negative  $\delta^{13}C_{vene}$  values (Rosing, 1999). This graphite has nanoscale morphologies consistent with pyrolyzation of structurally heterogeneous organic compounds during metamorphism. Iron isotopic signatures in Isua banded iron formations show isotope fractionations compatible with both biogenic and abiogenic processes (Dauphas et al., 2004). However, for many Isua carbonates, biogenic fractionation is the preferred interpretation (Craddock and Dauphas, 2011). In a departure from isotopic approaches, it is observed that units of Isua massive dolomite rocks (CaMg2CO<sub>3</sub>) have seawater-like rare earth element + yttrium signatures, indicating a sedimentary origin. Given that low temperature dolomite forms by microbial mediation, this is proposed as simple, robust evidence for early life.

Physical rather than chemical life evidence is also sought. In the 1970s, globular structures in metamorphosed silica-rich sedimentary rocks were interpreted as relict microfossils, but subsequent investigations have reinterpreted these as younger abiogenic structures. In a unique low deformation domain discovered in the 2010s, ~3.7 Ga dolomitic rocks preserve shallow water sedimentary structures and stromatolites (bio-structures), with the latter restricted to only about 2m<sup>2</sup> of outcrop.



Five decades of geological investigations have resulted in increasing and diverse lines of evidence for life on Earth by 3.7 Ga and probably by 3.85 Ga. Therefore, future research can leave the 'yes' or 'no' debate. Instead, via geologically-suitable samples, studies can focus on the early diversity of the ecological niches that life occupied and via an increasing array of stable isotope tools, its metabolic pathways. The oldest surviving sedimentary rocks on Earth are ~3.9 Ga, by which time life had probably emerged. Therefore, Mars, which has been geologically 'dead' for billions of years, might provide a better window on life's emergence.

**SESSION IX** 

### QUANTIFYING THE EFFECTS OF ABIOTIC N<sub>p</sub> PRODUCTION ON CARBON METABOLISM IN SERPENTINIZATION SYSTEMS

#### William D. Orsi

Department of Earth and Environmental Sciences, Paleontology and Geobiology, GeoBio-Center, Ludwig-Maximilians-University Munich, 80333 Munich, Germany

The submarine alkaline hydrothermal vent theory for the emergence of life involves thermodynamic disequilibria across geochemical gradients of pH, temperature, redox and H,... The theory holds that in pre-biotic alkaline hydrothermal vents, a primordial metabolism was characterized by CO, being reduced abiotically via H, produced abiotically from serpentinization: the naturally occurring reaction that occurs when water reacts with ultramafic seafloor rocks. How the abiotically produced hydrogen gas that is released by the serpentinization reaction is connected to carbon metabolism at hydrothermal settings in the deep sea is poorly understood. and has not yet been quantified. Here we report the first quantification of H<sub>2</sub> effects on metabolism of CO<sub>2</sub>, acetate, and formate in a serpentinization setting at the Mid-Atlantic Ridge, where seafloor spreading exposes new ultramafic rocks to seawater releasing abundant abiotic H2. The results show a strong coupling between abiotic H<sub>2</sub> production, and the utilization of CO<sub>2</sub> and acetate, but revealed that high H<sub>a</sub> concentrations significantly reduce the amount of formate that can be metabolized by micro-organisms due to inhibition of the enzyme formate dehydrogenase. Reconstruction of the metabolic pathways involved reveal the role of acetate kinase and several carbon fixation pathways that are stimulated by the presence of H<sub>2</sub> in the serpentinization setting. These data provide the first constraints on how H, influences metabolism in modern serpentinization settings. Experimental approaches to produce a laboratory simulation of a prebiotic alkaline hydrothermal vent using an anoxic 'iron ocean' analog to the Hadean ocean have furthermore yielded preliminary results, whereby chimneys composed of green rust and white rust can be created under controlled anoxic conditions. Preliminary results show that the highly reactive green rust minerals bind and thereby concentrate high molecular weight DNA, which would have been a possibly important mechanism concentrating biomolecules in similar alkaline hydrothermal settings in a pre-biotic world.

🞽 anutman@uow.edu.au

### Friday, 10 July, 11:25 CEST



### **SESSION X**

### SESSION X

### ROADMAP TO BUILDING A CELL

#### Christophe Danelon, Zhanar Abil

Department of Bionanoscience, Kavli Institute of Nanoscience, Delft University of Technology, The Netherlands

The cell is the basic unit of life. Our group is engaged in the long-term effort to build a synthetic cell using a bottom-up biology approach. The core architecture of our minimal synthetic cell consists of a cell-free gene expression system (called PURE system) encapsulated inside a lipid vesicle compartment (called liposome). Using in-liposome synthesis of proteins from DNA templates, we aim to reconstitute four essential cellular modules: DNA replication, vesicle growth through phospholipid biosynthesis, liposome division and biogenesis of the transcriptiontranslation machinery. Our latest results on these different research fronts will be presented. Finally, we will discuss a strategy to apply in vitro continuous evolution for the functional integration of these modules into a self-replicating autonomous cell.

### PREBIOTIC SELECTION PRESSURES SHAPE THE **EVOLUTION OF PROTOCELLS**



#### Sudha Rajamani

Department of Biology, Indian Institute of Science Education and Research, Pune, 411008, India

Protocells are primitive cellular entities that are thought to have emerged during the dawn of life on Earth. Their membranes would have been composed of mixtures of single chain amphiphiles such as fatty acids and their derivatives; moieties found in a complex prebiotic chemical landscape. The composition and the physico-chemical properties of these prebiological membranes would have been significantly affected and regulated by the physical environment that surrounded them. I will discuss what we have gleaned from studying the properties of prebiotically relevant membrane systems under pertinent selection pressures (e.g. varying pH, divalent ion concentrations etc). Our results demonstrate how environmental constraints could have shaped the landscape of early membranes. They also illustrate that hetergeneous membrane systems are more stable and robust to multiple selection pressures, thereby making them more suitable for supporting protocellular life.

### Friday, 10 July, 12:25 CEST



### **SESSION XI**

### SEARCHING FOR EARLY PROTEINS IN RANDOMNESS



#### Klára Hlouchová

Department of Cell Biology, Faculty of Science, Charles University, Biocev, Praque, Czech Republic

The earliest proteins had to evolve from thoroughly random sequences. Because today's proteins are mostly highly evolved for specific functions and often rely on defined structural arrangements, it has been assumed that earliest proteins would not be able to support the early biosphere without a dominant or significant aid of other biomolecules (such as RNA and small cofactors). To explore the potential of unnatural (unevolved) sequence space, we have performed a systematic computational and experimental exploration of random sequences. We found that the overall secondary structure and physicochemical properties of random and biological sequences are very similar. Random sequences can be both structured and disordered while the latter have lower aggregation properties and seem to better tolerated by living cells (Tretyachenko et al., Sci Rep 2017, 7.1: 15449). To better navigate protein sequence space, a new bioinformatic tool for combinatorial library design (CoLiDe) will be introduced, offering precise control over protein sequence composition, length and diversity (Tretyachenko & Voracek et al., submitted). This algorithm can be used to search for some functional phenomena that rely on specific amino acid composition and also to bias alphabets towards the amino acid composition of early biosphere. Two exemplary studies where protein composition was reduced to early amino acids will be shown in addition: (i) a ribosomal RNA-binding domain engineered from only 10 evolutionary old amino acids (lacking aromatic and positively charged residues), and (ii) a dephospho-CoA kinase mutated to lack all aromatic amino acids (Giacobelli et al. and Makarov et al., manuscripts in preparation). Our studies indicate that random (unevolved) sequences can give rise to both structured and disordered sequences. In addition, while early alphabets probably missed some of the most functional amino acids of today's proteins, our studies support that they could still serve specific molecular interactions and enzymatic activities.

### **SESSION XI**

### PREBIOTIC MEMBRANES BIND PROTOCELL BUILDING BLOCKS AND CATALYZE FORMATION OF BIOPOLYMERS

Roy A. Black, Zachary R. Cohen, Mengjun Xue, Avijit Hazra, Caitlin E. Cornell, Brenda Kessenich, Richard S. Johnson, Gary P. Drobny, Goiko Lalic, Sarah L. Keller

Departments of Chemistry and Genome Sciences, University of Washington, Seattle, WA USA

How were the prebiotic building blocks of biopolymers brought together and then joined to form RNA and protein, and how did these polymers co-localize with membranes? Membranes form spontaneously from prebiotic amphiphiles such as fatty acids. We have investigated whether membranes composed of decanoic acid, a fatty acid found in meteorites, could have bound RNA and protein building blocks and catalyzed polymer formation. We find that these membranes do bind nucleobases, sugars, nucleosides and amino acids. Within each of these classes, some compounds bind better than others, helping to explain the limited diversity of cellular components. Moreover, binding the building blocks stabilizes the decanoic acid vesicles against the disruptive effects of various salts, explaining how protocells persisted on the early Earth. To explore whether membranes promote the polymerization of building blocks, we dehydrated a solution containing decanoic acid vesicles and an amino acid-ethylester. We found that this procedure generates more dipeptide than dehydration in the absence of vesicles. Moreover, the dipeptide binds the vesicles better than the unjoined amino acid does. These results readily explain how peptides became associated with membranes. More broadly, we propose that biological evolution began with the evolution of increasingly stable fatty acid vesicles that bound and polymerized RNA and protein building blocks.

### Friday, 10 July, 15:25 CEST



### **SESSION XII**

### SESSION XII

### ONSET OF NATURAL SELECTION IN POPULATIONS OF AUTOCATALYTIC HETEROPOLYMERS

#### Sergei Maslov

Dept. of Bioengineering, Dept. of Physics, and Carl R. Woese Institute for Genomic Biology, University of Illinois at Urbana-Champaign, Urbana, IL USA

Reduction of the information entropy along with ever-increasing complexity is among the key signatures of life. Understanding the onset of such behavior in the early prebiotic world is essential for solving the problem of the origin of life. We studied a general problem of heteropolymers capable of template-assisted ligation based on Watson-Crick-like hybridization<sup>1,2</sup>. The system is driven out of equilibrium by cyclic changes in the environment. We modeled the dynamics of 2-mers, i.e., sequential pairs of specific monomers within the heteropolymer population<sup>1</sup>. While the possible number of them is Z<sup>2</sup> (where Z is the number of monomer types), we observe that most of the 2-mers get extinct, leaving no more than 2Z survivors. This leads to a dramatic reduction of the information entropy in the sequence space. This natural-selectionlike process ultimately results in a limited subset of polymer sequences. Importantly, the set of surviving sequences depends on the initial concentrations of monomers and remains exponentially large (2<sup>L</sup> reduced down from Z<sup>L</sup> for chains of length L) in each of realizations. Thus, an inhomogeneity in the initial conditions allows for a massively parallel search of the sequence space for biologically functional polymers, such as ribozymes. The problem has a surprising connection<sup>3</sup> to microbial ecology in which multiple species (analogous to autocatalytic 2-mers) compete for essential nutrients of two types (Z types of left and right ends of chains). Finally, I describe preliminary experimental results<sup>4</sup> validating the predictions of our model.

#### References

1 Tkachenko AV, Maslov S. Onset of natural selection in auto-catalytic heteropolymers (2018) J Chem Phys. 149, 134901 https://doi. org/10.1063/1.5048488.

2 Tkachenko AV, Maslov S. (2015) Spontaneous emergence of autocatalytic information-coding polymers. J Chem Phys. 143(4):045102. https://doi.org/10.1063/1.4922545.

3 Dubinkina V, Fridman Y, Pandey PP, Maslov S (2019) Multistability and regime shifts in microbial communities explained by competition for essential nutrients. eLife 8:e49720; : https://doi.org/10.7554/eLife.49720. Alternative stable states in a model of microbial community limited by multiple essential nutrients. bioRxiv 439547 [Preprint]. October 11, 2018. Available from: https://doi. org/10.1101/439547.

4 Kudella PW, Tkachenko AV, Maslov S, Braun D, Sequence self-selection by the network dynamics of random ligating oligomer pools, (in preparation) (2020)

### SELECTION VIA PHASE SEPARATION

#### Christoph A. Weber

Max Planck Institute for the Physics of Complex Systems, Dresden, Germany

Pre-biotic systems are complex aqueous mixtures composed of thousands of different heteropolymers which undergo chemical reactions. In such chemically-active, multi-component mixtures, enrichment and selection of a small set of components is crucial for the emergence of functional reaction cycles. However, in mixtures of a very large number of different components, each individual component is typically too diluted impeding the emergence of functionality through robust reaction cycles in pre-biotic mixtures.

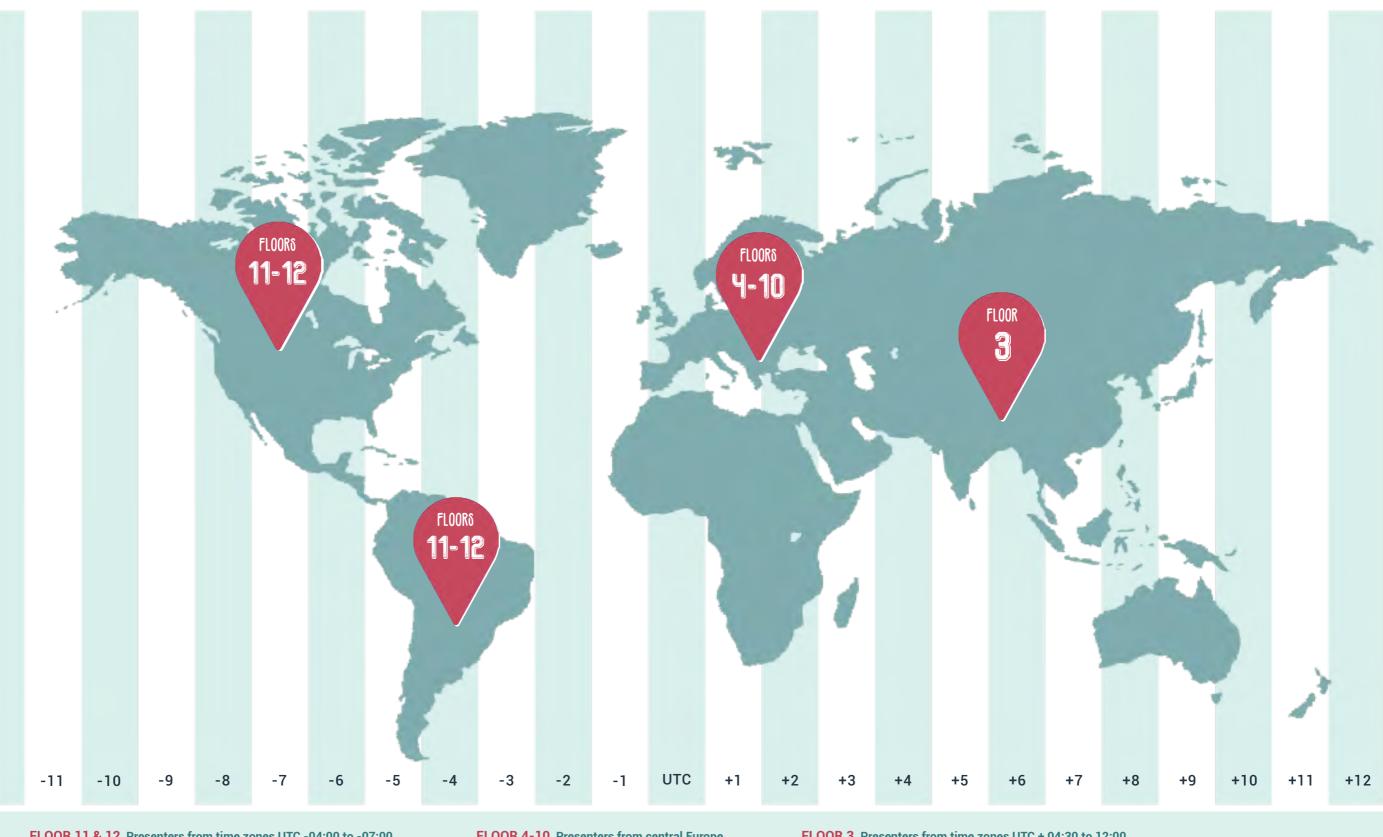
Here, we propose a selection mechanism relevant for prebiotic mixtures based on cycles of phase separation combined with material exchange of the dense phase with a reservoir. We find a selective enrichment of components up to two orders of magnitude. The selection kinetics coincides with a growth of the dense phase up to the system volume leading to a final state where the selected components are condensed. For a prebiotic soup, our findings indicate that cycles of phase separation and material exchange with a reservoir, e.g. the accumulation DNA gel in rock pores periodically filled with DNA rich aqueous solution, could provide a mechanism for the selection and enrichment of specific heteropolymers sequences in a multi-component mixture at the origin of life.



### Friday, 10 July, 16:25 CEST



# POSTERS BY TIME ZONES



FLOOR 11 & 12 Presenters from time zones UTC -04:00 to -07:00 Visit at late hours of the event or during poster session

FLOOR 4-10 Presenters from central Europe Aligns with meeting schedule.

**FLOOR 3** Presenters from time zones UTC + 04:30 to 12:00 Visit at early hours of the event. The presenters will probably not be available during the poster session.

### FLOOR 3

- SANDEEP AMETA 49 Darwinian properties and their trade-offs in autocatalytic reaction networks
- SHIKHA DAGAR 50 Geochemical constraints shaping the nonenzymatic oligomerization of cyclic nucleotides
- 51 MARYAM FATHIAN Levy swimmer in an asymmetric channel
- 52 TONY Z JIA DNA liquid crystal coacervates and polyester microdroplets as model systems in origins of life research
- 53 MINORU KURISU Reproduction of vesicles coupled with a membrane surface-confined template polymerization
- 54 YAMEI LI Geo-electrochemical decomposition of amino acids on icy planetesimals reproduces organics in carbonaceous meteorites
- 55 DEHAI LIANG Protocell with membraneless "Organelles" formed by liquid-liquid phase separation
- 56 JAYANTA NANDA Emergence of native peptide sequences in the context of Origins of Life

#### 57 SUSOVAN SARKAR The influence of compositional heterogeneity of model protocellular

membranes: Implications for the emergence of cellular life

- 58 HEMACHANDER SUBRAMANIAN Identification of evolutionary advantage of some fundamental properties of DNA from replicative potential maximization of primordial autocatalytic heteropolymers
- 59 **HIRONORI SUGIYAMA** Abiotic accumulation of small molecules and ions into cellular compartment against a concentration gradient
- 60 NOBUTO TAKEUCHI The origin of the central dogma through conflicting multilevel selection

ZHANAR ABIL Engineering evolvability of self-replicating DNA as a tool towards building a synthetic cell

FLOOR 4

62

- JAIME AGUDO-CANALEJO 63 Cooperatively enhanced reactivity and 'stabilitaxis' of dissociating oligomeric proteins
- GIACOMO BARTOLUCCI 64 Selection via phase separation
- ALEX BLOKHUIS 65 Autocatalysis in chemical networks: unifications and extensions
- CLAUDIA BONFIO 66 Towards the emergence of modern cell membranes
- JOANNA BRAU 67 Thermal stability of metalorganic compounds on volcanic olivine
- FEDERICO CAIMI 68 Self-assembly driven polymerization of activated nucleotides
- ENRICO SANDRO COLIZZI 69 Emergence of novelty and evolutionary transitions
- ÖMER KÜRSAT COSKUN & 70 THOMAS STEINER Tracing primordial metabolism reflected by microorganisms under hydrothermal conditions
- ANDRES DE LA ESCOSURA 71 Biohybrid materials and systems chemistry

72

**CLAUDIA DE MICCO** Anomalous fluctuations and selective extinction in primordial replicators: a "struggle for life" at the origin of biological chirality

CHRISTINA FELICITAS 74 DIRSCHERL Escalation of DNA monomer polymerization in thermal traps

FLOOR 5

77

78

80

81

82

84

- **CARSTEN DONAU** 75 Active coacervate droplets as a platform towards synthetic life
- OLIVIA DOPPLEB 76 Oligonucleotide assemblies as early ribosomes
  - JAN-PETER DUDA Tracking life at the break of dawn-Identification & interpretation of biosignatures in Earth's oldest rocks
  - **SELENE FORGET & MARKUS** MERINGER Computational exploration of lipid chemical space : Predicting assembly using QSPR models
- **TOMMASO FRACCIA** 79 Liquid crystal coacervates: a pathway for biopolymers coevolution and complex proto-cellular structuring
  - TARO FURUBAYASHI Emergence and diversification of a hostparasite RNA ecosystem through Darwinian evolution
  - **THOMAS GEISBERGER &** PHILIPPE DIEDERICH Chemical evolution of biomolecules formed under volcanic hydrothermal conditions
  - EFTAL GEZER Selection pressure: the latitudinal biodiversity gradient is driven by the latitudinal variation of ultraviolet radiation
- **TOBIAS GOEPPEL** 83 A nonequilibrium error filtering mechanism for enzyme-free copying of nucleic acid sequences
  - MAREN HAAS A mineral-catalysed, mechanochemical formose reaction

### FLOOR 6

87

92

- PHILIPP HONEGGER & 86 CHRISTOPH FLAMM Identification of autocatalysis in a r
  - network A graph-topological appr CLAUDIA HUBER
  - Towards a possible acetyleno/ carboxydotrophic core metabolism primordial conditions
- ALAN IANESELLI 88 Heated microdroplets of acidic wate induce the denaturation of oligonuc and the replication of longer strands
- MIKOŁAJ JANICKI 89 Photoreduction of thioanhydroaden the purine deoxyribonucleosides
- ANTON JOSEPH 90 Design of stealth cell-mimetic dendrimersomes
- NOZAIR KHAWAJA 91 Exploring the biogeochemistry of extraterrestrial active ocean worlds
  - SVEN KLUMPE Ultrastructural analysis of nuclear p complex assembly in early Drosoph embryos by cryo
- 93 KAI KOHLER & FLORIAN GA Evolutionary optimization of experi synthetic networks

FIB LiftOut

- 94 BALAZS KONNYU Dynamics and stability in prebiotic information integration: An RNA wo model from first principles
- NINA KOSTINA 95 Sugar-driven formation of artificial raft-domains with hierarchical perio nanoarrays on dendrimersome prot
- 96 PATRICK KUDELLA Sequence self-selection by the dynamics of a random oligomer pool network

# S Ily " 11

202

### FLOOR 7

reaction	98	ALEXANDRA KÜHNLEIN Sequence dependent gelation, accumulation and sedimentation
oroach 1 under	99	CHANDRASHEKHAR V. KULKARNI Estimating preferential localization of interacting molecules in model lipid membranes
ter	100	SUDARSHANA LAHA Droplets as biochemical reactors in living cells
cleotides ds	101	GABRIELLE LEVEAU Bridging the gap between chain formation and genetic copying of RNA
nosine to	102	YONGDA LI Biochemical methods for the detection of proteins as life signatures on Mars-like soils
	103	<b>TIM LICHTENBERG</b> Atmospheric speciation of rocky planets from magma ocean outgassing
s pore	104	<b>KAI LIU</b> Towards darwinian evolution of synthetic replicators
hila ARTNER imental	105	<b>SAVINO LONGO</b> Anomalous fluctuations and selective extinction in primordial replicators: a "struggle for life" at the origin of biological chirality
:	106	<b>OLIVER MAGUIRE</b> Cycling of orthophosphate under mild prebiotically plausible conditions
orld	107	<b>THOMAS MATREUX</b> Heat flows shift chemical equilibria by selective accumulation
l iodic tocells	108	GAIA MICCA LONGO New insights on prebiotic chemistry from plasma kinetics
amics of		

### FLOOR 8

- SAIBAL MITRA 110 The first step from molecules to life: Formation of large random molecules acting as micro-environments
- MAITANE MUÑOZ BASAGOITI 111 Physics and evolution of catalysts
- ATIDA NASUFOVSKA 112 Prebiotic chemical energy flux
- 113 JOANA PEREIRA On the origins of the protein world: A largescale computational approach to study the emergence of the first autonomously folding proteins
- **BENEDIKT PETER & ELIA SALIBI** 114 Freeze-thaw driven proliferation of RNA protocells
- 115 MARTINA PREINER The ambivalent role of water at the origins of life

2

. Ily

- 11 1

- KHOSROW RAHIMI 116 Amphiphilic comb-polymers solve the dilemma of polymer-based cell-mimetic membranes
- MEHRNOUSH RAHIMZADEH 117 Structural design of amphiphilic comb polymers to self-assemble into faceted membrane protocells
- ANA MARIA RESTREPO SIERRA 118 An evolutionary approach for building a synthetic cell
- PAULA CATALINA RODRIGUEZ 119 RAMIREZ Linking microbial diversity to carbon cycling in subseafloor sediments from the Namibian continental shelf
- **CESAR RODRIGUEZ-**120 EMMENEGGER Superselectivity in synthetic protocells

### FLOOR 9

129

131

- SAROJ KUMAR ROUT 122 On the amyloid world hypothesis 123 ALEXANDER RUF The challenging detection of nucleobases from preaccretional astrophysical ice analogs ANNALENA SALDITT 124 A thermal habitat that triggers the retention and RNA-catalyzed replication of RNA
- FABIAN SCHMIDT 125 The Origin of Interstellar CO<sub>2</sub>
- 126 CHRISTIAN SCHMITT RNA library screening for self-aminoacylating tRNA precursors
  - enzymes?
- 128 PETER RICHARD SCHREINER Carbohydrate formation in the absence of biosynthesis
  - Thermal gradient driven formation of homochiral domains in hydrogels starting from racemic polynucleotide mixtures
- 130 Understanding the genetic code from affinities of aminoacyl adenylates to pretransfer RNA motifs
  - Sugars program the hierarchical selfassembly in onion glycodendrimers
- EMILIE SONG 132 Probing RNA stability, formation, and catalysis in simulated prebiotic environments on the early Earth and in Space

### FLOOR 10

135

136

137

138

140

- CHRISTINA SPRINGSKLEE 134 Prebiotic synthesis in volcanic discharges: An experimental approach
  - TOMISLAV STOLAR The solid-state as a reaction medium for prebiotic chemical selection: drv heating enables selective pairing of model nucleobases
  - JAN TENBUSCH Super-flexible biomimetic vesicles
  - BEATRIZ VON DER ESCH Prebiotic pathway to DNA nucleosides
  - MARIIA VOROBII Building membrane machines to endocytize living bacteria: the battle between adhesion and flexibility
- 139 ANNA MARIA WAGNER How does spontaneous curvature induce the morphogenesis of dendrimersome vesicles?
  - CRAIG WALTON Phosphorous mineralogy on the Early Earth
- KARL WIENAND 141 Public outreach on 'Emergence of Life'
- 142 MAX WINKLER UV resistance of nucleosides - an experimental approach
- SREEKAR WUNNAVA 143 Acid-catalyzed polymerization of cyclic GMP
- 144 NOEL YEH MARTIN Out-of-equilibrium cellular mimics driven by thermal gradients

ZACHARY COHEN 146 Fatty acid membranes are stable in carbonate-rich, prebiotic lake enviro

FLOOR 11

- 147 SAURJA DASGUPTA Chemistry and catalysis join forces in ligation
- 148 HADI FARES Impact of wet-dry cycling on the phase compartmentalization behaviors of co coacervates
- JAY FORSYTHE 149 Proline incorporation in model prebi depsipeptides
- MCCAULEY MEYER 150 Nucleotide-level resolution of RNA for interactions within peptide-based cor coacervates
- 151 TRISHOOL NAMANI Role of amino acids on nonenzymatic clay-promoted oligomerization of act nucleotide
- ARTASH NATH 152 Using machine learning to improve pr of chemical composition of exoplane atmospheres
- ALINE NOVAIS 153 Exoplanetary atmospheric retrieval machine learning
- FATMA PIR CAKMAK 154 Prebiotically-relevant low polyion multivalency might improve functiona membraneless compartments
- **RAGHAV POUDYAL** 155 RNA world inside compartments: Acti of RNA catalysis by complex coacerv
- MICHAEL L. WONG & STUAR 156 BARTLETT Defining lyfe in the universe: From three privileged functions to four pillars
- WEN ZHANG 157 Deciphering nonenzymatic RNA polymerization through crystallography

- 127 TOBIAS SCHNITZER Catalytic peptides - Potential precursors of
- - PHILIPP SCHWINTEK
- ADRIANA SERRAO
- DOMINIK SÖDER

### FLOOR 12

	159	MARION ZULEMA ARMAS VAZQUEZ
onments		Computational study of physicochemical
		properties of an adenine synthesis route
		under UV radiation
n RNA	160	JULIAN CORZO
	100	Endolithic culturable bacteria in minerals
		from geologic samplings in Colombia
se and	161	ROMULO LEONCIO
omplex	101	Chemical oscillations in a theoretical system
		of dinitrosyl iron complex (DNIC) with thiol-
		containing ligands
iotic	409	LUIS DELAYE
	162	Was LUCA a hyperthermophilic prokaryote?
		The impact-bottleneck hypothesis revisited
lding		
mplex	163	FRANKY DJUTANTA
		Producing cell-like structures from oil films
		residing on ocean water by raindrop impacts
0	165	GIOVANNA GHIRLANDA
tivated	100	Membraneless organelles by design
	100	ROSA REYES
	166	What about the origin of death?
rediction		
tary	167	DAVID RODRIGUEZ
		Origin of life on ice. Is Mars a potential
		habitable planet?
using	168	ALFREDO RODRÍGUEZ ARTEAGA
		Extremophilic proteins and their resistance
		secrets
	169	LUIS MIGUEL RODRÍGUEZ
ality of	100	TORRES
		Contrast between the main life's origins
		abiogenic models
ivation	170	TOVILLA QUESADA RUBÉN DE
ates		JESÚS
		The importance of laboratory practices on the
RT		origin of life at the upper-middle level

# POSTER PRESENTERS INDEX

S.No	First Name	Last Name	Titles	Floor	Page	1º	S.No	First Name	Last Name	Titles	Floor	Ра
1.	Zhanar	Abil	Engineering evolvability of self-replicating DNA as a tool towards building a synthetic cell	4	62		61.	Kai	Liu	Towards darwinian evolution of synthetic replicators	7	10
2.	Jaime	Agudo-Canalejo	Cooperatively enhanced reactivity and 'stabilitaxis' of dissociating oligomeric proteins	4	63		62.	Savino	Longo	Anomalous fluctuations and selective extinction in primordial replicators: a "struggle for life" at the origin of biological chirality	7	10
3.	Sandeep	Ameta	Darwinian properties and their trade-offs in autocatalytic reaction networks	3	49		63.	Oliver	Maguire	Cycling of orthophosphate under mild prebiotically plausible conditions	7	10
4.	Marion Zulema	Armas Vazquez	Computational study of physicochemical properties of an adenine synthesis route under UV radiation	12	159	M1 11	64.	Thomas	Matreux	Heat flows shift chemical equilibria by selective accumulation	7	10
5.	Stuart	Bartlett	Defining lyfe in the universe: From three privileged functions to four pillars	11	156	- ,1	65.	Markus	Meringer	Computational exploration of lipid chemical space : Predicting assembly using QSPR models	5	7
6.	Giacomo	Bartolucci	Selection via phase separation	4	64	111	66.	McCauley	Meyer	Nucleotide-level resolution of RNA folding interactions within peptide-based complex coacervates	11	15
7.	Alex	Blokhuis	Autocatalysis in chemical networks: unifications and extensions	4	65	1	67.	Gaia	Micca Longo	New insights on prebiotic chemistry from plasma kinetics	7	10
8	Claudia	Bonfio	Towards the emergence of modern cell membranes	4	66	110	68.	Saibal	Mitra	The first step from molecules to life: Formation of large random molecules acting as micro-environments	8	1
9	Joanna	Brau	Thermal stability of metalorganic compounds on volcanic olivine	4	67		69.	Maitane	Muñoz Basagoiti	Physics and evolution of catalysts	8	1
10.	Federico	Caimi	Self-assembly driven polymerization of activated nucleotides	4	68		70.	Trishool	Namani	Role of amino acids on nonenzymatic clay-promoted oligomerization of activated nucleotide	11	1
11.	Zachary	Cohen	Fatty acid membranes are stable in carbonate-rich, prebiotic lake environments	11	146	94	71.	Jayanta	Nanda	Emergence of native peptide sequences in the context of Origins of Life	3	5
12.	Enrico Sandro	Colizzi		4		1	72.	Atida	Nasufovska	Prebiotic chemical energy flux		1'
	Julian	Corzo	Emergence of novelty and evolutionary transitions	10	69	for-	72.	Artash	Nath		11	15
13.			Endolithic culturable bacteria in minerals from geologic samplings in Colombia	12	160	10				Using machine learning to improve prediction of chemical composition of exoplanetary atmospheres	11	15
14.	Omer Kürsat	Coskun	Tracing primordial metabolism reflected by microorganisms under hydrothermal conditions	4	70		74.	Aline	Novais	Exoplanetary atmospheric retrieval using machine learning		
15.	Romulo Leoncio	Cruz Simbron	Chemical oscillations in a theoretical system of dinitrosyl iron complex (DNIC) with thiol-containing ligands	12	161		75.	Joana	Pereira	On the origins of the protein world: A large-scale computational approach to study the emergence of the first autonomously folding protein	is 8	1
16.	Shikha	Dagar	Geochemical constraints shaping the nonenzymatic oligomerization of cyclic nucleotides	3	50		76.	Benedikt	Peter	Freeze-thaw driven proliferation of RNA protocells	8	1
17.	Saurja	DasGupta	Chemistry and catalysis join forces in RNA ligation	11	147	11.	77.	Fatma	Pir Cakmak	Prebiotically-relevant low polyion multivalency might improve functionality of membraneless compartments	11	15
18.	Andres	de la Escosura	Biohybrid materials and systems chemistry	4	71		78.	Raghav	Poudyal	RNA world inside compartments: Activation of RNA catalysis by complex coacervates	11	1
19.	Claudia	De Micco	Anomalous fluctuations and selective extinction in primordial replicators: a "struggle for life" at the origin of biological chirality	4	72		79.	Martina	Preiner	The ambivalent role of water at the origins of life	8	1
20.	Luis	Delaye	Was LUCA a hyperthermophilic prokaryote? The impact-bottleneck hypothesis revisited	12	162		80.	Khosrow	Rahimi	Amphiphilic comb-polymers solve the dilemma of polymer-based cell-mimetic membranes	8	1
21.	Philippe	Diederich	Chemical evolution of biomolecules formed under volcanic hydrothermal conditions	5	81		81.	Mehrnoush	Rahimzadeh	Structural design of amphiphilic comb polymers to self-assemble into faceted membrane protocells	8	1
22.	Christina Felicitas	Dirscherl	Escalation of DNA monomer polymerization in thermal traps	5	74		82.	Ana Maria	Restrepo Sierra	An evolutionary approach for building a synthetic cell	8	1
23.	Franky	Djutanta	Producing cell-like structures from oil films residing on ocean water by raindrop impacts	12	163	A	83.	Rosa	Reyes	What about the origin of death?	12	1
24.	Carsten	Donau	Active coacervate droplets as a platform towards synthetic life	5	75	7 8	84.	David	Rodriguez	Origin of life on ice. Is Mars a potential habitable planet?	12	1
25.	Olivia	Doppleb	Oligonucleotide assemblies as early ribosomes	5	76	$\lambda = $	85.	Alfredo	Rodríguez Arteaga	Extremophilic proteins and their resistance secrets	12	1
26.	Jan-Peter	Duda	Tracking life at the break of dawn—Identification & interpretation of biosignatures in Earth's oldest rocks	5	77	F	86.	Paula Catalina	Rodriguez Ramirez	Linking microbial diversity to carbon cycling in subseafloor sediments from the Namibian continental shelf	8	1
27.	Hadi	Fares	Impact of wet-dry cycling on the phase and compartmentalization behaviors of complex coacervates	11	148	1	87.	Luis Miguel	Rodríguez Torres	Contrast between the main life's origins abiogenic models	12	1
28.	Maryam	Fathian	Levy swimmer in an asymmetric channel	3	51		88.	Cesar	-	r Superselectivity in synthetic protocells	8	1
29.	Christoph	Flamm	Identification of autocatalysis in a reaction network – A graph-topological approach	6	86		89.	Saroj Kumar	Rout	On the amyloid world hypothesis	9	1
	Selene	Forget		5		Allet	90.	Alexander			, ,	1
30.			Computational exploration of lipid chemical space : Predicting assembly using QSPR models	11	78	W.			Ruf	The challenging detection of nucleobases from preaccretional astrophysical ice analogs	9	1
31.	Jay T	Forsythe	Proline incorporation in model prebiotic depsipeptides	-	149	11.5	91.	Annalena	Salditt	A thermal habitat that triggers the retention and RNA-catalyzed replication of RNA	9	
32.	Tommaso _	Fraccia	Liquid crystal coacervates: a pathway for biopolymers coevolution and complex proto-cellular structuring	5	79	-2 "	92.	Elia	Salibi	Freeze-thaw driven proliferation of RNA protocells	8	1
33.	Taro	Furubayashi	Emergence and diversification of a host-parasite RNA ecosystem through Darwinian evolution	5	80		93.	Susovan	Sarkar	The influence of compositional heterogeneity of model protocellular membranes: Implications for the emergence of cellular life	3	
34.	Florian	Gartner	Evolutionary optimization of experimental synthetic networks	6	93	-	94.	Fabian	Schmidt	The Origin of Interstellar CO2	9	1
35.	Thomas	Geisberger	Chemical evolution of biomolecules formed under volcanic hydrothermal conditions	5	81		95.	Christian	Schmitt	RNA library screening for self-aminoacylating tRNA precursors	9	1
36.	Eftal	Gezer	Selection pressure: the latitudinal biodiversity gradient is driven by the latitudinal variation of ultraviolet radiation	5	82		96.	Tobias	Schnitzer	Catalytic peptides – Potential precursors of enzymes?	9	1i
37.	Giovanna	Ghirlanda	Membraneless organelles by design	12	165	-	97.	Peter Richard	Schreiner	Carbohydrate formation in the absence of biosynthesis	9	18
38.	Tobias	Goeppel	A nonequilibrium error filtering mechanism for enzyme-free copying of nucleic acid sequences	5	83		98.	Philipp	Schwintek	Thermal gradient driven formation of homochiral domains in hydrogels starting from racemic polynucleotide mixtures	9	18
39.	Maren	Haas	A mineral-catalysed, mechanochemical formose reaction	5	84	2	99.	Adriana	Serrao	Understanding the genetic code from affinities of aminoacyl adenylates to pre-transfer RNA motifs	9	18
40.	Philipp	Honegger	Identification of autocatalysis in a reaction network – A graph-topological approach	6	86		100.	Dominik	Söder	Sugars program the hierarchical self-assembly in onion glycodendrimers	9	1
41.	Claudia	Huber	Towards a possible acetyleno/carboxydotrophic core metabolism under primordial conditions	6	87		101.	Emilie	Song	Probing RNA stability, formation, and catalysis in simulated prebiotic environments on the early Earth and in Space	9	18
42.	Alan	Ianeselli	Heated microdroplets of acidic water induce the denaturation of oligonucleotides and the replication of longer strands	6	88		102.	Christina	Springsklee	Prebiotic synthesis in volcanic discharges: An experimental approach	10	18
43.	Mikołaj	Janicki	Photoreduction of thioanhydroadenosine to the purine deoxyribonucleosides	6	89		103.	Thomas	Steiner	Tracing primordial metabolism reflected by microorganisms under hydrothermal conditions	4	
44.	Tony Z	Jia	DNA liquid crystal coacervates and polyester microdroplets as model systems in origins of life research	3	52		104.	Tomislav	Stolar	The solid-state as a reaction medium for prebiotic chemical selection: dry heating enables selective pairing of model nucleobases	10	1
45.	Anton	Joseph	Design of stealth cell-mimetic dendrimersomes	6	90	えき	105.	Hemachander	Subramanian	Identification of evolutionary advantage of some fundamental properties of DNA from replicative potential maximization of primordial	3	
46.	Nozair	Khawaja	Exploring the biogeochemistry of extraterrestrial active ocean worlds	6	91	dus-				autocatalytic heteropolymers	2	
47.	Sven	Klumpe	Ultrastructural analysis of nuclear pore complex assembly in early Drosophila embryos by cryoFIB LiftOut	ĥ	92	17	106.	Hironori	Sugiyama	Abiotic accumulation of small molecules and ions into cellular compartment against a concentration gradient	2	
48.	Kai	Kohler	Evolutionary optimization of experimental synthetic networks	a a	92 93	1	100.	Nobuto	Takeuchi		2	, i
	Balazs			e e		31/	107.	Jan		The origin of the central dogma through conflicting multilevel selection	- J 10	1
49. 50		Konnyu Kostina	Dynamics and stability in prebiotic information integration: An RNA world model from first principles	0	94	12/1			Tenbusch	Super-flexible biomimetic vesicles	10	1
50.	Nina	Kostina	Sugar-driven formation of artificial raft-domains with hierarchical periodic nanoarrays on dendrimersome protocells	0	95	1 3.1	109.	Rubén de Jesús	Tovilla Quesada	The importance of laboratory practices on the origin of life at the upper-middle level	12	1
51.	Patrick	Kudella	Sequence self-selection by the dynamics of a random oligomer pool network	6 	96		110.	Beatriz	von der Esch	Prebiotic pathway to DNA nucleosides	10	
52.	Alexandra	Kühnlein	Sequence dependent gelation, accumulation and sedimentation	7	98		111.	Mariia	Vorobii	Building membrane machines to endocytize living bacteria: the battle between adhesion and flexibility	10	1
53.	Chandrashekhar V.	Kulkarni	Estimating preferential localization of interacting molecules in model lipid membranes	7	99	N. Con	112.	Anna Maria	Wagner	How does spontaneous curvature induce the morphogenesis of dendrimersome vesicles?	10	1
54.	Minoru	Kurisu	Reproduction of vesicles coupled with a membrane surface-confined template polymerization	3	53		113.	Craig	Walton	Phosphorous mineralogy on the Early Earth	10	1
55.	Sudarshana	Laha	Droplets as biochemical reactors in living cells	7	100	1 7/1	114.	Karl	Wienand	Public outreach on 'Emergence of Life'	10	1
56.	Gabrielle	Leveau	Bridging the gap between chain formation and genetic copying of RNA	7	101	R. 1/4-	115.	Max	Winkler	UV resistance of nucleosides - an experimental approach	10	1
57.	Yamei	LI	Geo-electrochemical decomposition of amino acids on icy planetesimals reproduces organics in carbonaceous meteorites	3	54	YE	116.	Michael L	Wong	Defining lyfe in the universe: From three privileged functions to four pillars	11	1
58.	Yongda	Li	Biochemical methods for the detection of proteins as life signatures on Mars-like soils	7	102	V	117.	Sreekar	Wunnava	Acid-catalyzed polymerization of cyclic GMP	10	1
	Dehai	Liang	Protocell with membraneless "Organelles" formed by liquid-liquid phase separation	3	55		118.	Noel	Yeh Martin	Out-of-equilibrium cellular mimics driven by thermal gradients	10	1
59.	Denai											



### INDEX - FLOOR 3 (UTC + 04.30 TO 12.00)

Visit at early hours of the event. The presenters will probably not be available during the poster session

**49** SANDEEP AMETA

Darwinian properties and their trade-offs in autocatalytic reaction networks

50 SHIKHA DAGAR

Geochemical constraints shaping the nonenzymatic oligomerization of cyclic nucleotides

**51** MARYAM FATHIAN Levy swimmer in an asymmetric channel

\_\_\_\_\_

52 TONY Z JIA

DNA liquid crystal coacervates and polyester microdroplets as model systems in origins of life research

53 MINORU KURISU

Reproduction of vesicles coupled with a membrane surface-confined template polymerization

54 YAMEI LI

Geo-electrochemical decomposition of amino acids on icy planetesimals reproduces organics in carbonaceous meteorites

55 DEHAI LIANG

Protocell with membraneless "Organelles" formed by liquid-liquid phase separation

**56** JAYANTA NANDA

Emergence of native peptide sequences in the context of Origins of Life

57 SUSOVAN SARKAR

The influence of compositional heterogeneity of model protocellular membranes: Implications for the emergence of cellular life

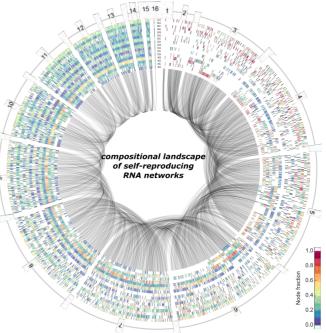
### 58 HEMACHANDER SUBRAMANIAN

Identification of evolutionary advantage of some fundamental properties of DNA from replicative potential maximization of primordial autocatalytic heteropolymers

59 HIRONORI SUGIYAMA

Abiotic accumulation of small molecules and ions into cellular compartment against a concentration gradient

**60 NOBUTO TAKEUCHI** The origin of the central dogma through conflicting multilevel selection FLOOR 3



### DARWINIAN PROPERTIES AND THEIR TRADE-OFFS IN AUTOCATALYTIC REACTION NETWORKS



Ameta S.<sup>¥,1,3</sup>, Arsène S.<sup>¥,1</sup>\*, Foulon S.<sup>1</sup>, Saudemont B.<sup>1</sup>, Clifton B.E.<sup>2</sup>, Griffiths A.D.<sup>1\*</sup>, Nghe P.<sup>1\*</sup>

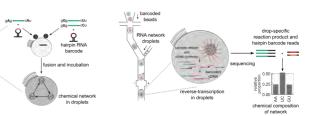
<sup>1</sup> Laboratoire de Biochimie, ESPCI Paris, 10 Rue Vauquelin, 75005 Paris, France <sup>2</sup> School of Chemistry and Biochemistry, Georgia Institute of Technology, Atlanta, Georgia 30332-0400

<sup>3</sup> Simons Centre for the Study of Living Machines, National Centre for Biological Sciences, Tata Institute of Fundamental Research, Bengaluru 560065, India.

\*Both authors contributed equally: sandeepameta@ncbs.res.in \*Corresponding authors: andrew. griffiths@espci.fr, philippe.nghe@espci. fr

> 1 Butlerov and 2 Miras H. N., e 3 Nanda, J., et 4 Sievers, D. ar 5 Ameta, S.\*, A 6 Arsene, S\*., *J* 7 Vaidya, N., et

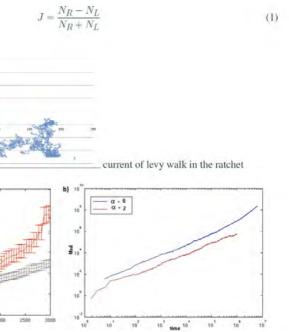
Discovering autocatalytic chemistries that can evolve is a major goal in systems chemistry and a critical step towards understanding the origin of life. Autocatalytic networks have been discovered in various chemistries<sup>1-4</sup>, but we lack a general understanding of how network topology controls the Darwinian properties of variation, differential reproduction, and heredity, which are mediated by the chemical composition. Using barcoded sequencing and droplet microfluidics, we establish a landscape of thousands of networks of RNAs<sup>5</sup> (figure) that catalyze their own formation from fragments<sup>6,7</sup>, and derive relationships between network topology and chemical composition<sup>5</sup>. We find that strong variations arise from catalytic innovations perturbing weakly connected networks, and that reproduction increases with global connectivity. These rules imply trade-offs between reproduction and variation, and between compositional persistence and variation along trajectories of network complexification. Overall, connectivity in reaction networks provides a lever to balance variation (to explore chemical states) with reproduction and heredity (persistence being necessary for selection to act), as required for chemical evolution.

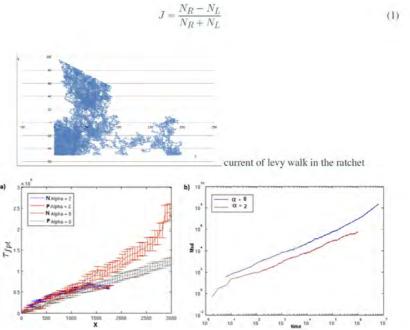


- 1 Butlerov and A.M. (1861) Zeitschrift fur Chemie 4: 549-560.
- 2 Miras H. N., et al. (2019) ChemRxiv, Preprint. https://doi.org/10.26434/chemrxiv.9598442.v1. 3 Nanda, J., et al. (2017) Nature Communications 8: 434.
- 4 Sievers, D. and G. von Kiedrowski (1994) Nature 369: 221-4.
- 5 Ameta, S.\*, Arsene S.\*, et al. (2019) bioRxiv (https://doi.org/10.1101/726497).
- 6 Arsene, S\*., Ameta S\*., et al. (2018) Nucleic Acids Research 46: 9660-9666.
- 7 Vaidya, N., et al. (2012) Nature 491: 72-7.

### FLOOR 3

The Molecular Origins of Life Conference, Munich 2020



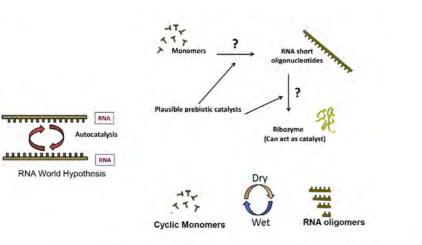


### LEVY SWIMMER IN AN ASYMMETRIC CHANNEL



Maryam Fathian<sup>1</sup>, Ali Najafi<sup>1,2</sup> <sup>1</sup> Department of Physics; University of 7anian

<sup>2</sup> Institute for Advanced Studies in Basic Sciences (IASBS), Zanjan



Geochemical constraints facilitating nonenzymatic oligomerization of cyclic nucleotides

CO+L lob

Nucleobase

Cyclic nucleotide

### GEOCHEMICAL CONSTRAINTS SHAPING THE NONENZYMATIC **OLIGOMERIZATION OF CYCLIC NUCLEOTIDES**



Shikha Dagar, Susovan Sarkar and Sudha Rajamani Department of Biology, Indian Institute of Science Education and Research, Pune 411008, India

1 Gilbert, W. The RNA world, Nature (1986) 2 Zaug, A. & Cech, T. The intervening sequence RNA of Tetrahymena is an enzyme. Science (80-. ). 231, 470-475 (1986). 3 Glasner, M. E. et al. Metal ion requirements for structure and catalysis of an RNA ligase ribozyme. Biochemistry 41, 8103-8112 (2002).

4 NASA Spaceward Bound India Program; Ladakh India 2016: spacewardbound.astrobiologyindia.in/field-site-ladakh/

About one-third of the extant enzymes are metalloenzymes. This ubiquitous nature of metal ions in extant life highlights their potential role in the emergence and evolution of early life on Earth. Specifically, metal ions are thought to have played the role of catalysts on early Earth in prebiotically pertinent processes. Among several competiting theories that hypothesize how life would have originated on Earth, RNA World Hypothesis happens to be the most prevalent one. It suggests that RNA would have been the first relevant biomolecule to set the stage for life's emergence, and whose formation would have been driven by nonenzymatic processes1. This is due to its capability to act as a catalyst in addition to being a genetic material (Carl Woese, Francis Crick and Leslie Orgel, 1960s)<sup>2</sup>. However, spontaneous generation of RNA, which involves energitically uphill condensation reactions among nucleotides, is thought to be non-trivial. Nonetheless, even if long RNAs could indeed form by nonenzymatic means, they would require metal ions to fold appropriately and catalyze reactions<sup>3</sup>.

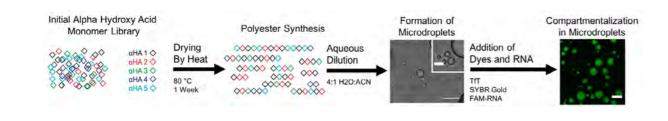
In this study, the effect of metal ions on the nonenzymatic oligomerization of prebiotically relevant cyclic nucleotides was investigated. Reactions were performed under, both, laboratory simulated prebiotic conditions, and using water samples collected from an astrobiologically relevant early Earth analogue site in Ladakh, India<sup>4</sup>. This was done to evaluate the robustness of the results obtained from a predetermined laboratory set up. Furthermore, it also enables an understanding of how a more 'realistic' scenario, wherein multiple entities like metal ions and other co-solutes are present, could affect the otherwise 'controlled' reactions. Under aqueous conditions, intact oligomers up to tetramers were observed. However, in the reactions performed under analogue conditions, even-though the hot-spring water samples seemed to enhance the rate of oligomerization, they also led to the quick destabilization of the resultant oligomers. Our results suggest that the presence of ions could effectively act as a selection pressure under prebiotic scenarios, thereby playing a vital role in shaping the evolutionary landscape of a putative RNA World.

GO Weblink to the Poster Room

We numerically study the dynamics of a model Levy walker, moving in a 2-dimensional medium confined by the walls of an asymmetric channel. We show that, as a result of both asymmetric potential due to the channel and also the power law step size distribution, the Levy walker will achieve a net directed velocity in the direction preferred by the channel. Other statistical properties of the walker such as mean first passage time and the mean square displacements are also examined.

### FLOOR 3

of Aniline



### DNA LIQUID CRYSTAL COACERVATES AND POLYESTER MICRODROPLETS AS MODEL SYSTEMS IN ORIGINS OF LIFE RESEARCH



Tony Z. Jia<sup>1,2</sup>, Kuhan Chandru<sup>3,4</sup>, Yayoi Hongo<sup>1</sup>, Rehana Afrin<sup>1</sup>, Tomohiro Usui<sup>1,5</sup>, Kunihiro Myojo<sup>6</sup>, H. James Cleaves II<sup>1,2,7</sup>, Tommaso P. Fraccia<sup>8</sup> <sup>1</sup> Farth-Life Science Institute, Tokyo

Institute of Technology <sup>2</sup> Blue Marble Space Institute of Science <sup>3</sup> University of Chemistry and Technology, Prague <sup>4</sup>Space Science Centre (ANGKASA), Institute of Climate Change, National

University of Malaysia <sup>5</sup> Institute of Space and Astronautical Science, Japan Aerospace Exploration Agency

<sup>6</sup> Department of Earth and Planetary Science, Tokyo Institute of Technology <sup>7</sup>Institute for Advanced Study, Princeton University <sup>8</sup> Institut Pierre-Gilles de Gennes, Chimie

Biologie et Innovation, ESPCI Paris, PSL University

Liquid-liquid phase separation (LLPS) controls important biological processes including catalysis and gene regulation. Due to their ease of in vitro assembly into membraneless compartments and their presence within modern cells, LLPS systems have been postulated to be one potential form that the first cells on Earth took on. Here, we present work regarding the structure, assembly, and function of various in vitro LLPS systems, such as DNA liquid crystal coacervates and polyester microdroplets, that produce membraneless compartments which may have been relevant to the emergence of primitive compartments. These compartments exhibit various properties, such as compartmentalization and self-assembly, and catalysis, potentially providing scaffolds that could have effected the assembly of more complex chemical structures. While there are still a number of remaining open questions regarding LLPS systems as models for primitive membraneless cells, including how modern biologies acquired such membraneless organelles, understanding the exact connection between LLPS systems and primitive cells will shed light into how primitive cells transitioned to modern cells.

**REPRODUCTION OF VESICLES COUPLED WITH** A MEMBRANE SURFACE-CONFINED TEMPLATE POLYMERIZATION

Vesicle

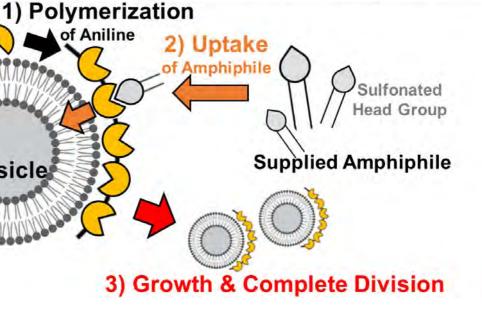


Minoru Kurisu<sup>1</sup>, Harutaka Aoki<sup>1</sup>, Takehiro Jimbo<sup>1</sup>, Yuka Sakuma<sup>1</sup>, Sandra Luginbuhl<sup>2</sup>, Peter Walde<sup>2</sup>, Masayuki Imai<sup>1</sup> Department of Physics, Tohoku University, Japan, <sup>2</sup> Department of Materials, ETH Zürich,

Switzerland

vesicles<sup>3</sup>.

### **Reproduction of Vesicle coupled with Surface-Confined Polymerization**



Molecular assembly system that have autonomous reproduction ability can be considered as minimal cell-like systems, which bridges non-living and living forms of matter<sup>1</sup>. Here we show the reproduction of cell-sized vesicles coupled with polymerization on the surface of vesicles. The particular reaction used is the template polymerization of aniline occurring on the surface of AOT vesicles, which yields polyaniline emeraldine salt form (PANI)<sup>2</sup>. When AOT micelles are microinjected to AOT vesicles during polymerization, the AOT - PANI-ES vesicles selectively incorporate them in their membrane, which leads to a growth of the vesicle. If the AOT vesicles contained cholesterol, the vesicle not only showed growth, but also division, i.e., reproduction of

1 J. Von Neumann, Cerebral mechanisms in behavior, 1, 1 (1951) 2 K.Junker et al., RSC Adv., 2, 6478 (2012) 3 M.Kurisu et al., Communications Chemistry, 2:117 (2019)

### **GEO-ELECTROCHEMICAL** DECOMPOSITION OF AMINO ACIDS ON ICY PLANETESIMALS REPRODUCES ORGANICS IN CARBONACEOUS **METEORITES**

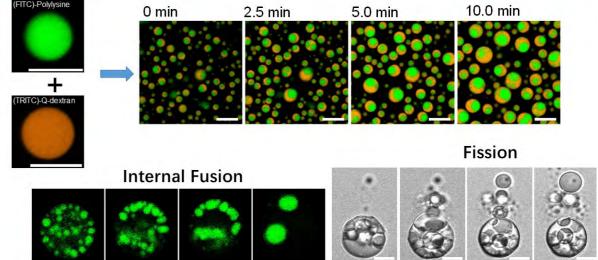


Li Yamei<sup>1</sup>, Norio Kitadai<sup>1,2</sup>, Ryuhei Nakamura<sup>1</sup>, Yasuhito Sekine<sup>1</sup> Earth-Life Science Institute, Tokyo Institute of Technology Japan Agency for Marine-Earth Science and Technology

Organics in carbonaceous meteorites are the remnants of primitive solar system chemistry and parent-body processes. Amino acids are ubiquitously found in carbonaceous chondrites, and their genesis and conversion have important implications on the origin of biomolecules, the geophysical evolution and the geochemical processes that occurred in their parent bodies (icy planetesimals).

In Murchison meteorite and several reported carbonaceous meteorites, in addition to amino acids, other types of water-soluble, low molecular weight compounds make a complex suite that includes hydroxycarboxylic acid, aliphatic monoamines, monocarboxylic acids, alcohols, and others. However, the synthetic origin for their formation leading to their occurrence in meteorites remain poorly understood, becoming a long-standing enigma.

It has been largely considered that water-rock interactions cause the alteration of organic distribution and associated minerals in carbonaceous bodies, however, how does energy transduction proceed in such environment and how the chemical processes have shaped the organic distributions remain poorly understood. Here we report that geo-electrochemical processes can decompose aliphatic amino acids into monocarboxylic acids, alcohols, amines, and hydroxycarboxylic acids. Iron and nickel sulfides work as effective catalysts for the redox-mediated activation of amino acids. Such geo-electrochemical processes are driven spontaneously by the water-rock interactions, which generate steep redox, pH and chemical gradients between the interior and exterior of the icy planetesimals.



### PROTOCELL WITH **MEMBRANELESS** "ORGANELLES" FORMED **BY LIQUID-LIQUID** PHASE SEPARATION

#### Hairong Jing, Qingwen Bai, Ya'nan Lin, Haojing Chang, Dehai Liang

College of Chemistry and Molecular Engineering, Peking University, Beijing, 100871, China

Construction of protocell with hierarchical structure and living functions by prebiotically plausible components and mechanism is still a great challenge. Growing evidences demonstrate that the membraneless organelles, which facilitate many essential cellular process, are formed by RNA, protein and other biopolymers via liquid-liquid phase separation (LLPS). The formation of protocell on the early Earth could follow the same principle. In this work, we develop a novel coacervatebased protocell containing membraneless subcompartments via spontaneous liquid-liquid phase separation simply by mixing singlestranded oligonucleotides (ss-oligo), quaternized dextran (Q-dextran), and poly(L-lysine) (PLL) together. The resulting biphasic droplet, with PLL/ss-oligo phase being the interior subcompartments and Q-dextran/ss-oligo phase as the surrounding medium, is capable of sequestering and partitioning biomolecules into distinct regions. When the droplet is exposed to salt or dextranase, the surrounding Q-dextran/ ss-oligo phase experiences a dynamic adjustment and dissociates, which induce the internal subcompartments to chaotically move and fuse. Besides, the external electric field at lower strength can drive the biphasic droplet to undergo a deviated circulation concomitant with the fusion of localized subcompartments, while high-strength electric field can induce the directional movement of subcompartments and polarize the whole droplet, resulting in the release of daughter droplets in a controllable manner. Our study highlights that liquid-liquid phase separation of biopolymers is a powerful strategy to construct hierarchically structured protocells resembling the morphology and functions of living cells, and provides a step towards a better understanding of the transition mechanism from non-living to living matter under prebiotic conditions.



Figure 1 : Schematic presentation of the template assisted peptide replication.

### EMERGENCE OF NATIVE PEPTIDE SEQUENCES IN THE CONTEXT OF ORIGINS OF LIFE

#### Jayanta Nanda<sup>1,2</sup> and Gonen Ashkenasy<sup>1</sup>

<sup>1</sup> Department of Chemistry, Ben-Gurion University of the Negev, Beer-Sheva, Israel

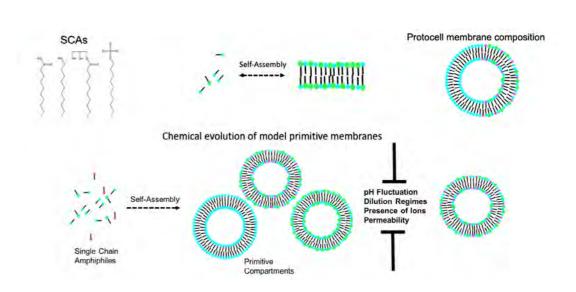
<sup>2</sup>Department of Chemistry, Indian Institute of Engineering Science and Technology, Shibpur, Howrah, India

Biopolymer syntheses in living cells are perfected by an elaborate error correction machinery, which was not applicable during polymerization on early Earth. Scientists are consequently striving to identify mechanisms by which functional polymers were selected and further amplified from complex prebiotic mixtures. Here we show the instrumental role of non-enzymatic replication in the enrichment of certain product(s). To this end, we analyzed a complex web of reactions in  $\beta$ -sheet peptide networks, focusing on the formation of specific intermediate compounds and template-assisted replication<sup>1,2</sup>. Remarkably, we find that the formation of several products in a mixture is not critically harmful, since efficient and selective templateassisted reactions serve as a backbone correction mechanism, namely, for keeping the concentration of the peptide containing the native backbone equal to, or even higher than, the concentrations of the other products. We suggest that these findings may shed light on molecular evolution processes that led to current biology<sup>3</sup>.

1 Rubinov, B.; Wagner, N.; Matmor, M.; Regev, O.; Ashkenasy, N.; Ashkenasy, G. ACS Nano 2012, 6.7893.

2 Rubinov, B.; Wagner, N.; Rapaport, H.; Ashkenasy, G. Angewandte Chemie International Edition 2009. 48. 6683.

3 Nanda, J.; Rubinov, B.; Ivnitski, D.; Mukherjee, R.; Shtelman, E.; Motro, Y.; Miller, Y.; Wagner, N.; Cohen-Luria, R.; Ashkenasy, G. Nature Communications 2017, 8

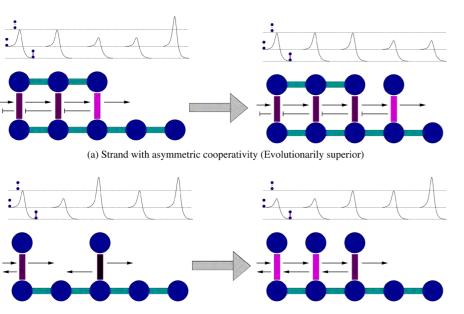


THE INFLUENCE OF COMPOSITIONAL HETEROGENEITY OF MODEL PROTOCELLULAR MEMBRANES: IMPLICATIONS FOR THE EMERGENCE OF **CELLULAR LIFE** 



Verma, Sudha Rajamani Department of Biology, Indian Institute of Science Education and Research, Pune 411008 India

Protocell membranes are thought to be comprised of mixtures of single chain amphiphiles, such as fatty acids long-chain alcohols, etc. which would have been part of the complex prebiotic chemical space. The physico-chemical properties of these prebiological membranes would have been significantly affected, and regulated, by the physical environment that they were present in. In this study, the physicochemical properties of two different chain length membrane systems i.e. C18 and C11, were systematically characterized, under prebiotically pertinent environmental conditions. The membrane systems have been designed to be composed of fatty acid and/or their corresponding alcohol and their glycerol monoester derivatives, to make a range of membrane combinations (e.g. binary and tertiary systems). The properties of those membranes were evaluated as a function of multiple factors including their composition, stability under alkaline pH, in the presence of Mg<sup>2+</sup> ions, dilution regimes. The permeability of C11 based mixed membrane systems were also investigated. These environmental constraints would have acted as important prebiotic selection pressures to shape the evolution of prebiological membranes. Our results demonstrate that complex membrane systems are more stable and robust to multiple selection pressures, thereby making them more suitable for supporting protocellular life. Furthermore, different fatty acid derivatives conferred varying degrees of stability when mixed with their respective fatty acid moiety. Importantly, the aforesaid depended on the chain length of the system, and the selection pressure that was applied. Significantly, when the systems were subjected to multiple selection pressures in a consecutive manner, only the heterogeneous membrane systems survived the race. These results highlight the requirement of compositional complexity, and underscore its implications for the emergence of mixed membrane systems during the dawn of life on Earth.



(b) Strand with symmetric cooperativity (Evolutionarily inferior)

**IDENTIFICATION OF** EVOLUTIONARY ADVANTAGE OF SOME FUNDAMENTAL PROPERTIES OF DNA FROM REPLICATIVE POTENTIAL MAXIMI-ZATION OF PRIMORDIAL AUTOCATALYTIC **HETEROPOLYMERS** 



Hemachander Subramanian<sup>1,2</sup> and Robert A. Gatenby<sup>2</sup> Department of Physics, NIT Durgapur, West Bengal, India <sup>2</sup> Integrated Mathematical Oncology, H.

Lee Moffitt Cancer Center and Research Institute, Tampa, FL, U.S.A

DNA in all living systems shares common properties that are remarkably well suited to its function, suggesting refinement by evolution. However, DNA also shares some counter-intuitive properties that confer no obvious benefit, such as strand directionality and antiparallel strand orientation, which together result in the complicated lagging strand replication. The evolutionary dynamics that led to these properties of DNA remain unknown. By carefully examining the physico-chemical requirements for evolutionarily successful primordial self-replicators, we theoretically show that asymmetric uni-directional self-replicators would have an evolutionary advantage over bidirectional self-replicators. The competing requirements of low and high kinetic barriers for induction and retention of monomers respectively are simultaneously satisfied through asymmetric kinetic influence of interstrand bonds, resulting in evolutionarily successful unidirectional self-replicators. The advantage of anti-parallel strand orientation stems from the increased rate of replication, achieved by dividing the DNA into predictable, independently and simultaneously replicating segments, as opposed to sequentially replicating the entire DNA, thereby parallelizing the replication process.

### FLOOR 3

Flow

ABIOTIC ACCUMULATION OF SMALL MOLECULES AND IONS INTO CELLULAR COMPARTMENT AGAINST A CONCENTRATION GRADIENT

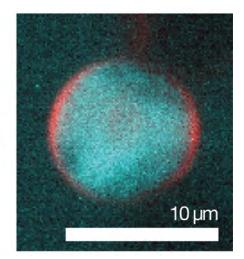


Hironori Sugiyama<sup>1</sup>, Toshihisa Osaki<sup>2,3</sup>, Shoji Takeuchi<sup>2,4</sup>, Taro Toyota<sup>1,5</sup> Department of Basic Science, Graduate School of Arts and Sciences, The University of Tokyo <sup>2</sup> Institute of Industrial Science, The University of Tokyo <sup>3</sup>Kanagawa Institute of Industrial Science and Technology <sup>4</sup>Department of Mechano-Informatics, Graduate School of Information Science and Technology, The University of Tokyo <sup>5</sup>Universal Biology Institute, The

University of Tokyo

origins of life. 119-145.

hemachander@gmail.com GO Weblink to the Poster Room

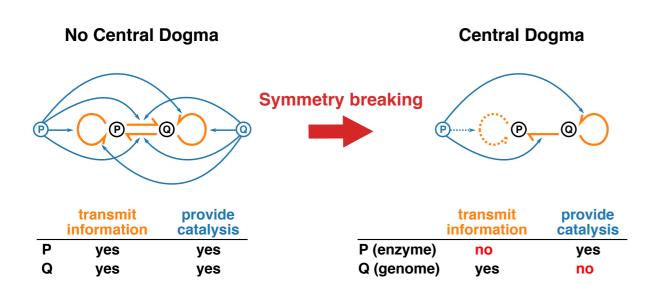


At the emergence of cell-like chemical systems at the origin of life, compartment is one of indispensable elements<sup>1</sup>. On one hand, compartmentalization is advantageous for the stable self-replication cycle against the parasitic molecules<sup>2,3</sup>. On the other hand, low permeability of phospholipid membrane hinders repeatable and continuous supply of molecules and ions inside. An abiotic process to accumulate substances into preformed liposomes against the concentration gradient would be prerequisite for maintenance, propagation, and development of metabolic system. In this study, we present that without proteins, cell-sized liposomes under hydrodynamic environment repeatedly accumulate small molecules and ions even against the concentration gradient. Notably, an analogue of adeno-sine triphosphate was also accumulated. We investigated this intri-guing and unexplored class of transportation of substrates across the lipid membrane with in-house-developed automated observation platform based on the microfluidic device (termed as MANSIONs). We discuss that the mechanism of this accumulation is probably ex-plained by a unique partitioning at the liposomal membrane exposed to the external flow in a constrained space. The hydrodynamic ac-cumulation could provide a breakthrough step for driving a metabo-lic pathway at the

1 Segré et. al., The lipid world, Origins of life and evolution of the biosphere 2001, 31 (1-2).

2 Ichihashi et.al., Darwinian evolution in a translation-coupled RNA replication system within a cell-like compartment. Nat. Commun. 2013, 4, 2494,

3 Matsumura et. al., Transient compartmentalization of RNA replicators prevents extinction due to parasites. Science 2016, 354 (6317), 1293-1296



### THE ORIGIN OF THE CENTRAL DOGMA THROUGH CONFLICTING MULTILEVEL SELECTION



#### Nobuto Takeuchi<sup>1,3</sup> and Kaneko Kunihiko<sup>2,3</sup>

School of Biological Sciences, the University of Auckland, Private Bag 92019, 1142 Auckland, New Zealand; <sup>2</sup> Graduate School of Arts and Sciences. the University of Tokyo, Komaba 3-8-1, Meguro-ku, Tokyo 153-8902, Japan; <sup>3</sup> Research Center for Complex Systems Biology, Universal Biology Institute, the University of Tokyo, Komaba 3-8-1, Meguro-ku, Tokyo 153-8902, Japan

The central dogma of molecular biology rests on two kinds of asymmetry between genomes and enzymes: informatic asymmetry, where information flows from genomes to enzymes but not from enzymes to genomes; and catalytic asymmetry, where enzymes provide chemical catalysis but genomes do not. How did these asymmetries originate? Here we show that these asymmetries can spontaneously arise from conflict between selection at the molecular level and selection at the cellular level<sup>1</sup>. We developed a model consisting of a population of protocells, each containing a population of replicating catalytic molecules. The molecules are assumed to face a trade-off between serving as catalysts and serving as templates. This trade-off causes conflicting multilevel selection: serving as catalysts is favoured by selection between protocells, whereas serving as templates is favoured by selection between molecules within protocells. This conflict induces informatic and catalytic symmetry breaking, whereby the molecules differentiate into genomes and enzymes, establishing the central dogma. We show mathematically that the symmetry breaking is caused by a positive feedback between Fisher's reproductive values and the relative impact of selection at different levels. This feedback induces a division of labour between genomes and enzymes, provided variation at the molecular level is sufficiently large relative to variation at the cellular level, a condition that is expected to hinder the evolution of altruism. Taken together, our results suggest that the central dogma is a logical consequence of conflicting multilevel selection.

1 Takeuchi and Kaneko 2019 The origin of the central dogma through conflicting multilevel selection. Proc. R. Soc. B. 286 20191359 link

Aligns with meeting schedule

62	ZHANAR ABIL Engineering evolvability of self-replicating L
63	JAIME AGUDO-CANALEJO Cooperatively enhanced reactivity and 'stabil
64	GIACOMO BARTOLUCCI Selection via phase separation
65	ALEX BLOKHUIS Autocatalysis in chemical networks: unifica
66	CLAUDIA BONFIO Towards the emergence of modern cell memb
67	JOANNA BRAU Thermal stability of metalorganic compounds
68	FEDERICO CAIMI Self-assembly driven polymerization of activa
69	ENRICO SANDRO COLIZZI Emergence of novelty and evolutionary tran
70	ÖMER KÜRSAT COSKUN & THOMAS STEINE Tracing primordial metabolism reflected by m
71	ANDRES DE LA ESCOSURA Biohybrid materials and systems chemistry
72	<b>CLAUDIA DE MICCO</b> Anomalous fluctuations and selective extinct origin of biological chirality



### INDEX - FLOOR 4

DNA as a tool towards building a synthetic cell

itaxis' of dissociating oligomeric proteins

tions and extensions

branes

s on volcanic olivine

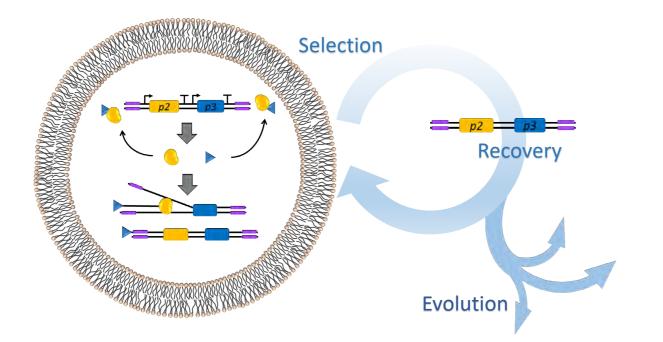
ated nucleotides

sitions

ER

nicroorganisms under hydrothermal conditions

ion in primordial replicators: a "struggle for life" at the



### ENGINEERING EVOLVABILITY OF SELF-**REPLICATING DNA AS A** TOOL TOWARDS BUILDING A SYNTHETIC CELL

#### Zhanar Abil, Christophe Danelon

Department of Bionanoscience, Kavli Institute of Nanoscience, Delft University of Technology, van der Maasweg 9, Delft 2629 HZ, The Netherlands

A consensus is that chemical evolution preceded biological evolution, and was the main driver behind the emergence of life. Our vision is to build a synthetic cell model via evolution so that we better understand life's properties and how life originated via chemical evolution. Our goal is not to determine the most plausible evolutionary path for emergence of terrain life, but to understand broader evolutionary principles and processes that can lead to emergence of self-replicating, autonomous, functionally integrated entities. To achieve this vision, we are developing a platform for a system's level continuous in vitro evolution of a synthetic cell model. We will start by evolving a simple system composed of self-replicating DNA encoding for a viral DNA polymerase and its complementary terminal protein in liposomes. Herein, we present the work done to enable long-term directed evolution of this bi-cistronic DNA fragment. As a proof of concept, we demonstrated that single molecules of DNA can be encapsulated in a polydisperse population of giant unilamellar vesicles, and that in vitro expressed replication proteins can amplify the parental linear DNA. We also show that enrichment of self-replicating DNA can be observed from a mixture with non-self-amplifying DNA, which suggests that in-liposome evolution of a DNA polymerase is possible. Moreover, we show that engineered exonuclease-deficient DNA polymerase variants are functionally active and may be potentially used as mutator DNA polymerases in a continuous in vitro evolution scenario. This work constitutes a major milestone towards in vitro evolution of a selfreplicating DNA genome in a synthetic cell model.

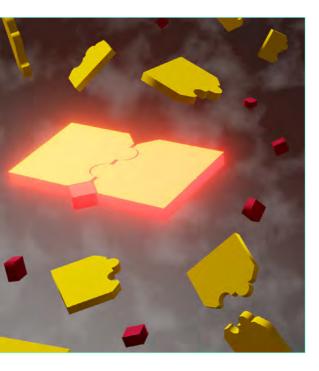
# COOPERATIVELY ENHANCED REACTIVITY AND 'STABILITAXIS' OF DISSOCIATING **OLIGOMERIC PROTEINS**



J. Agudo-Canalejo<sup>1</sup>, P. Illien<sup>2</sup>, and R. Golestanian<sup>1,2</sup>

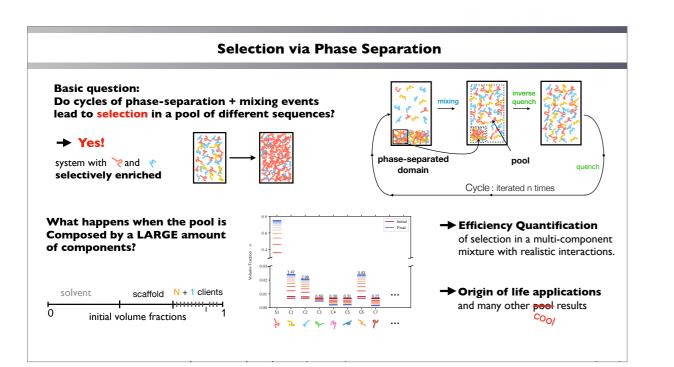
Max Planck Institute for Dynamics and Self-Organization (MPIDS), D-37077 Göttingen, Germany <sup>2</sup> Sorbonne Université, CNRS Laboratoire PHENIX, UMR CNRS 8234, 75005 Paris, France <sup>3</sup> Rudolf Peierls Centre for Theoretical Physics, University of Oxford, Oxford

OX1 3PU, UK



Many functional units in biology, such as enzymes or molecular motors, are composed of several subunits that can reversibly assemble and disassemble. This includes oligomeric proteins composed of several smaller monomers, as well as protein complexes assembled from a few proteins. By studying the generic spatial transport properties of such proteins, we investigate here whether their ability to reversibly associate and dissociate may confer them a functional advantage with respect to non-dissociating proteins. In uniform environments with positionindependent association-dissociation, we find that enhanced diffusion in the monomeric state coupled to reassociation into the functional oligomeric form leads to enhanced reactivity with distant targets. In non-uniform environments with position-dependent associationdissociation, caused e.g. by spatial gradients of an inhibiting chemical, we find that dissociating proteins generically tend to accumulate in regions where they are most stable, a process that we term 'stabilitaxis'.

J. Agudo-Canalejo, P. Illien, and R. Golestanian, Proc. Natl. Acad. Sci. U.S.A. (2020, in press). arXiv:1911.02350



### **SELECTION VIA PHASE SEPARATION**



Giacomo Bartolucci<sup>1,2</sup>, Yash Rana<sup>1,2</sup>, Alexandra Kühnlein<sup>3</sup>. Christof Mast<sup>3</sup>. Dieter Braun<sup>3</sup>, and Christoph A. Weber<sup>1,2</sup> Max Planck for the Physics of Complex Systems, Dresden

<sup>2</sup>Center for Systems Biology, Dresden <sup>3</sup>Ludwig Maximilian University, MünchenInstitute of Nanoscience, Delft University of Technology, van der Maasweg 9, Delft 2629 HZ, The Netherlands

Living cells and pre-biotic systems are complex aqueous mixtures composed of thousands of different heteropolymers. In such multicomponent mixtures, enrichment and selection of a small set of components is important to achieve biological function. However, when the number of components increases, components are more diluted impeding a significant enrichment of selected components. Here, we propose a selection mechanism relevant for prebiotic mixtures based on cycles of phase separation combined with material exchange of the dense phase with a reservoir. We find a selective enrichment of components up to two orders of magnitude coinciding with a growth of the dense phase up to the system volume. Such enrichment of selective components is robust also in mixtures composed of a large number of components. For a prebiotic soup, our findings indicate that cycles of phase separation and material exchange with a reservoir, e.g. the accumulation DNA gel in rock pores periodically filled with DNA rich aqueous solution<sup>1</sup>, could provide a mechanism for the selection and enrichment of specific heteropolymers sequences in a multi-component mixture at the origin of life.

1 M. Morasch, A. Kühnlein, Christof Mast, Dieter Braun et al. Nature Chemistry 2019

### Type III Type IV Type V Type I Type II **○**+**■\$■**+**○\$○**+**■** 0 **+** 1 reaction, 1 Reaction, 3 compartments: 1 compartment: Autocatalysis allowed (type III) Autocatalysis excluded Branching process Extinction probabilities

### **AUTOCATALYSIS IN** CHEMICAL NETWORKS: **UNIFICATIONS AND** EXTENSIONS

Netherlands

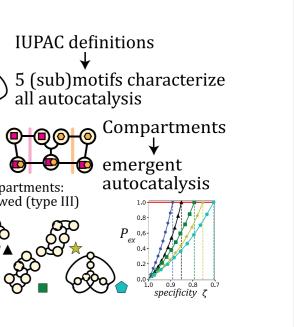


#### Alex Blokhuis<sup>1,2,3,4</sup>, David Lacoste<sup>1</sup>, Philippe Nahe<sup>2</sup>

Gulliver Laboratory, UMR CNRS 7083, PSL University, 10 rue Vauquelin, Paris F-75231. France

<sup>2</sup> Laboratoire de Biochimie, UMR CNRS 8231 Chimie Biologie et Innovation, ESPCI Paris, PSL University, 10 rue Vauquelin, Paris, France <sup>3</sup> Centre for Systems Chemistry, Stratingh Institute, University of Groningen, Nijenborgh 4, 9747 AG Groningen, The Netherlands <sup>4</sup> Groningen Institute for Evolutionary Life Sciences, University of Groningen, Nijenborgh 7, 9747 AG Groningen, The

fied. be widespread.



Autocatalysis is a multifarious phenomenon encountered in all branches of chemistry, and a shared feature in all scenarios for abiogenesis. While formalizations exist for particular instances, autocatalysis in chemistry lacks a unifying framework that captures all reported autocatalysis, and unknown forms of autocatalysis may still have to be identi-

Here, we introduce this framework, by deriving general properties in chemical reaction networks for catalysis and autocatalysis that follow directly from basic definitions in chemistry. These definitions imply minimal structural motifs in autocatalytic networks which come in five types, of which two have not yet been encountered. Branching processes allow to assess the kinetic viability of networks, which depends on such motifs. We further extend the range of conceivable autocatalysis by showing that autocatalytic motifs readily emerge in a multicompartment setting. Autocatalysis can thereby be realized from a single uncatalyzed chemical reaction, suggesting the phenomenon may in fact

Preprint, https://doi.org/10.26434/chemrxiv.12317273.v1

Blokhuis, Alex (2019): Physical Aspects of Origins of Life Scenarios, Thesis, PSL Research University, ESPCI. https://hal.archives-ouvertes.fr/tel-02566386/

Blokhuis, Alex; Lacoste, David; Nghe, Philippe (2020): Autocatalysis in Chemical Networks: Unifications and Extensions, ChemBxiv

THERMAL STABILITY OF

**METALORGANIC** 

COMPOUNDS ON

**VOLCANIC OLIVINE** 

Bettina Scheu<sup>1</sup>, and Donald B. Dingwell<sup>1</sup>

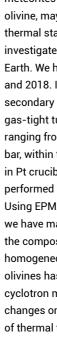
Joanna Brau<sup>1</sup>, Marco Matzka<sup>2</sup>, Philippe Schmitt-

Kopplin<sup>2,3</sup>, Norbert Hertkorn<sup>2</sup>, Werner Ertel-Ingrisch<sup>1</sup>,

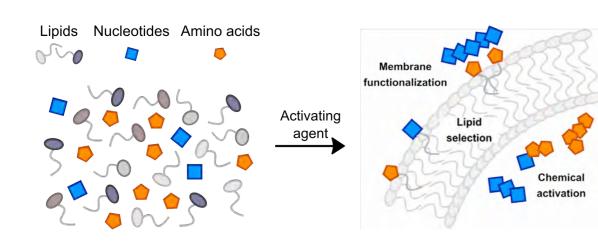
<sup>1</sup>Ludwig-Maximilians-Universität München, Germany;

<sup>2</sup> Helmholtz Zentrum München, Neuherberg, Germany;

<sup>3</sup> Technische Universität München, Freising, Germany



2824



### TOWARDS THE EMERGENCE OF MODERN CELL MEMBRANES



C. Bonfio, J.D. Sutherland MRC Laboratory of Molecular Biology -Cambridge (United Kingdom)

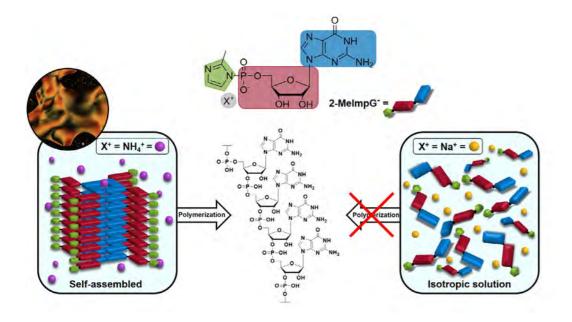
The complexity of modern biochemistry suggests that a systems chemistry approach is required to understand and potentially recapitulate the intricate network of prebiotic reactions that led to theemergence of life. Early cells probably relied upon compatible and interconnected chemistries to linkRNA, peptides and membranes. In this context, understanding how and when phospholipid membranesappeared on early Earth is critical to elucidating the prebiotic pathways that led to the emergence of primitive cells. Starting with a mixture of activated carboxylic acids of different lengths, iterative cyclingof acylation and hydrolysis steps allowed for the selection of longer-chain acylglycerol-phosphatesthrough accumulation-induced compartmentalization of self-assembling amphiphiles at the expense of non-self-assembling shorter chain analogues. Our results suggest that a selection pathway based onenergy-dissipative cycling could have driven the selectivesynthesis of phospholipids on the early Earth<sup>1</sup>. Moreover, I will show that several types of vesicles, formed from prebiotically plausiblemixtures of amphiphiles, allow activation of amino acids, peptides and nucleotides. Interestingly, activation chemistry drives the advantageous conversion of reactive amphiphiles into inertcyclophospholipids, thus supporting their potential role as major constituents of primitive cells.Activation of prebiotic building blocks within fatty acid-based vesicles yields lipidated species capable of localizing and functionalizing primitive membranes. Our findings describe a potentially prebioticnetwork of reactions in which the components of primitive cells could have selectively undergoneactivation and reacted to yield new species, which enabled the emergence of cells with increasinglyadvanced functionalities<sup>2</sup>.

1 Bonfio C. et al., J. Am. Chem. Soc.2019, 141, 3934-3939 2 Bonfio C. et al., submitted

Previously unknown class of metalorganic compounds revealed in meteorites<sup>1</sup> also found on the surfaces of silicate phases such as olivine, may have been involved in the emergence of life. Here, the thermal stability of such organic compounds has been experimentally investigated under conditions which simulate those extant on the early Earth. We have studied olivines from the Hawaiian eruptions of 1959 and 2018. Individual mineral grains have been hand-picked to be free of secondary phases such as pyroxene or melt. We use a high temperature gas-tight tube furnace under CO-CO<sub>2</sub> gas mixture at temperatures ranging from 950°C to 1350°C and oxygen fugacity ranging from 10-12 bar, within the stability field of olivine. The samples were contained in Pt crucibles and held for dwell times of 1 to 8 h. Quenching was performed by lifting the samples vertically out of the tube furnace. Using EPMA (electron microprobe analyzer) and RAMAN spectroscopy, we have mapped the state of the olivine samples. We observe that the composition of the individual mineral grains remains stable and homogeneous with thermal treatment. The metalorganic cargo of these olivines has been analyzed using FT-ICR- MS (Fourier Transform ion cyclotron mass spectrometry). Preliminary results reveal systematic changes or organic molecular composition depending on time and heat of thermal treatment whose origins will be discussed...

1 A. Ruf, B. Kanawati, N. Hertkorn, Q. Yin, F. Moritz, M. Harir, M. Lucio, B. Michalke, J. Wimpenny, S. Shilobreeva, B. Bronsky, V. Saraykin, Z. Gabelica, R. D. Gougeon, E. Quirico, S. Ralew, T. Jakubowski, H. Haack, M. Gonsior, P. Jenniskens, N. W. Hinman, P. Schmitt-Kopplin. Previously unknown class of metalorganic compounds revealed in meteorites. PNAS 114 (2017) 2819-





### SELF-ASSEMBLY DRIVEN POLYMERIZATION OF ACTIVATED NUCLEOTIDES



Federico Caimi<sup>1</sup>, Marco Todisco<sup>1</sup>, Diego Colombo<sup>1</sup>, Sara Sattin<sup>2</sup>, Tommaso **Bellini**<sup>1</sup>

<sup>1</sup> Dipartimento di Biotecnologie Mediche e Medicina Traslazionale, Università degli Studi di Milano, via Vanvitelli 32, 20129 Milano, Italy <sup>2</sup> Dipartimento di Chimica, Università degli Studi di Milano, via Golgi 19, 20133 Milano, Italy

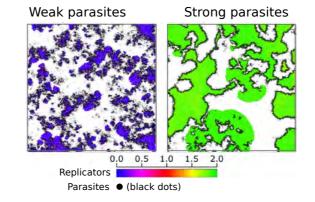
One of the most mind-bending problems about the "RNA World" hypothesis is the lack of a process to explain the ligation of monomeric nucleotides into linear chains.

Many mechanisms for the abiotic polymerization have been proposed, but no one without issues. Among them phosphoroimidazolides, carrying the energy required to form phosphodiester bonds, can perform the nonenzymatic copying of RNA templates.1 However, this process relies on the presence of a preformed RNA chain to drive the polymerization, favoring the reactivity and preventing the formation of cyclic products. It has been recently discovered that nucleic acids, at high concentration, can self-assemble into linear aggregates<sup>2</sup>, that in turn enhance their chance of ligating when in the presence of condensing agents like carbodiimides.<sup>3</sup> Here we report that imidazole-activated guanosines can self-assemble forming G-quadruplex columns and order into nematic liquid crystal phases, analogously to guanosine monophosphate. These columns are formed by stacked tetrads of Hoogsteen-paired guanines, which resemble the secondary structures found in guanine-rich DNA and RNA strands. We find that the organization provided by these structures drives, without the need of templating strands, the polymerization of phosphoroimidazolides into oligos limited in length by the peculiar reaction mechanism of these molecules. With a simple simulation we find that in a static structure the polymerization can run into some breakpoints hampering the formation of longer oligos. To introduce some dynamics to the system we designed some wet-dry cycles adding fresh activated nucleotide to each cycle increasing the polymerization up to decamers.

By demonstrating a path of polymerization through self-assembly, our findings support the idea of a primeval world in which RNA strands emerged from pools of self-assembled molecules.

1 J. Am. Chem. Soc. 2016, 138, 11996-12002 2 PNAS 2008, 105 (4), 1111-7 3 ACS Nano 2018 12 (10), 9750-9762

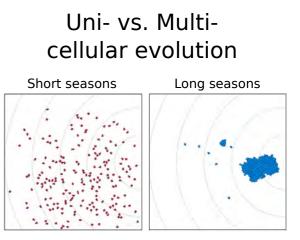
### Spatial patterns in the RNA world



### EMERGENCE OF NOVELTY AND EVOLUTIONARY TRANSITIONS



Enrico Sandro Colizzi Leiden University, Origins Center



Novel functions and traits have appeared throughout the four billion years of evolutionary history. The emergence of novelty is particularly salient during Major Evolutionary Transitions - where new levels of individuality are generated by the self-organisation of its constituting components - because entire new spaces of possibilities are opened. Evolutionary novelty is typically understood "a posteriori" by tracing back the steps that generated it.

I show that novelty can be understood - by mathematical modelling also without presuming the result. I present two very different models, one about the Origin of Life and the other about the Evolution of Multicellular Life, and show that novelty arises in a similar way. In the Origin of Life model, RNA-like replicators can evolve the degree to which they replicate one another. In the Origin of Multicellularity model, cells follow a noisy signal that lead to resources, and can evolve the degree to which they stick to each other. In both models, the self-organisation of replicators (RNAs or cells) into higher-level individuals (RNA replicator-parasite waves or multicellular organisms) create functional properties that did not exist outside of the higherlevel context (hence novel) and that profoundly affect the evolutionary dynamics of the replicators.

In conclusion, the interplay between self-organisation and evolution readily generates novelty.

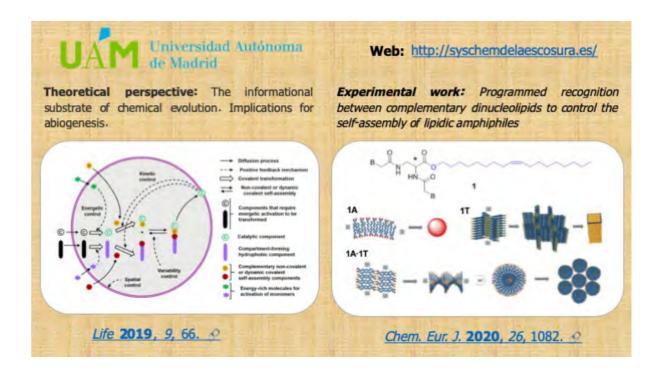
### TRACING PRIMORDIAL METABOLISM REFLECTED BY MICROORGANISMS UNDER HYDROTHERMAL CONDITIONS

#### Ömer K. Coskun<sup>1</sup>, Thomas Steiner<sup>2</sup>, William Orsi<sup>1,3</sup>, Wolfgang Eisenreich<sup>2</sup>

<sup>1</sup> Department of Earth and Environmental Sciences, Ludwig-Maximilians Universität, 80333 Munich, Germany <sup>2</sup> Chair of Biochemistry, Department of Chemistry, Technische Universität München, Garching, Germany <sup>3</sup> GeoBio-CenterLMU, Ludwig-Maximilians Universität, 80333 Munich, Germany

The origin of life is hypothesized to be linked to autocatalytic carbon fixation pathways under primordial hydrothermal conditions<sup>1</sup>. The main goal of this project is to provide experimental, field, and theoretical constraints on the functioning and evolution of carbon fixation pathways while combining microbiology and chemistry. Using isotopologue profiling analysis, we investigate carbon fixation in different heterotropic and autotrophic model organisms, namely Bacillus subtilis<sup>2</sup>, Hippea maritima, Pyrobaculum arsenaticum and Raoultella planticola. These data will be compared to microbial communities expressing the rTCA and reductive acetyl-CoA pathways sampled from Loki's castle in collaboration with Bergen University, active volcanism driven hydrothermal area in Milos (in collaboration with Bergen University) and geothermal springs/wells in Eastern Anatolia in Turkey (in collaboration with Istanbul University). Twenty good to high quality metagenomic bins were constructed from samples taken from the Turkey and three bins affiliated to deeply branching hydrogen oxidizing Aguificae were found. Further evaluation on metagenomic bins, future fieldworks and metatransciptomic studies of hydrothermal samples will validate the experimental results under natural conditions, by assessing how natural hydrothermal settings can favor the expression of either pathway or pathway components. Our project will result in novel insights into early metabolic evolution under hydrothermal conditions.

1 Fuchs, G. Alternative pathways of carbon dioxide fixation: insights into the early evolution of life? Annu Rev Microbiol 65. 631-658, doi:10.1146/annurev-micro-090110-102801 (2011). 2 Spona-Friedl M. Substrate-dependent CO2-fixation in heterotrophic bacteria revealed by stable isotope labelling FEMS Microbiol Ecol 96, fiaa080, doi: 10.1093/femsec/fiaa080 (2020).



### **BIOHYBRID MATERIALS** AND SYSTEMS CHEMISTRY

Andres de la Escosura<sup>1,2</sup>, Sonia Vela-Gallego<sup>1</sup>, Stefania Kalantzi<sup>1</sup>, Noemi Nogal-Rodriguez<sup>1</sup>, Santiago Guisan<sup>1</sup> <sup>1</sup> Departament of Organic Chemistry, Universidad Autónoma de Madrid, Campus de Cantoblanco 28049, Madrid, Spain.

<sup>2</sup> Institute for Advanced Research in Chemistry (IAdChem) Campus de Cantoblanco 28049 Madrid Spain

lines.

Abiogenesis", Life, 9, 66, Chem. Soc. Rev., 47, 19. 170050.

The study of complex molecular networks and supramolecular assemblies is a clear objective of the field so-called systems chemistry, which is expected to have a great impact in the area of origins-of-life research and as biohybrid materials in materials science. With regards to the origins of life, a pertinent question is whether artificial cells could be constructed from non-natural components. In order to provide clues about this question, we have started research lines on nucleic acid hybrids and nucleolipid compartments. The study and combination of these components is an interesting approach because it allows exploring some properties of life without the restrictions of the historical pathway that Darwinian evolution took. Concerning the approach to biohybrid materials, we focus our work on supramolecular biohybrids for biomedical light management, which combine different photoactive molecules with peptide, protein and nucleic acid nanostructures. In this poster presentation we will shortly discuss some of these research

S. Morales-Reina, C. Giri, M. Leclerco, S. Vela-Gallego, I. de la Torre, J. R. Caston, M. Surin, A. de la Escosura, 2020. "Programmed Recognition Between Complementary Dinucleolipids to Control the Self-Assembly of Lipidic Amphiphiles", Chem. Eur. J., 26, 1082.

A. de la Escosura, 2019. "The Informational Substrate of Chemical Evolution: Impli-cations for

V. Almeida-Marrero, E. van de Winckle, E. Anaya-Plaza, T. Torres, A. de la Escosura, 2018. "Porphyrinoid Biohybrid Materials as an Emerging Toolbox for Biomedical Light Management",

K. Ruiz-Mirazo, C. Briones, A. de la Escosura, 2017. "Chemical Roots of Biological Evolution: The Origins of Life as a Process of Development of Autonomous Functional Systems", Open Biol. 7,

K. Ruiz-Mirazo, C. Briones, A. de la Escosura, 2014. "Prebiotic Systems Chemistry: New Perspectives for the Origins of Life", Chem. Rev. 114, 285-366.

# ANOMALOUS FLUCTUATIONS AND **SELECTIVE EXTINCTION** IN PRIMORDIAL **REPLICATORS:** A "STRUGGLE FOR LIFE" AT THE ORIGIN OF **BIOLOGICAL CHIRALITY**



Savino Longo<sup>1,3</sup>, Claudia De Micco<sup>2</sup>, Miriana Carmela Chincoli<sup>1</sup>, Gaia Micca Longo<sup>1,3</sup>

Department of Chemistry – Università degli Studi di Bari Aldo Moro – Via Orabona 4 – 70125 Bari, Italy <sup>2</sup> Department of Bioscience, Biotechnology and Biopharmaceutics -Università degli Studi di Bari Aldo Moro – Via Orabona 4 – 70125 Bari, Italy <sup>3</sup> Istituto per la Scienza e Tecnologia dei Plasmi – Consiglio Nazionale delle Ricerche, Bari Section – Via Amendola 122/D – 70125 Bari, Italy

One of the most distinctive signs of life as we know it, is the presence of a single chiral variant in living organisms and one of the greatest ambitions for biochemistry and astrobiology is to provide an explanation of this predominance. Several mechanisms were proposed in the past, here we propose a different scenario: anomalous fluctuations associated with a self-replication process can lead to selective extinction of one of the two variants. The idea is based on three key-points: a) the simulation of early biological processes as a "board game"; b) the presence of large fluctuations during an autocatalytic process; c) the presence of a limited source of chemical energy, inducing a form of competition in a primordial replicator population. In order to demonstrate this mechanism, a computational model is developed, describing the "struggle for life" of two different kinds of primordial replicators on a "chessboard" with periodic boundary conditions. Each replicator employs enzymes of different chirality on a non-chiral substrate. The replication occurs randomly and with a fixed probability, providing that a sufficient amount of chemical energy is locally available. Results clearly show that strong fluctuations in the number of individuals of each species and a subsequent selective extinction of one of the two are observed. In the next step of our research, a more structured variant of the mechanism will be considered. This will concern experimental simulations with more complicated assumptions. More complex organisms and biological behavior will be investigated. Phenotypes, potentially favorable in the "struggle for life" will be introduced. Phenotypes are acquired on the basis of a genotype; therefore, our model will be implemented with a genetic code which could be inherited or even mutated producing different phenotypes. These studies may contribute to shed light on the transition occurred during the biochemical evolution of our planet.

Longo, S. (2019). Anomalous fluctuations and selective extinction in populations of primordial, EANA 2019, Orléans

# INDEX - FLOOR 5

74	CHRISTINA FELICITAS DIRSCHERL
	Escalation of DNA monomer polymerization i
75	CARSTEN DONAU
	Active coacervate droplets as a platform towar
76	OLIVIA DOPPLEB
	Oligonucleotide assemblies as early ribosomes
77	JAN-PETER DUDA
	Tracking life at the break of dawn—Identificat oldest rocks
78	SELENE FORGET & MARKUS MERINGER
	Computational exploration of lipid chemical sp
79	TOMMASO FRACCIA
	Liquid crystal coacervates: a pathway for biopo structuring
80	TARO FURUBAYASHI
	Emergence and diversification of a host-parasi
81	THOMAS GEISBERGER
	Chemical evolution of biomolecules formed u
82	EFTAL GEZER
	Selection pressure: the latitudinal biodiversity g
83	TOBIAS GOEPPEL
	A nonequilibrium error filtering mechanism for
84	MAREN HAAS
	A mineral-catalysed, mechanochemical formos



Aligns with meeting schedule

in thermal traps

ds synthetic life

tion & interpretation of biosignatures in Earth's

ace : Predicting assembly using QSPR models plymers coevolution and complex proto-cellular

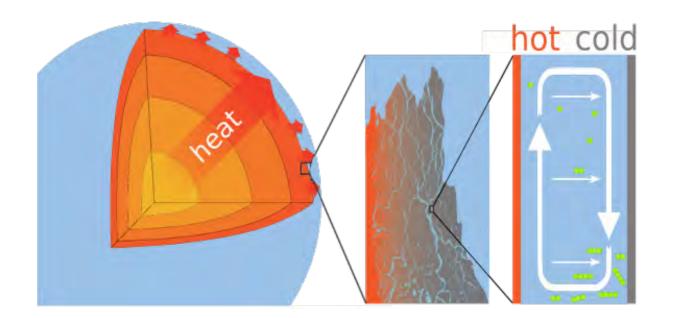
te RNA ecosystem through Darwinian evolution

Inder volcanic hydrothermal conditions

gradient is driven by the latitudinal variation of

enzyme-free copying of nucleic acid sequences

se reaction



# ESCALATION OF DNA MONOMER POLYMERIZATION IN THERMAL TRAPS



Christina Felicitas Dirscherl, Dieter Braun

Systems Biophysics, Functional Nanosystems, Ludwig-Maximilian University Munich, Amalienstr. 54, 80799 Munich, Germany

Highly diluted starting concentrations and the 'tyranny of the shortest' are the main problems for de novo strand formation of DNA/RNA on Early Earth. A possible scenario that could have overcome these issues are thermogravitational traps in hydrothermal vent systems<sup>1</sup>. We demonstrate that these traps can accumulate solutions of single molecules despite their high diffusivity. We prove that there are nonenzymatic chemistries that are designed to polymerize RNA also polymerize DNA mononucleotides. However, the concentrations of the strands decrease exponentially with length leaving the long strands to be the inferior species. In non-equilibrium conditions provided by thermogravitational traps an accumulation of DNA strands is taking place that is exponentially length dependent<sup>2</sup>. We show that the trapping procedure significantly increases the probability that longer strands are linked together and consequently leads to an escalation of DNA polymerization. If additionally, two complementary monomers are present in the system, self-templated ligation can take place after the first spontaneous formation of oligomers.

1 Thermal trap for DNA replication. C.B. Mast & D. Braun. Physical Review Letters 104,188102 (2010)

2 Escalation of polymerization in a thermal gradient. C.B. Mast, S. Schink, U. Gerland & D. Braun. PNAS 110 8030-8035 (2013)

# ACTIVE COACERVATE DROPLETS AS A PLATFORM TOWARDS SYNTHETIC LIFE



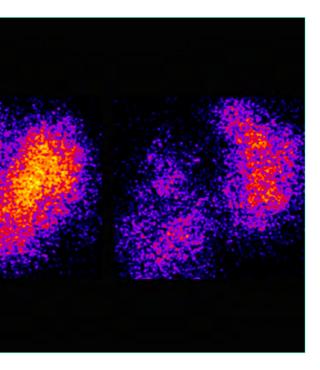
Carsten Donau<sup>1</sup>, Fabian Spaeth<sup>1</sup>, Marilyne Sosson<sup>1</sup>, Brigitte Kriebisch<sup>1</sup>, Fabian Schnitter<sup>1</sup>, Marta Tena-Solsona<sup>1,2</sup>, Hyun-Seo Kang<sup>1,3</sup>, Elia Salibi<sup>4</sup>. Michael Sattler<sup>1,3</sup>. Hannes Mutschler<sup>4</sup>. Job Boekhoven<sup>1,2</sup> <sup>1</sup> Department of Chemistry, Technical University of Munich, Lichtenbergstrasse 4, 85748 Garching, Germany <sup>2</sup> Institute for Advanced Study, Technical University of Munich, Lichtenbergstrasse 2a, 85748 Garching, Germany

<sup>3</sup> Institute of Structural Biology, Helmholtz Zentrum München.

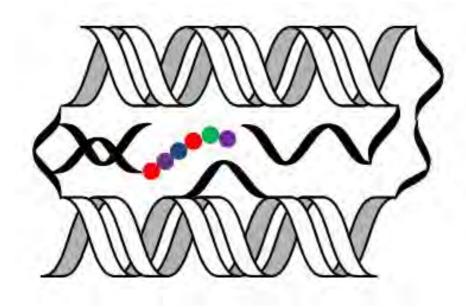
Ingolstädter Landstrasse 1, 85764 Neuherberg

<sup>4</sup> Max Planck Institute of Biochemistry, Am Klopferspitz 18, 82152 Martinsried

Membraneless organelles like stress granules are active liquid-liquid phase-separated droplets that are involved in many intracellular processes. Their active and dynamic behavior is often regulated by ATP-dependent reactions. However, how exactly membraneless organelles control their dynamic composition remains poorly understood. Herein<sup>1</sup>, we present a model for membraneless organelles based on RNA-containing active coacervate droplets regulated by a fuel-driven reaction cycle. These droplets emerge when fuel is present, but decay without. Moreover, we find these droplets can transiently up-concentrate functional RNA, and that this up-take is accelerated by the chemical reaction cycle. Finally, we show that in their pathway towards decay, these droplets self-divide asymmetri-cally. Self-division combined with emergence, decay, rapid exchange of building blocks, and functionality are all hallmarks of life, and we believe that our work could be a stepping stone towards its synthesis.



1 ChemRXiv: "Active Coacervate Droplets as a Model for Membraneless Orga-nelles and a Platform Towards Synthetic Life"



# OLIGONUCLEOTIDE ASSEMBLIES AS EARLY RIBOSOMES



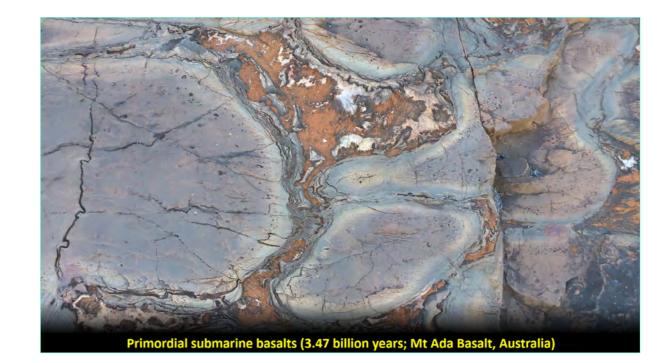
Olivia Doppleb<sup>1</sup>, Arthur Ermatov<sup>2</sup>, Biswarup Jash<sup>1</sup>, Rainer-Joachim Schwarz<sup>1</sup>, Tim Liedl<sup>2</sup>, Clemens Richert<sup>1</sup> Institute of Organic Chemistry, University of Stuttgart <sup>2</sup> Department of Physics, LMU Munich

As Francis Crick et al. wrote, "The origin of protein synthesis is a notoriously difficult problem. We do not mean by this the formation of random polypeptides but the origin of the synthesis of polypeptides directed, however crudely, by a nucleic acid template and of such a nature that it could evolve by steps into the present genetic code."1 This difficult problem is still unsolved.<sup>2</sup> The earliest organisms capable of RNA-induced peptide synthesis must have possessed a much simpler form of molecular machinery than the ribosomal apparatus. Based on the experimental observation of unencoded formation of 'peptido RNA' from amino acids and nucleotides in condensation buffer<sup>3,4</sup>, we are studying the effect of multistrand assemblies on the nanometer scale on peptide and peptido RNA formation. This includes assemblies with RNA primers that possess binding pockets for amino acids and templated reactions with tRNAs.5

In particular, we are investigating reactions in small folded DNA or RNA structures comprising 65 base pairs or less and gaps of one or two nucleotides. Such assemblies can have a profound effect on peptide chain growth in aqueous condensation buffers.

1 Crick, F. H. C.; Brenner, S.; Klug, A.; Pieczenik, G. Orig. Life 7, 389-397 (1976). 2 Yarus, M. Life 7, 13 (2017). 3 Jauker, M.; Griesser, H.; Richert, C. Angew. Chem. Int. Ed. 54, 14564-14569 (2015).

4 Griesser, H.; Tremmel, P.; Kervio, E.; Pfeffer, C.; Steiner, U.E.; Richert, C. Angew. Chem. Int. Ed. 56 1219-1223 (2017) 5 Schwarz, R.-J.; Richert, C., Nanoscale 9, 7047-7054 (2017).



TRACKING LIFE AT THE BREAK OF DAWN-**IDENTIFICATION &** INTERPRETATION OF **BIOSIGNATURES IN** EARTH'S OLDEST ROCKS



J.-P. Duda<sup>1,2</sup>, H. Mißbach<sup>1</sup>, M. Reinhardt<sup>3</sup>, V. Thiel<sup>1</sup>, & J. Reitner<sup>1,2</sup> <sup>1</sup> Georg-August-Universität Göttingen, Göttingen, Germany <sup>2</sup>Göttingen Academy of Sciences and Humanities, Göttingen, Germany <sup>3</sup> Linnaeus University, Kalmar, Sweden

When and under what conditions did life emerge? The geological record holds detailed information relevant to these questions. Particularly important are geologically stable signatures that can be used to track primordial life and, potentially, provide information on its nature (i.e., biosignatures). However, the identification of unequivocal biosignatures in Earth's oldest rocks is incredibly challenging. For instance, candidate signatures must be demonstrably syngenetic with the formation of the host rock and assuredly be of biological origin. Once established, the interpretation of biosignatures is a further challenge we face. This is because biological processes are extremely complex, and so are the resulting signatures. In this presentation, we will demonstrate how we can yet extract valuable information from the geological record by combining field observations with a variety of rock-based approaches (e.g., analytical imaging techniques, organic-geochemical analyses)<sup>1,2</sup>. Furthermore, we will show how we can use younger analogue systems (e.g., hydrothermal Lake Magadi, Kenya<sup>3</sup>) and laboratory experiments (e.g., Fischer-Tropsch-type synthesis of organic matter<sup>4</sup>) to fill remaining gaps in understanding. This integrative approach has allowed us to successfully identify and interpret unequivocal biosignatures in some of Earth's oldest rocks (e.g., 3.4-Byr-old mineralized microbial mats1, 3.5-Byr-old organic biomolecules2). Furthermore, it has provided detailed insights into the nature of early Earth's habitats. In the long run, our studies will help to develop a solid understanding of how life emerged on our planet (and possibly beyond)...

1 Duda et al. (2016) PloS One 11(1), e0147629 (10.1371/journal.pone.0147629) 2 Duda et al. (2018) Biogeosci. 15(5), 1535-1548 (10.5194/bg-15-1535-2018) 3 Reinhardt et al. (2019) Biogeosci. 16(12), 2443-2465 (10.5194/bg-16-2443-2019) 4 Mißbach et al. (2018) Org. Geochem. 119, 110–121 (10.1016/j.orggeochem.2018.02.012)

The abiotic formation of biopolymers from their monomeric building blocks and the emergence of cellular structure are still unsolved problems in the origin of life investigation. Here we report the formation of liquid crystal phases inside complex coacervate microdroplets in mixtures of short dsDNA and cationic peptides1. This system reveals a much more complex phase diagram than normal unstructured coacervates and reversibly transitions between each phase state with continuity upon variations in temperature and added salt, DNA and peptide concentration. Coacervation decreases the global dsDNA concentration required for the nucleation of all LC mesophases previously observed in bulk<sup>2</sup>. Remarkably and counterintuitively, in drying-wetting cycles, this system can escape precipitation simply by drying, and accesses ordered, yet fluid, phases by dilution of a homogeneous liquid phase. Additionally, we show that the dense coacervate phase enhances the rate of non-enzymatic ligation reactions, resulting in the elongation of constituent short dsDNA up to more than 10 times their initial length, while the reaction is ineffective in the diluted phase. The cooperative assembly of nucleic acids monomers<sup>3</sup> and oligomeric peptides templated by LC ordering inside the coacervate phase can constitute a pathway for the formation and co-evolution of life polymers. LC-coacervates can generate multi-phase compartments with different degrees of order<sup>4</sup> and permeability to guest molecules, which can result in effective and structured model protocells.

Structure Enumeration Algorithms And have were here with New Computed Amphiphiles **Computing Critical Packing Parameter** Predicting CMC with QSPR Models Experimental assay New Candidates for Compartimentalization

# COMPUTATIONAL **EXPLORATION OF LIPID** CHEMICAL SPACE : PREDICTING ASSEMBLY USING QSPR MODELS



Selene Forget, Ric Gillams, Tony Jia Markus Meringer, Jim Cleaves Ecole Normale Supérieure

Compartmentalization is likely to have been essential for the emergence of life. Compartmentalization allows for the creation of unique chemical conditions that can be maintained out of equilibrium with the environment and the exclusion of parasites. Confining organic molecules also helps limit diffusion, increases concentration and can thus influence both the thermodynamics and kinetics of prebiotic reactions. Biology currently predominantly uses phospholipids to construct cell membranes. However, there are many other types of organic compounds that can form stable compartments in water, and many of these may have been abundant in the prebiotic environment. In this study we explore this alternative lipid chemical space by using structure enumeration algorithms to compute an exhaustive combinatorial library of surfactant molecules. We then predict the propensity of these compounds to self-assemble into membranes using quantitative structure-property relationship (QSPR) models on critical micelle concentration (CMC). Combined with critical packing parameter calculations, these models can allow identification of novel molecule types which can be experimentally assayed as candidates for the emergence of protocells.

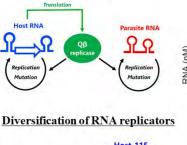
# COACERVATES: A PATHWAY FOR BIO-POLYMERS COEVOLUTION AND COMPLEX PROTO-**CELLULAR STRUCTURING**

LIQUID CRYSTAL

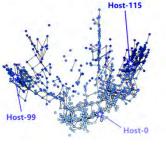
Tommaso P. Fraccia<sup>1</sup>, Nicolas Martin<sup>2</sup> and Tony Z. Jia<sup>3,4</sup> <sup>1</sup> Institut Pierre-Gilles de Gennes, CBI UMR 8231 ESPCI Paris - CNRS, PSL University, Paris, France <sup>2</sup>Centre de Recherche Paul Pascal, UMR 5031 CNRS, Bordeaux, France <sup>3</sup>Earth-Life Science Institute, Tokyo Institute of Technology, Tokyo, Japan <sup>4</sup> Blue Marble Space Institute of Science, Seattle Washington USA

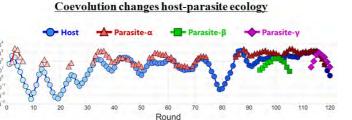
selene.forget@ens.fr , markus.meringer@dlr.de GO Weblink to the Poster Room

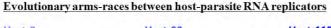
1 Fraccia, T. P., Jia, T. Z. chemRxiv (2020) doi:10.26434/chemrxiv.12220418.v2 2 Nakata, M. et al. Science 318, 1276-1279 (2007) 3 Smith, G. P. et al. Proc. Natl. Acad. Sci. U. S. A. 115, E7658-E7664 (2018) 4 Shakya, A. & King, J. T. Biophys. J. 115, 1840–1847 (2018)

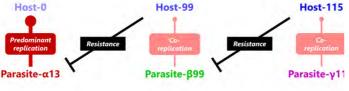


Host-parasite RNA ecosystem









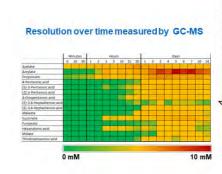
EMERGENCE AND **DIVERSIFICATION OF A** HOST-PARASITE RNA ECOSYSTEM THROUGH DARWINIAN EVOLUTION

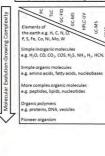


Taro Furubayashi<sup>1</sup>, Kensuke Ueda<sup>2</sup>, Yohsuke Bansho<sup>3</sup>, Daisuke Motooka<sup>4</sup>, Shota Nakamura<sup>4</sup>, Ryo Mizuuchi<sup>5,6</sup>, Norikazu Ichihashi<sup>2,3,5,7,8</sup> Laboratoire Gulliver, CNRS, ESPCI Paris,

PSL Research University, Paris, France <sup>2</sup> Department of Bioinformatic Engineering, Graduate School of Information Science and Technology, Osaka University, Osaka, Japan <sup>3</sup>Graduate School of Frontier Biosciences. Osaka University, Osaka, Japan <sup>4</sup> Research Institute for Microbial Diseases Osaka University, Osaka, Japan <sup>5</sup> Komaba Institute for Science, The University of Tokyo, Tokyo, Japan <sup>6</sup> JST, PRESTO, Kawaguchi, Saitama 332-0012. Japan <sup>7</sup> Department of Life Science, Graduate School of Arts and Science. The University of Tokyo, Tokyo, Japan <sup>8</sup> Universal Biology Institute, The University of Tokyo, 3-8-1 Komaba, Meguro-ku, Tokyo, Japan

In prebiotic evolution, molecular self-replicators are considered to develop into diverse, complex living organisms. The appearance of parasitic replicators is believed inevitable in this process. However, the role of parasitic replicators in prebiotic evolution remains elusive. Here, we demonstrated experimental coevolution of RNA self-replicators (host RNAs) and emerging parasitic replicators (parasitic RNAs) using an RNA-protein replication system we developed. During a long-term replication experiment, a clonal population of the host RNA turned into an evolving host-parasite ecosystem through the continuous emergence of new types of host and parasitic RNAs produced by replication errors. The host and parasitic RNAs diversified into at least two and three different lineages, respectively, and they exhibited evolutionary arms-race dynamics. The parasitic RNA accumulated unique mutations, thus adding a new genetic variation to the whole replicator ensemble. These results provide the first experimental evidence that the coevolutionary interplay between host-parasite molecules plays a key role in generating diversity and complexity in prebiotic molecular evolution.

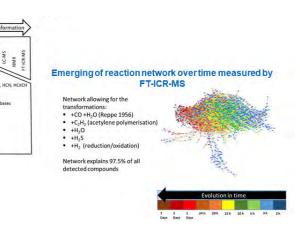




CHEMICAL EVOLUTION OF BIOMOLECULES FORMED UNDER **VOLCANIC HYDRO-**THERMAL CONDITIONS



Thomas Geisberger<sup>1</sup>, Philippe Diederich<sup>2</sup>, Claudia Huber<sup>1</sup> and Philippe Schmitt-Kopplin<sup>2</sup> <sup>1</sup> Lehrstuhl für Biochemie, Technische Universität München <sup>2</sup> Research Unit Analytical Biogeochemistry, HelmholtzZentrum München



Reactions mimicking volcanic-hydrothermal vent conditions are performed. Main group volatiles (e.g. CO., COS, HCN, C.H., H.S,  $N_{2}$ ,  $H_{2}$ ,  $NH_{3}$ , HCN,  $P_{4}O_{10}$ ) in aqueous solution react on the surfaces of crustal catalytic transition metal minerals (e.g. FeS, NiS, CoS)<sup>1</sup>. The basic setup of the experiment consists of NiS in water under an acetylene and carbon monoxide atmosphere. The mixture is kept at 378 K for several hours and the temporal evolution of the reaction mixture was monitored. Later experiments added NH<sub>2</sub> to the system. In addition to previously established GC/MS analysis, the combination of high-resolving analytical tools<sup>2,3</sup> and data-analytical approaches<sup>4</sup> will enable us to decipher the complex reaction mixtures. Fourier transform ion cyclotron resonance mass spectrometry (FT-ICR-MS), gas chromatography-MS (GC-MS) and nuclear magnetic resonance (NMR) methods provide complementary profiles of the complex product mixtures for both low and high molecular weight compounds, including a remarkable variety of CHOS derivatives.

1 C. Huber, G. Wächtershäuser, Science 1997, 276, 245-247. 2 P. Schmitt-Kopplin et al., Proc Natl Acad Sci U S A 2010, 107, 2763-2768. 3 O. P. Popova, et al., Chelyabinsk Airburst, Science 2013, 342, 1069-1073. 4 D. Tziotis et al. (2011) EJMS 17.4:415-421

# **SELECTION PRESSURE:** THE LATITUDINAL BIODIVERSITY GRADIENT IS DRIVEN BY THE LATITUDINAL VARIATION OF ULTRAVIOLET RADIATION



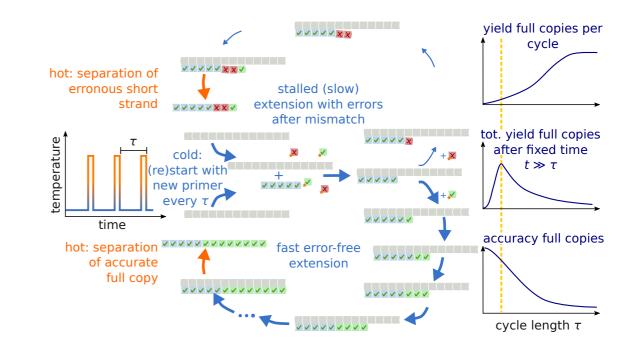
### Eftal Gezer

Department of Physics, Gebze Technical University, Gebze, 41400, Kocaeli, Turkey.

The fundamental principle of life is Darwinian evolution. Ultraviolet radiation is a known mutagen and a factor for selection pressure on Earth's prebiotics. Ultraviolet irradiance peaks at the tropics and minimises at the poles. Species richness also peaks at the tropics and minimises at the poles. Previous literature reviews hypothesise that increased biomass and species richness at the tropics might be caused by the solar energy presence, but the mechanism is unclear. In this study, the role of latitudinal variation of ultraviolet irradiance on the latitudinal biodiversity gradient is investigated. A mathematical relation between latitudinal biodiversity gradient and latitudinal variation of ultraviolet radiation is discussed. The preliminary approach shows that the latitudinal biodiversity gradient might be driven by the latitudinal variation of ultraviolet radiation.

- H. IKEHATA and T. ONO, Journal of Radiation Research 52, 115 (2011).
- C. Sagan, Journal of Theoretical Biology 39, 195 (1973)
- C. S. Cockell, Journal of Theoretical Biology 193, 717 (1998)
- F. S. Johnson, T. Mo, and A. E. S. Green, Photochemistry and Photobiology 23, 179 (1976).
- P. D. Mannion, P. Upchurch, R. B. J. Benson, and A. Goswami, Trends in Ecology & Evolution 29, 42 (2014).
- T M Blackburn and K J Gaston Biodiversity Letters 3 44 (1996) K. J. Gaston, T. M. Blackburn, Pattern and Process in Macroecology (Blackwell Science Ltd. n.d.). pp. 35-96 (2000)
- T. M. Blackburn and K. J. Gaston, Ecography 19, 369 (2006). D. M. Kaufman, Journal of Mammalogy 76, 322 (1995).





# A NONEQUILIBRIUM ERROR FILTERING MECHANISM FOR ENZYME-FREE COPYING OF NUCLEIC ACID δεουενсεδ



Tobias Göppel<sup>1</sup>, Benedikt Obermayer<sup>2</sup>. Joachim Rosenberger<sup>1</sup>, Bernhard Altaner<sup>1</sup>, Gabrielle Leveau<sup>3</sup>, Clemens Richert<sup>3</sup>, Irene Chen<sup>4</sup>, and Ulrich Gerland<sup>1</sup>

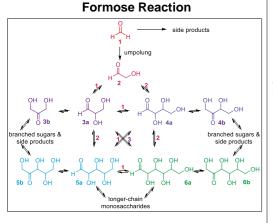
<sup>1</sup> Physics of Complex Biosystems, Physics Department, Technical University of Munich, James-Franck-Str. 1, D-85748 Garching, Germany

<sup>2</sup> Systems Biology of Gene Regulatory Elements, Berlin Institute for Medical Systems Biology, Max Delbrück Center for Molecular Medicine in the Helmholtz Association. Robert-Rössle-Straße 10, D-13125 Berlin, Germany; Core Unit Bioinformatics, Berlin Institute of Health, Charité -Universitätsmedizin Berlin, Charit latz 1, D-10117 Berlin, Germany.

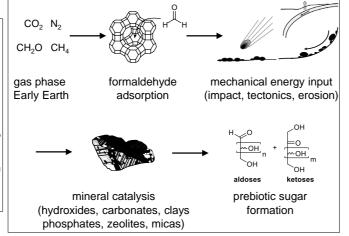
<sup>3</sup>Institute of Organic Chemistry, University of Stuttgart, Pfaffenwaldring 55, 70569 Stuttgart, Germany <sup>4</sup>Department of Chemistry and Biochemistry and Program in Biomolecular Science and Engineering, University of California, Santa Barbara, Santa Barbara, California 93106 United States; Department of Chemical and Biomolecular Engineering, University of California, Los Angeles, Los Angeles, California 90095, United States

GO Weblink to the Poster Room eqezer@qtu.edu.tr

Accurate copying of nucleic acid sequences by template-directed polymerization is essential for self-replicating and evolving systems. Modern cells achieve error rates as low as 10-9 with sophisticated enzymatic machineries that consume free energy to repeatedly discriminate between correct and incorrect nucleotides. In contrast, experiments probing template-directed, nonenzymatic extension of RNA and DNA as potential prebiotic copying processes find error rates of roughly 10-1, making it impossible to reliably transmit information from one strand generation to another. However, it is observed that initial incorporation errors trigger a cascade of consecutive errors and significantly reduce the speed of downstream extension, an effect called stalling. This phenomenon opens the door to an early error reduction mechanism: Using computer simulations and mathematical modelling, we show that error cascades can be exploited to discriminate between faithful and faulty polymerization products by means of their global kinetics. Limiting the time window for the polymerization process prevents erroneous strands to complete resulting in a pool where fulllength products show an enhanced accuracy. Such a mechanism does not require any additional energy input to the extension reaction itself and can be controlled externally. However, filtering out strands is at the expense of the overall yield and hence a characterization of the fidelityyield trade-off is needed. The yield problem can be circumvented though a repeated copying process where a strand serves as a template multiple times. Such a process may be induced by temperature cycles occurring naturally in the vicinity of hydrothermal vents which were common on the early Earth.



### Mechanochemistry as a scenario for the Early Earth



### A MINERAL-CATALYSED. MECHANOCHEMICAL FORMOSE REACTION

### Maren Haas<sup>1,2</sup>, Saskia Lamour<sup>1,2</sup>, Sarah B. Christ<sup>2</sup>, Oliver Trapp<sup>1,2</sup>

<sup>1</sup> Max-Planck-Institute for Astronomy, Königstuhl 17, 69117 Heidelberg, Germany <sup>2</sup>Department of Chemistry & Pharmacy, Ludwig-Maximilians-University, Butenandtstr. 5-13, 81377 Munich, Germany

The formose reaction builds up monosaccharides from formaldehyde and glycolaldehyde and a basic catalyst and is one of the possible formation pathways for carbohydrates on the Early Earth.<sup>1,2</sup> However, it suffers from side reactions and deterioration of the products under the highly alkaline reaction conditions.<sup>3</sup>

Most of the postulated scenarios for prebiotic reactions are based on aqueous reaction media like the "warm little pond" or hydrothermal vents.<sup>4</sup> Solvent-free reactions represent an alternative setting for prebiotic reactions and can be accelerated by mechanic energy inputs like lithospheric activity, weathering, erosion, diagenesis, tectonics or meteor impacts. Under laboratory conditions ball mills are used for the investigation of mechanochemical reactions.

We present the combination of the formose reaction and the mechanochemical setting in the context of the origins of life. The formation of carbohydrates by ball milling formaldehyde and glycolaldehyde with mineral catalysts is observed.<sup>5</sup> These reactions gain from higher selectivity and decreased decomposition in comparison to the aqueous reaction. Two-step derivatization and subsequent analysis by gas chromato-graphy-mass spectrometry is used to investigate the formed reaction mixtures.6

1 A. Butlerow, Justus Liebigs Ann. Chem. 1861, 120, 295-298. 2 D. Kopetzki, M. Antonietti, New J. Chem. 2011, 35, 1787-1794. 3 R. Larralde, M. P. Robertson, S. L. Miller, Proc. Natl. Acad. Sci. 1995, 92, 8158-8160. 4 J. L. Bada, Earth Planet. Sci. Lett. 2004, 226, 1-15. 5 S. Lamour, S. Pallmann, M. Haas, O. Trapp, Life 2019, 9, 52.

6 M. Haas, S. Lamour, O. Trapp, J. Chromatogr. A 2018, 1568, 160-167.

# INDEX - FLOOR 6

86	PHILIPP HONEGGER & CHRISTOPH FLAMM Identification of autocatalysis in a reaction r
87	CLAUDIA HUBER Towards a possible acetyleno/carboxydotroph
88	ALAN IANESELLI Heated microdroplets of acidic water induce the of longer strands
89	MIKOŁAJ JANICKI Photoreduction of thioanhydroadenosine to
90	ANTON JOSEPH Design of stealth cell-mimetic dendrimersome
91	NOZAIR KHAWAJA Exploring the biogeochemistry of extraterrestr
92	<b>SVEN KLUMPE</b> Ultrastructural analysis of nuclear pore compl FIB LiftOut
93	KAI KOHLER & FLORIAN GARTNER Evolutionary optimization of experimental sy
94	BALAZS KONNYU Dynamics and stability in prebiotic information
95	NINA KOSTINA Sugar-driven formation of artificial raft-domai dendrimersome protocells
96	<b>PATRICK KUDELLA</b> Sequence self-selection by the dynamics of a



Aligns with meeting schedule

network – A graph-topological approach

hic core metabolism under primordial conditions

he denaturation of oligonucleotides and the replication

the purine deoxyribonucleosides

rial active ocean worlds

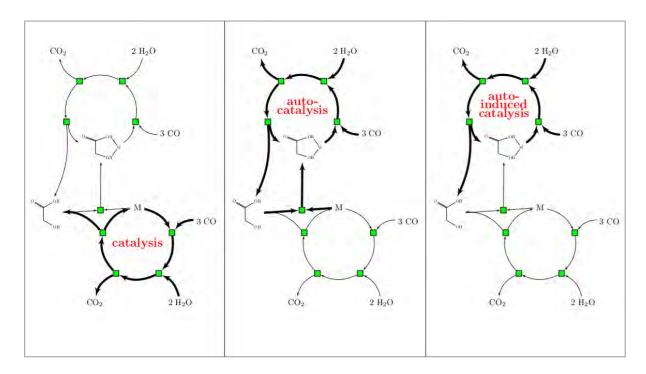
lex assembly in early Drosophila embryos by cryo

ynthetic networks

n integration: An RNA world model from first principles

ins with hierarchical periodic nanoarrays on

random oligomer pool network



# **IDENTIFICATION** OF AUTOCATALYSIS IN A **REACTION NETWORK - A GRAPH-TOPOLOGICAL APPROACH**



### Philipp Honegger<sup>1</sup> and Christoph Flamm

Institute of Computational Biological Chemistry, University of Vienna, Währinger Straße 17, 1090 Vienna,

Austria Institute of Theoretical Chemistry, University of Vienna, Währinger Straße 17, 1090 Vienna, Austria

The concept of autocatalysis refers to a chemical reaction producing copies of its catalyst. Autocatalysis is indispensible to abiogenetical questions due to the ability of such systems to feed on input molecules, grow and reproduce. Autocatalytic species possess the necessary robustness against environmental shocks and continuous molecular degradation,<sup>1</sup> and they allow for the emergence of enantiomeric excesses ex nihilo by ampliying statistical fluctuations of enantiomer ratios.<sup>2,3</sup> Even evolved modern-day metabolisms are suspected to contain ancestral autocatalytic cores.4

We present the structural requirements autocatalysis imposes on the underlying chemical reaction networks and show examples from chemistry and biology. This permits the comprehensive detection of possible autocatalytic reactions. In this context, we also show its reverse counterpart, destructive autocatalysis. Reverse-autocatalytic motifs present in the reaction network are able to counteract productive autocatalytic cycles.

Our graph-topological analysis also differentiates between autocatalysis and autoinduction. The latter refers to chemical reactions where a product enhances the reaction rate of its production without producing more catalyst, for instance by starting material activation. Autoinductive reactions can exhibit similar kinetic signatures, but they do not possess the persistence aspect that distinguishes autocatalysis from common catalysis.1

1 Blackmond, D. G. Angew. Chem. Int. Ed. 2009, 48, 386-390. 2 Frank, F. C. Biochimica et biophysica acta 1953, 11, 459-463. 3 Blackmond, D. G. Angew, Chem, Int. Ed. 2009, 48, 2648-2654 4 Kun, Á.: Papp, B.: Szathmáry E. Genome biology 2008, 9, R51 TOWARDS A POSSIBLE ACETYLENO/

# CARBOXYDOTROPHIC CORE METABOLISM **UNDER PRIMORDIAL** CONDITIONS

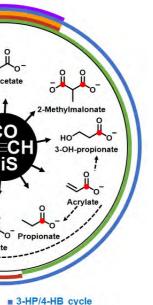


Thomas Geisberger<sup>1</sup>, Jessica Sobotta<sup>1</sup>, Thomas Steiner<sup>1</sup>, Christian Seitz<sup>1</sup>, Günter Wächtershäuser<sup>2</sup>, Wolfgang Eisenreich<sup>1</sup>, Claudia Huber Lehrstuhl für Biochemie. Department Chemie, Technische Universität

München, Lichtenbergstraße 4, 85748 Garching, Germany <sup>2</sup> 209 Mill Race Drive, Chapel Hill, NC 27514, USA

Theories concerning the origin and early evolution of life have to consider carbon fixation as well as the evolution of metabolism. The extant biosphere owes its existence mainly to CO<sub>2</sub>-fixation. Due to the low reactivity of CO<sub>2</sub>, alternative geochemically available carbon sources should be considered. Acetylene (C<sub>2</sub>H<sub>2</sub>) and carbon monoxide (CO) can be found in hydrothermal exhalations<sup>1,2</sup> and iron-nickel-sulfides were present in early earth's crust<sup>3</sup>. We could show a one-pot carbon fixation of acetylene and carbon monoxide by aqueous nickel sulfide (NiS) under hydrothermal (>100°C) conditions<sup>4</sup>. We found more than ten key  $C_{2,4}$  constituents of the extant four central CO<sub>2</sub>-fixation cycles of the domains Bacteria and Archaea<sup>5-7</sup>. Some of the organic products engage in the same interconversions as seen in the extant central CO<sub>2</sub>fixation cycles. A primordial, non-cyclic acetyleno/carboxydotrophic metabolism is suggested. It is based on aqueous organo-metal chemistry, from which the extant central CO<sub>2</sub>-fixation cycles based on thioester chemistry would have evolved by piecemeal modifications. 1 S. Igari et al.;Chikyukagau 34, 7 (2000). 2 J.R. Holloway & J.G. Blank; Rev. mineral. 30, 187 (1994). 3 R.M. Hazen; Sci. Am. 302, 58 (2010).

8 T. Geisberger et al; Life, 9, 50 (2019)

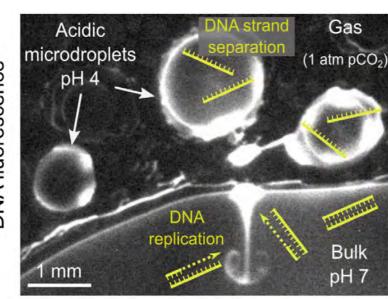


rTCA cycle DC/4-HB cycle 3-HP bicycle

■ rAcCoA

- 4 J. Sobotta, T. Geisberger et al.; Life, 10 (4), 35, (2020)
- 5 G. Fuchs.; Ann. Rev. microbial. 65, 631 (2011).
- 6 I.A.Berg; Appl. Environ. Microbiol. 77, 1925 (2011). 7 M. Hügler & S.M.Sievert: Ann. Rev. mar. scie. 3. 261 (2011).

**DNA fluorescence** 



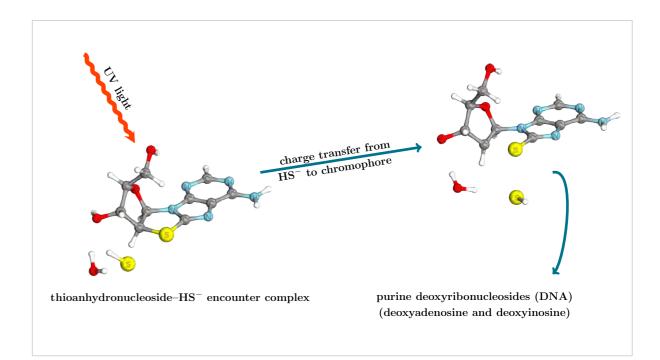
# **HEATED MICRODROPLETS** OF ACIDIC WATER INDUCE THE DENATURATION OF **OLIGONUCLEOTIDES AND** THE REPLICATION OF LONGER STRANDS



### Alan Ianeselli<sup>1,2</sup>, Miguel Atienza Juanatey<sup>1</sup>, Christof B. Mast<sup>1,2</sup> and Dieter Braun<sup>1,2</sup>

Systems Biophysics, Ludwig Maximilian University Munich Amalienstraße 54. 80799 München. Germany <sup>2</sup>Center for NanoScience (CeNS), Ludwig Maximilian University. Amalienstraße 54, 80799 München, Germany

The understanding of how the replication and the evolution of DNA or RNA sequences could have occurred on the primordial Earth is still an open debate in the Origin of Life community. Paradigms like Spiegelman's "tyranny of the shortest"1 and the "strand separation problem"2 are still some of the major barriers that most of the current models are not able to surpass. In this work, we created a prebiotically plausible physical system that tackles the aforementioned problems and could host the replication and the evolution of oligonucleotides autonomously. Our system consists of a thermal gradient across a rock pore containing liquid and gas enriched in CO<sub>2</sub>. It periodically generated acidic microdroplets of condensation water that denatured DNA and RNA duplexes at moderate temperatures. At the same time, oligonucleotides of increasing length were accumulated at the gaswater boundaries and were preferentially denatured. The system was able to host enzymatic DNA replication at temperatures lower than the melting temperature and promoted the replication of longer sequences, avoiding the dominance of the shorter sequences in the pool.



### PHOTOREDUCTION OF THIOANHYDROADENOSINE TO THE PURINE DEOXYRIBONUCLEOSIDES

Mikołaj J. Janicki<sup>1</sup>, Rafał Szabla<sup>2,3</sup>, Robert W. Góra<sup>1</sup> <sup>1</sup> Department of Physical and Quantum Chemistry, Wrocław University of Science and Technology, Faculty of Chemistry, Wybrzeże Wyspiańskiego 27, 50-370, Wrocław Poland

<sup>2</sup>EaStCHEM, School of Chemistry, University of Edinburgh, Joseph Black Building, David Brewster Road, Edinburgh, EH9 3FJ, UK

<sup>3</sup>Institute of Physics, Polish Academy of Sciences, Al. Lotników 32/46, PL-02668 Warsaw, Poland.

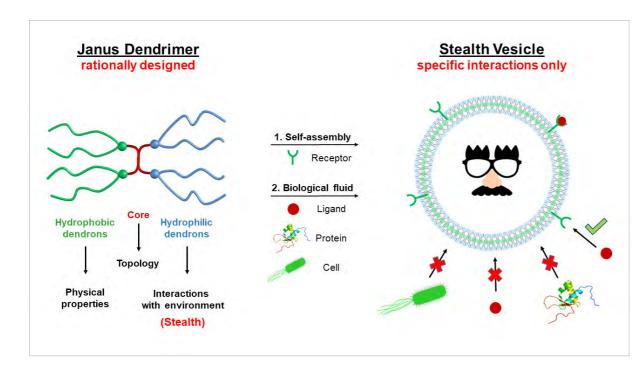
In recent decades, a lot of efforts have been put into the exploration of abiotic synthesis of RNA and DNA nucleosides from prebiotically plausible feedstock molecules. Substantial progress has been made in the prebiotic synthesis of pyrimidine ribonucleosides. Despite many attempts, the abiotic synthetic routes towards DNA molecules, under the credible geochemical scenario, have not been found so far. The available results seem to support the RNA world hypothesis assuming RNA molecule could have been the first genetic polymer at the primordial Earth.1 Furthermore, in this scenario, DNA polymer is appearing at a later stage of the development of early life. Our collaborative research performed with prof. John Sutherland's group enabled to discover the selective prebiotic synthesis of the purine deoxyribonucleosides showing that RNA and DNA may have coexisted at the origins of life.<sup>2</sup> Photoreduction of thioanhydroadenosine plays a key role in the newly uncovered prebiotic route to deoxyribonucleosides. To explain the mechanisms underlying photoreduction of thioanhydronucleosides, we performed quantum-chemical calculations of the excited-state potential energy surfaces to provide a mechanistic rationale for the UV-driven chemical reaction.

1 Gilbert W., Origin of life: The RNA world, Nature 319, 618 (1986) 2 Xu J., Chmela V., Green N.J., Russell D. A., Janicki M. J., Góra R. W., Szabla R., Bond A. D., Sutherland J. D., Selective prebiotic formation of RNA pyrimidine and DNA purine nucleosides, Nature, Accepted Manuscript

<sup>1</sup> Mills, D. R.; Peterson, R. L.; Spiegelman, S. An Extracellular Darwinian Experiment with a Self-Duplicating Nucleic Acid Molecule, Proc. Natl. Acad. Sci. U. S. A. 1967, 58 (1), 217-224, https:// doi.org/10.1073/pnas.58.1.217

<sup>2</sup> Szostak, J. W. The Eightfold Path to Non-Enzymatic RNA Replication. J. Syst. Chem. 2012, 3 (1), 2. https://doi.org/10.1186/1759-2208-3-2.





# DESIGN OF STEALTH **CELL-MIMETIC** DENDRIMERSOMES

### A. Joseph and C. Rodriguez-Emmenegger DWI - Leibniz Institute for Interactive Materials, Forckenbeckstraße 50, 52074 Aachen, Germany.

The development of bottom-up synthetic cells requires the implementation of receptors into their membrane while the rest of the vesicle surface should be repellent towards non-specific adsorptions of proteins, sugars and cells. The current gold standards for stealth vesicle are co-assembled from natural lipid and 5 mol% poly(ethylene glycol) (PEG) containing synthetic lipid. However, the steric constrains of large-sized PEG and the force exert by these polymers to the membrane destabilizes the resulting liposome, which necessitates the stabilization by cholesterol. Therefore, a roughly equimolar ratio of natural phospholipid and cholesterol is required. While cholesterol stabilizes the liposome membrane, it also reduces its flexibility and lateral mobility - two physical properties that are essential for biomimicry.<sup>1</sup> This is the reason why new strategies are needed to generate monocomponent synthetic cell membranes with stealth properties while maintaining a biomimetic system.

Amphiphilic Janus dendrimers (JDs) are known to self-assemble into dendrimersomes (DSs) which amalgamate many biomimetic physical properties such as a bilayer thickness of ~ 5 nm, high flexibility, mechanical stability and lateral mobility.<sup>2</sup> Due to their modular synthesis, their physical properties and their interactions with the environment can be tuned by rational design. We synthesized JDs that incorporate PEG or zwitterionic moieties, known to prevent protein and cell adsorption onto surfaces, in their hydrophilic section.<sup>3</sup> With DSs self-assembled from these JDs, we aim to study selective interactions between specific ligands and receptors in the future.

1 M. L. Immordino et al., Int. J. Nanomedicine 2006, 1, 297-315. 2 V. Percec et al., Science 2010, 328, 1009-1014. 3 I. Banerjee et al., Adv. Mater. 2011, 23, 690-718

# EXPLORING THE BIOGEOCHEMISTRY OF EXTRATERRESTRIAL ACTIVE OCEAN WORLDS

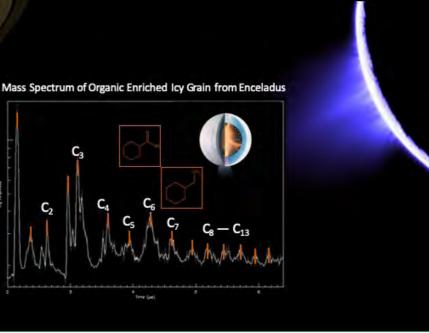


Nozair Khawaia, Fabian Klenner Jon Hillier, Frank Postberg, Marie Dannenmann, Zenghui Zou Inst. of Geological Sciences, Freie University Berlin, Germany

active ocean worlds. 1 Waite et al. (2017) Science

2 Hsu et al. (2015) Nature

GO Weblink to the Poster Room



Saturn's moon Enceladus (and potentially Jupiter's moon Europa) has a subsurface liquid water ocean that interacts with its rocky core via hydrothermal vents. From its ocean, Enceladus emits a plume of gas and ice grains into space through fractures near its south pole. This plume was sampled by mass spectrometers on the Cassini spacecraft - the Cosmic Dust Analyzer (CDA) and the Ion and Neutral Mass Spectrometer (INMS). Alkaline hydrothermal vents on Enceladus, likely to be similar to those believed to be possible sites for the emergence of life in Earth's oceans, were confirmed by INMS and CDA detections of H<sub>2</sub>, CH<sub>4</sub><sup>1</sup> and nanometre-sized silica particles<sup>2</sup>. Additional discoveries of biologically-relevant N-, O- and aromatic low- and high-mass organic compounds<sup>3,4</sup> reveal complex and reactive organic chemistry within the moon. We use Laser-Induced Liquid Beam Ion Desorption (LILBID) time of flight mass spectrometry, a proven analogue for CDA in situ ice grain mass spectrometry, to infer the biogeochemistry of Enceladus' ocean by comparing the LILBID mass spectra of selected compounds with those from CDA. Geochemically-relevant salts, as well as biologicallyrelevant organics, such as amino acids, fatty acids and peptides<sup>5a</sup>, have been tested, and identified in the resulting mass spectra. Biotic and abiotic mass spectral fingerprints could be discriminated and biomolecule detection limits were found to be at the µM to nM level<sup>5b</sup>, while those for salts (including sulfates and phosphates) are currently being measured and applied to CDA spectra. Further biological samples such as bacterial DNA and lysed cell material are under evaluation. The results we summarise here aid planning for future space missions to

3 Postberg et al. (2018) Nature 4 Khawaja et al. (2019) MNRAS 5 Klenner et al. (2020a & b) Astrobiology

# ULTRASTRUCTURAL ANALYSIS OF NUCLEAR PORE COMPLEX ASSEMBLY IN EARLY DROSOPHILA EMBRYOS BY CRYOFIB LIFTOUT



Sven Klumpe<sup>1,3</sup>, Bernhard Hampoelz<sup>2,3</sup>, Philipp Erdmann<sup>1</sup>, Janina Baumbach<sup>2,3</sup>, Martin Beck<sup>2,3</sup>, Jürgen M. Plitzko<sup>1</sup> <sup>1</sup> Max Planck Institute for Biochemistry, Martinsried. Germany

 <sup>2</sup> European Molecular Biology Laboratory, Structural and Computational Biology Unit, Heidelberg, Germany
<sup>3</sup> Max Planck Institute for Biophysics, Frankfurt am Main, Germany For more than a century, Drosophila melanogaster has been one of the most prevalent model organisms in developmental biology. Recently discovered, D. melanogaster shows a non-canonical pathway of nuclear pore complex (NPC) biogenesis in early development that is likely present in other metazoans<sup>1</sup>. NPCs pre-assemble within so called annulate lamellae (AL) - sheets of endoplasmic reticulum - that insert into the nuclear envelope (NE) during early development to maintain a constant NPC/NE ratio. Due to the loss of ultrastructural information in classical plastic-embedded samples, the structural details of this process remain elusive. Structural analysis of multicellular organisms at cryo-temperatures by transmission electron microscopy has recently been made possible by cryo-focused ion beam milling and lift-out<sup>2</sup>. However, the procedure is tedious and time-consuming. Here, we present software and hardware solutions to improve the throughput and streamline the procedure for a reliable and robust workflow. With these developments, we are now able to study the ultrastructure of NPC assembly and AL insertion during early Drosophila development at unprecedented resolution. The membrane remodeling and protein condensation principles<sup>3</sup> that underlie this process have the potential to give ideas on the governing mechanisms in early eukaryotic life.

1 Hampoelz, B., et al., Pre-assembled Nuclear Pores Insert into the Nuclear Envelope during Early Development. Cell, 2016. 166(3): p. 664-678.

2 Schaffer, M., et al., A cryo-FIB lift-out technique enables molecular-resolution cryo-ET within native Caenorhabditis elegans tissue. Nat Methods, 2019. 16(8): p. 757-762. 3 Hampoelz, B., et al., Nuclear Pores Assemble from Nucleoporin Condensates During Oogenesis. Cell, 2019. 179(3): p. 671-686 e17.

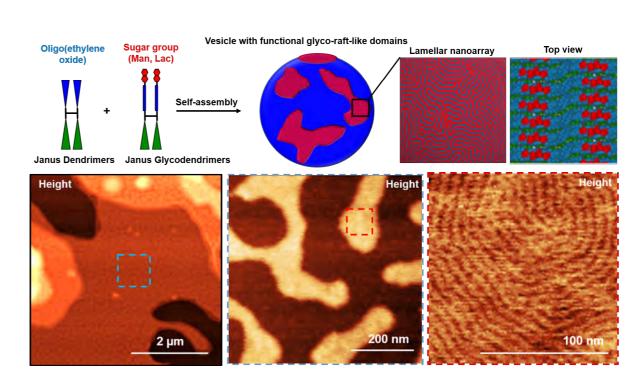
# EVOLUTIONARY OPTIMIZATION OF EXPERIMENTAL SYNTHETIC NETWORKS



Severin Angerpointner<sup>1</sup>, Florian Gartner<sup>1</sup>, Kai Kohler<sup>2</sup>, Maren Haas<sup>2</sup>, Erwin Frey<sup>1</sup>, Oliver Trapp<sup>2</sup> <sup>1</sup> University of Munich (LMU), Theresienstr. 37, 80333 Munich, Germany <sup>2</sup> University of Munich (LMU), Butenandtstr. 7-13, 81377 Munich, Germany Up to date, the formose reaction is the most plausible pathway in molecular evolution leading to the formation of sugar molecules. A plethora of monosaccharides is formed by dimerization of formaldehyde, subsequent (retro-)aldol reactions and aldose-ketose-isomerizations in the presence of basic catalysts.<sup>1</sup> Albeit Breslow had suggested an auticatalytic cycle starting as soon as traces of glycolaldehyde have been formed,<sup>2</sup> the initial dimerization of formaldehyde has not yet been mechanistically elucidated. In this collaboration project an extended formose reaction network will be comprehensively studied. The interplay of improved separation and analytic techniques with modern computer-aided simulations based on sloppy models<sup>3</sup> is the basis for tackling these challenges.

klumpe@biochem.mpg.de 📀 Weblink to the Poster Room

A. Butlerow, Ann. Chem. Pharm. 1861, 120, 295-298.
R. Breslow, Tetrahedron Lett. 1959, 1, 22-26.
M. Transtrum et al., J. Chem. Phys. 2015, 143, 01091.



**SUGAR-DRIVEN** FORMATION OF ARTIFICIAL RAFT-DOMAINS WITH **HIERARCHICAL PERIODIC** NANOARRAYS ON DENDRIMERSOME PROTOCELLS



Nina Yu. Kostina<sup>1</sup>, Dominik Söder<sup>1</sup>, Anna Wagner<sup>1</sup>, Tamás Haraszti<sup>1</sup>, Khosrow Rahimi<sup>1</sup>, Virgil Percec<sup>2</sup>, César Rodriguez-Emmenegger<sup>1</sup> <sup>1</sup> DWI- Leibniz Institute for Interactive

Materials and Institute of Technical and Macromolecular Chemistry RWTH Aachen University Forckenbeckstraße 50, 52074 Aachen, Germany <sup>2</sup> Roy & Diana Vagelos Laboratories, Department of Chemistry, University of Pennsylvania Philadelphia, Pennsylvania 19104-6323, United States.

DYNAMICS AND **STABILITY IN PREBIOTIC** INFORMATION **INTEGRATION: AN RNA** WORLD MODEL FROM FIRST PRINCIPLES

Model

Results

The MCRS on

al surfaces

E,

 $(\mathbf{P}) = \mathbf{M} = \mathbf{E}$ 



### András Szilágyi<sup>1,2,3</sup>, Balázs Könnyű<sup>1,4</sup> & Tamás Czárán<sup>1,2,5</sup>

<sup>1</sup> Evolutionary Systems Research Group, MTA Centre for Ecological Research, Klebelsberg K. u 3, 8327, Tihany, Hungary.

<sup>2</sup>MTA–ELTE Theoretical Biology and Evolutionary Ecology Research Group, Eötvös Loránd University, Pázmány P. s. 1C, 1117, Budapest, Hungary. <sup>3</sup> Center for Conceptual Foundation of Science, Parmenides Foundation, Kirchplatz 1, 82049, Pullach/Munich, Germany

<sup>4</sup> Department of Plant Systematics, Ecology and Theoretical Biology, Institute of Biology, Eötvös Loránd University, Pázmány P. s. 1 C, 1117, Budapest, Hungary. <sup>5</sup> Biocomplexity Group, Niels Bohr Institute, Copenhagen University, Blegdamsvej 17, 2100, Copenhagen, Denmark

The robust coevolution of catalytically active, metabolically cooperating prebiotic RNA replicators were investigated using an RNA World model of the origin of life based on physically and chemically plausible first principles. The Metabolically Coupled Replicator System assumes RNA replicators to supply metabolically essential catalytic activities indispensable to produce nucleotide monomers for their own template replication. Using external chemicals as the resource and the necessary ribozyme activities, Watson-Crick type replication produces complementary strands burdened by high-rate point mutations (insertions, deletions, substitutions). Metabolic ribozyme activities, replicabilities and decay rates are assigned to certain sequence and/ or folding (thermodynamical) properties of single-stranded RNA molecules. Short and loosely folded sequences are given replication advantage, longer and tightly folded ones are better metabolic ribozymes and more resistant to hydrolytic decay. We show that the surface-bound MCRS evolves stable and metabolically functional communities of replicators of almost equal lengths, replicabilities and ribozyme activities. Being highly resistant to the invasion of parasitic (nonfunctional) replicators, it is also stable in the evolutionary sense. The template replication mechanism selects for catalytic "promiscuity": the two (complementary) strands of the same evolved replicator will often carry more than a single catalytically active motif, thus maximizing functionality in a minimum of genetic information.

Complementarity and folding

G :5'-ACCGACAGCG...GGCUUGGC-3'

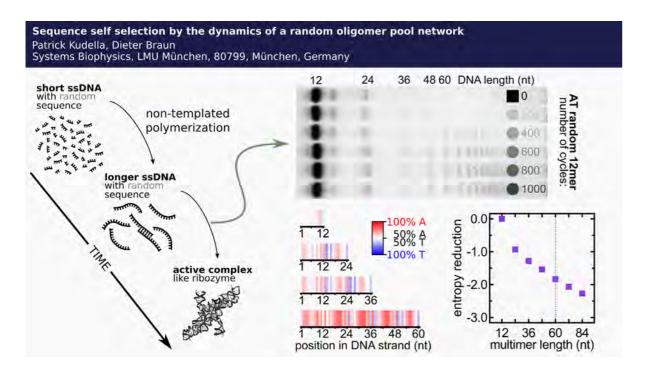
G :3'-UGGCUGUCGC...CCGAACCA-5'

000



The seminal fluid mosaic model of the cell membranes suggests a lipid bilayer sea, in which cholesterol, proteins, glycoconjugates, and other components are swimming. Complementing this view, a microsegregated rafts model predicts clusters of components that function as relay stations for intracellular signaling and trafficking. Despite decades of work, the structure and function of these domains in cells remains difficult to resolve. Their immense complexity of functions arises from the combination of the chemical diversity of sugar moieties and their spatial 3D presentation. To tackle this challenge, we synthesized Janus dendrimers (JDs) and Janus glycodendrimers (JGDs) decorated with sugar moieties that in water self-assemble into vesicles that function as biological membrane mimics. We discovered that JGDs, on which the sugar groups were diluted in a defined way among tri(ethylene oxide) units, gave rise to lamellar or hexagonal nanoarrays and elicits higher bioactivity to sugar-binding proteins.<sup>1,2</sup> But could such nanoassemblies be preorganized into functional domains that mimic raft-domain in cells? And could the phase-separation be driven by solely hydrophilic interactions between amphiphilic molecules without the addition of cholesterol? We study the co-assembly of JDs (hydrophilic dendron decorated by tri(ethylene oxide)) with JGDs (decorated by sugar moieties). Our studies revealed that the sizes and nanoarrays of raft-like domains can be controlled by the ratio of JD to JGD and sugar type and that phase separation occurs only by hydrophilic interactions between tri(ethylene oxide) and sugar groups. The studies on understanding the formation of glycan periodic nanoarrays may help to shed light on cell communication and signaling and provide a powerful example in which structure determines function.

1 Rodriguez-Emmenegger, C. et al. Proc. Natl. Acad. Sci. U S A, 116, 5376-5382 (2019). 2 Xiao, Q. et al. Proc. Natl. Acad. Sci. U S A (just accepted)(2020)



# SEQUENCE SELF-**SELECTION BY THE** DYNAMICS OF A RANDOM **OLIGOMER POOL NETWORK**



Patrick Kudella, Dieter Braun Systems Biophysics, LMU München, 80799, München, Germany

Replication of information on oligonucleotides such as RNA or DNA is essential for the emergence of life<sup>1,2</sup>. Previous studies focus on the replication of single sequences, but we believe it is the key to monitor selection dynamics and replication starting in an already completely random pool of sequences. Theoretical work suggests a significant reduction in sequence entropy and therein a selection of a subset of the "fittest" sequences in a dawn of life scenario<sup>3</sup>. We expect a nonlinear ligation dynamic had set in, once polymerization was able to create oligomers long enough for hybridization and thus capable of structure formation and templated ligation. This "mechanism naturally involves the information transmission from a template to the newly ligated chain, thus opening an exciting possibility of long-term memory and evolvability"<sup>3</sup>. In the experiments we show how a system of 12mer random sequence DNA reduces its information entropy dramatically under an enzyme based template ligation reaction and temperature cycling. We obtained more than 12 million individual strands by Next Generation Sequencing (NGS). Utilizing a selfwritten LabView code we can study different mechanisms such as the emergence of a position dependent sequence pattern and a segregation into mutually complementary pools of A-rich and T-rich sequences. Both effects are associated with the development of a multiscale ligation landscape with multiple mutually catalyzing subpopulations.

1 Doudna, J. A. & Szostak, J. W. RNA-catalysed synthesis of complementary-strand BNA, Nature 339, 519-522 (1989). 2 Walter, G. The RNA World. Nature 319, 618 (1986) 3 Tkachenko, A. V & Maslov, S. Spontaneous emergence of autocatalytic informationcoding polymers. J. Chem. Phys. 143, 045102 (2015).

### INDEX - FLOOR 7

Aligns with meeting schedule

98	ALEXANDRA KÜHNLEIN Sequence dependent gelation, accumulation
99	CHANDRASHEKHAR V. KULKARNI Estimating preferential localization of interacti
100	SUDARSHANA LAHA Droplets as biochemical reactors in living cells
101	GABRIELLE LEVEAU Bridging the gap between chain formation an
102	YONGDA LI Biochemical methods for the detection of prot
103	TIM LICHTENBERG Atmospheric speciation of rocky planets from
104	KAI LIU Towards darwinian evolution of synthetic repli
105	SAVINO LONGO Anomalous fluctuations and selective extinc the origin of biological chirality
106	OLIVER MAGUIRE Cycling of orthophosphate under mild prebioti
107	THOMAS MATREUX Heat flows shift chemical equilibria by selectiv
108	GAIA MICCA LONGO New insights on prebiotic chemistry from plas



and sedimentation

ing molecules in model lipid membranes

nd genetic copying of RNA

teins as life signatures on Mars-like soils

magma ocean outgassing

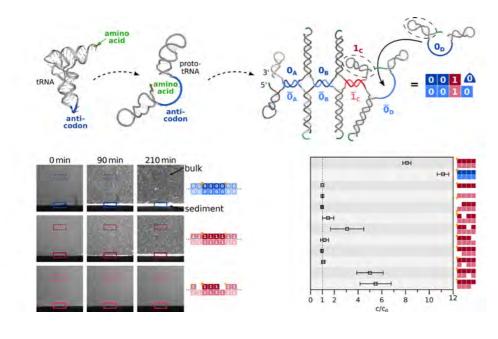
icators

ction in primordial replicators: a "struggle for life" at

ically plausible conditions

ve accumulation

sma kinetics



# SEQUENCE DEPENDENT **GELATION** ACCUMULATION AND **SEDIMENTATION**



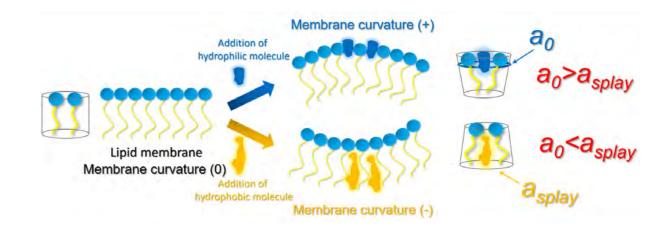
Alexandra Kühnlein<sup>1</sup>, Thomas Matreux<sup>1</sup>, Deni Szokoli<sup>2</sup>. Hannes Mutschler<sup>2</sup>. Dieter Braun<sup>1</sup>, Christof B. Mast<sup>1</sup> Biophysics and Center for NanoScience, LMU Munich, Amalienstraße 54, 80799 München <sup>2</sup> Max Planck Institute of Biochemistry, Martinsried, Germany

The origins of biological information constitute a major challenge for understanding the origins of life. Under Darwinian evolution, a localized, homogeneous genotype is selected. A first attempt would be to start from a random sequence pool. The emerging state of matter converges towards a phenotype that is optimal under a given selection pressure, e.g. a physical non-equilibrium. To jumpstart Darwinian evolution, the random sequence pools must show physical phenotypes. The question is, however, how to obtain such a process without already evolved replicases in a naïve system.

Firstly, we show preliminary results that indicate a self-selection of sequences by cooperative binding. We started from a pool of eight complementary hairpins, inspired from tRNA, designed to build a hybridization based self-replicator<sup>1</sup>. To initialize the sample, we heat it up to 95°C to melt all prevailing hydrogen bonds. Upon subsequent cooling, we observe sudden agglomeration of DNA in the system. These agglomerates sediment under gravity. If one of the eight sequences are missing, no significant gelation and no sedimentation occurs. Secondly, we subject a pool of random sequences to thermal gradients, where convection and thermophoresis can lead to size-dependent accumulation.

Through analysis on native PAGE gels<sup>2</sup>, Illumina-Sequencing and subsequent MFE calculations we analyse if and how the initially random sequence pools are biased by the temperature gradient. We speculate that in the long run, only a reduced number of cooperative binding sequences could remain in such a non-equilibrium setting.

1 Krammer, H., Möller, F. M., & Braun, D. (2012). Thermal, autonomous replicator made from transfer RNA, Physical review letters, 108(23), 238104. 2 Chizzolini, F., Passalacqua, L. F., Oumais, M., Dingilian, A. I., Szostak, J. W., & Lupták, A. (2019). Large phenotypic enhancement of structured random RNA pools. Journal of the American Chemical Society.



ESTIMATING PREFERENTIAL LOCALIZATION OF INTERACTING MOLECULES IN MODEL LIPID MEMBRANES

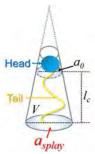


Chandrashekhar V Kulkarni Centre for Smart Materials, School of Physical Sciences and Computing, University of Central Lancashire, Preston PR1 2HR, United Kingdom.

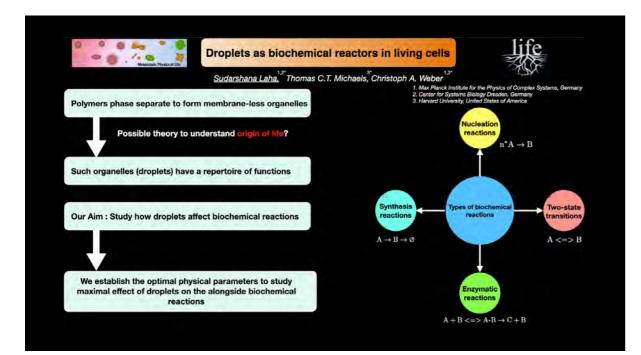
exhibiting hydrophilic, hydrophobic or amphiphilic characters

Spectroscopic techniques, in particular NMR, adequately reveal physicochemical interactions among a diverse set of molecules. However, when it comes to viscoelastic systems, including lipid phases, the applicability of these techniques remains limited due to poor signal quality caused by restricted diffusion. Here we present a novel method to estimate preferential localization of various molecules in lipid systems. This method is based on our recent work on 'chain splay'1 and its relevance in quantifying molecular shapes of amphiphilic molecules, including lipids.

Figure 1: Commonly adopted truncated (inverse) conical shape by a majority of lipid molecules By employing molecular level parameters (Figure 1), namely cross-sectional area at the head group (a0), lipid chain length (lc), molecular volume (V) and chain splay area (asplay) [1], one can perform quantitative estimation of an average shape of the amphiphilic molecule. Here, we show that by comparing above parameters with the parameters obtained after adding an additive molecule, it is possible to assess where that molecule prefers to localize within the membrane. This work finds great potential in formulating lipid systems for loading and interaction of a wide range of drugs and biomolecules



1 Kulkarni, C.V.\* Calculating the 'Chain Splay' of Amphiphilic Molecules: Towards Quantifying the Molecular Shapes (2019) Chemistry and Physics of Lipids, 218, pp 16-21.



# DROPLETS Αδ **BIOCHEMICAL REACTORS** IN LIVING CELLS

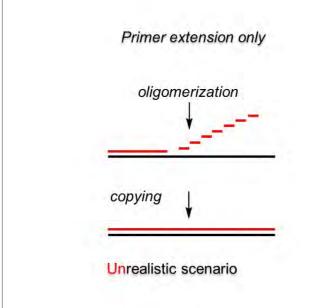


### Sudarshana Laha<sup>1,2</sup>, Thomas C.T

Michaels<sup>3</sup> and Christoph A. Weber<sup>1,2</sup> <sup>1</sup> Max Planck Institute for the Physics of Complex Systems, Germany <sup>2</sup>Center for Systems Biology Dresden, Germany <sup>3</sup> Harvard University, United States of America

Living cells use compartments (droplets) to spatially organise molecules that can undergo fuel-driven chemi-cal reactions<sup>1</sup>. Not much is known about the mechanisms underlying such spatial control of chemical reactions and how much the properties of chemical reactions are altered by the compartments relative to homogenous systems. Here, we derive a theoretical framework to study fuel driven chemical reactions in the presence of compartments. We study two state transitions like phosphorylation via hydrolysis of ATP and enzymatic reactions. For two state transitions, we find that the ratio of phosphorylated product can be regula-ted by droplets by two orders of magnitude relative to the homogenous state. In the case of enzymatic reac-tions, we show that the initial rate of product formation can be increased by more than ten fold. We further calculate analytically the optimal conditions of designing the system. Our studies exemplify the enormous potential of phase separated compartments as biochemical reactors in living cells and enhancing the effect of enzymes. Understanding the control of biochemical reactions via compartments is key to elucidate the func-tionality of stress granules for the cell and is also crucial for biochemical communication among synthetic cells and RNA catalysis in coacervate protocells.

1 Considerations and challenges in studying liquid-liquid phase separation and biomolecular condensates, Tanja Mittag et al., Cell, 2019

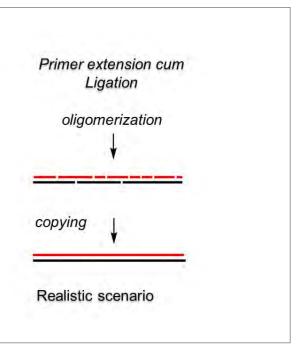


# BRIDGING THE GAP BETWEEN CHAIN FORMATION & GENETIC **COPYING OF RNA**

Gabrielle Leveau<sup>1</sup>, Tobias Göppel<sup>2</sup>, Daniel Pfeffer<sup>1</sup>, Joachim Rosenberger<sup>2</sup>, Ulrich Gerland<sup>2</sup>, Clemens Richert<sup>1</sup>

<sup>1</sup> Institute of Organic Chemistry, University of Stuttgart <sup>2</sup>Department of Physics, Technical University Munich

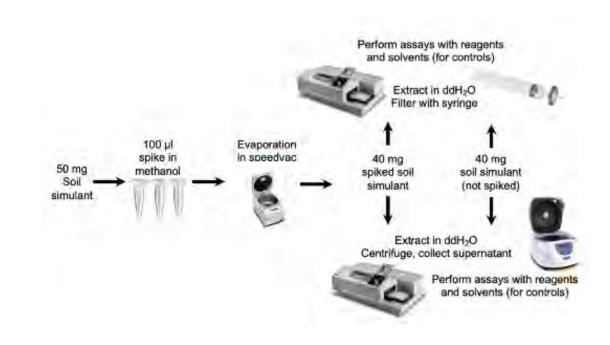
genetic information.



The copying of RNA sequences is the template-directed reaction underlying replication. It is interesting to ask how such a process may have occurred in a prebiotic context to ensure the transmission of

A minimal living system, from which more complex systems can arise via Darwinian evolution, must be able to replicate its genetic information. Steps toward such a minimal system should include the formation of oligonucleotide chains and their copying through the formation of complementary strands. Thus far, creating a minimal evolving system in the laboratory has remained elusive for RNA, and there are gaps in our understanding of this aspect of prebiotic evolution. One prominent gap is between the formation of short oligonucleotides from nucleotides and the spontaneous emergence of a copying process that can maintain and propagate the sequence information encoded in a minimal genome. Experiments in the absence of a polymerase show a sequence fidelity too low for inheriting even short genes. They are also not prebiotically plausible, given that they rely on one primertemplate complex that is extended by monomers only, as in modern-day PCR reactions, and not a statistical mixture of different strand lengths expected for oligomerization products. Still, selective and efficient incorporation of activated monomers has been demonstrated, using immobilized template/primer systems.1 We are expanding this work to enzyme-free ligation of dimers or trimers to an RNA primer using in situ activation conditions.<sup>2</sup> A quantitative modeling of RNA copying is also being conducted. The challenge to achieve replication of a short RNA sequence, a feat recently accomplished for 3'-amino-2',3'dideoynucleotides and an initial DNA template,3 will be discussed.

[1] Deck, C.; Jauker, M.; Richert, C. Nat. Chem. 2011, 3, 603. [2] Sosson, M.: Pfeffer, D.: Richert, C. Nucleic Acids Res. 2019, 47, 3836. [3] Hänle, E.; Richert, C. Angew. Chem. 2018, 130, 9049.

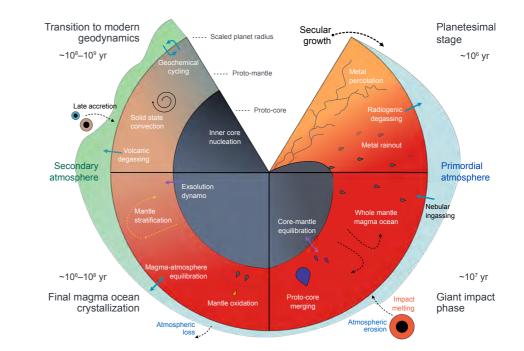


## **BIOCHEMICAL METHODS** FOR THE DETECTION OF PROTEINS AS LIFE **SIGNATURES ON** MARS-LIKE SOILS



### Yongda Li, David Collins, Konstantinos Grintzalis School of Biotechnology, Dublin City University, Dublin, Ireland

One of the principal objectives for planetary exploration is the search for traces of past life and evidence of conditions that may have supported life. Such analytical approaches often rely on the detection of specific markers (i.e. biomolecular signatures) and require the development of highly specialised methods and complex automated analytical platforms. The high cost of such missions is matched by the high risk of failure (i.e. mechanical failure, crash landing, loss of vehicle during launch, loss of contact, instrument failures, etc). Development of simple, highly specific, and conclusive assays with a minimum number of steps would mitigate many of the risks and improve the efficacy of such explorative tests. Astrobiological research is also oriented in Marslike soil samples on Earth which provide an analogue for experimental design and simulation. In this study, we present simple methods to quantify proteins on Martian soil simulants with a long-term goal the development of a fully automated platform for future exploration missions



# ATMOSPHERIC **SPECIATION OF ROCKY** PLANETS FROM MAGMA **OCEAN OUTGASSING**



Tim Lichtenberg<sup>1</sup>, Dan J. Bower<sup>2</sup>, Mark Hammond<sup>3</sup>, Shang-Min Tsai<sup>1</sup>, Paolo A. Sossi4

Atmospheric, Oceanic and Planetary Physics, University of Oxford <sup>2</sup> Center for Space and Habitability, University of Bern <sup>3</sup> Department of the Geophysical Sciences, University of Chicago <sup>4</sup> Institute of Geochemistry and Petrology, ETH Zurich

1 Bower, D. J., Kitzmann, D., Wolf, A. S., et al. (2019). Linking the evolution of terrestrial interi-ors and an early outgassed atmos to astrophysical observations. Astron. Astrophys. 631, A103 2 Zahnle, K., Schaefer, L., Fegley, B. (2010). Earth's earliest atmospheres. Cold Spring Harbor perspectives in biology, 2, a004895

3 Bonati, I., Lichtenberg, T., Bower, D. J., et al. (2019). Direct imaging of molten protoplanets in nearby young stellar associations. Astron Astrophys. 621, A125

4 Kreidberg, L., Koll, D. D., Morley, C., et al. (2019). Absence of a thick atmosphere on the terres-trial exoplanet LHS 3844b.Nature 573

5 Hamano, K., Abe, Y., Genda, H. (2013). Emergence of two types of terrestrial planet on solidifi-cation of magma ocean. Nature 497, 607-610

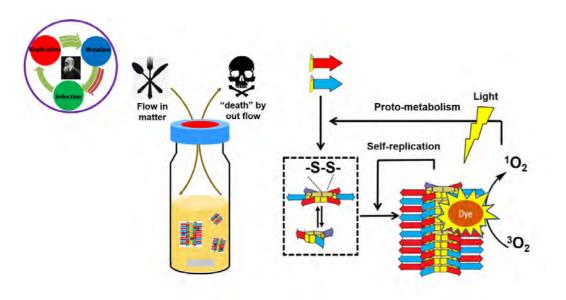
6 Benner, S. A., Bell, E. A., Biondi, E., et al. (2019). When Did Life Likely Emerge on Earth in an RNA First Process? ChemSystemsChem. 7 Sasselov, D. D., Grotzinger, J. P., Sutherland, J. D. (2020). The origin of life as a planetary phe-nomenon. Sci. Adv. 6, eaax3419.

cycles<sup>2</sup>.

The earliest atmospheres of rocky planets originate from extensive volatile release during one or more magma ocean epochs that occur during primary and late-stage assembly of the planet<sup>1</sup>. These epochs represent the most extreme cycling of volatiles between the interior and atmosphere in the history of a planet, and establish the initial distribution of the major volatile elements (C, H, N, O, S) between different chemical reservoirs that subsequently evolve via geological

Crucially, the erosion or recycling of primary atmospheres bear upon the nature of the long-lived secondary atmospheres that will be probed with current and future observing facilities3. Furthermore, the chemical speciation of the atmosphere arising from magma ocean processes can potentially be probed with present-day observations of tidallylocked rocky super-Earths<sup>4</sup>. The speciation in turn strongly influences the climatic history of rocky planets, for instance the occurrence rate of planets that are locked in long-term runaway greenhouse states<sup>5</sup>. We will present an integrated framework to model the build-up of the earliest atmospheres from magma ocean outgassing using a coupled model of mantle dynamics and atmospheric evolution. We consider the diversity of atmospheres that can arise for a range of initial planetary bulk compositions, and show how even small variations in volatile abundances can result in dramatically different atmospheric compositions. We will discuss our results in light of the prospects for untangling the diversity of rocky planetary atmospheric compositions and their potential effects on the redox state of the earliest mantle geochemistry and atmospheric speciation relevant for surficial prebiotic chemical environments6,7.

Only through the lense of coupled evolutionary models of terrestrial interiors and atmospheres can we begin to deconvolve the imprint of formation from that of evolution, with consequences for how we interpret the diversity revealed by astrophysical observables, and their relation to the earliest planetary conditions of our home world.



# TOWARDS DARWINIAN **EVOLUTION OF SYNTHETIC** REPLICATORS



k.liu@rug.nl

Kai Liu, Sijbren Otto

Centre for Systems Chemistry, Stratingh Institute, University of Groningen, Nijenborgh 4, 9747 AG Groningen, The Netherlands

NASA's working definition for life is "a self-sustaining chemical system capable of Darwinian evolution"1. In biology, Darwinian evolution can be considered to be the result of an interplay of replication, mutation, and selection<sup>2</sup>. These concepts can be extended to chemical systems. Otto group has realized exponential self-replication through elongation/ breakage of a fibre<sup>3</sup>. Starting from two different building blocks, diversification of replicators is achieved through cross-catalysis4. Recently we find replicator can recruit and activate a photocatalytic cofactor, catalysing the synthesis of its own precursors to promote replication5. The photocatalysis is presumed to work as "encoded function" that may be selected in a replication-deconstruction regime. When mutant replicators with different catalytic ability are subjected to flow condition, most photoactive ones become dominant, reminiscent of "survival of the fittest".

1 Ruiz-Mirazo, K., Peretó, J. & Moreno, A. Origins. Life. Evol. B. 34, 323-346 (2004). 2 Higgs, P. G. J. Mol. 84, 225-235 (2017).

3 Colomb-Delsuc, M., Mattia, E., Sadownik, J. W. & Otto, S. Nat. Commun. 6, 1-7 (2015). 4 Sadownik, J. W., Mattia, E., Nowak, P. & Otto, S. Nat. Chem. 8, 264 (2016). 5 Santiago, G. M., Liu, K., Browne, W. R. & Otto, S. ChemRxiv (2019), doi.org/10.26434/ chemrxiv 10002122 v1

ANOMALOUS FLUCTUATIONS AND **SELECTIVE EXTINCTION** IN PRIMORDIAL **REPLICATORS:** A "STRUGGLE FOR LIFE" AT THE ORIGIN OF **BIOLOGICAL CHIRALITY** 

1..... ...... ...... 11..1.......

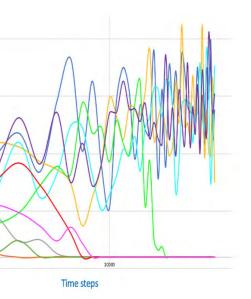
.....10...00...... 

...... 



Savino Longo<sup>1,2</sup>, Gaia Micca Longo<sup>1,2</sup>, Miriana Carmela Chincoli <sup>1</sup> Department of Chemistry – Università degli Studi di Bari Aldo Moro – Via Orabona 4 - 70125 Bari, Italy

<sup>2</sup> Istituto per la Scienza e Tecnologia dei Plasmi – Consiglio Nazionale delle Ricerche, Bari Section – Via Amendola 122/D - 70125 Bari, Italy



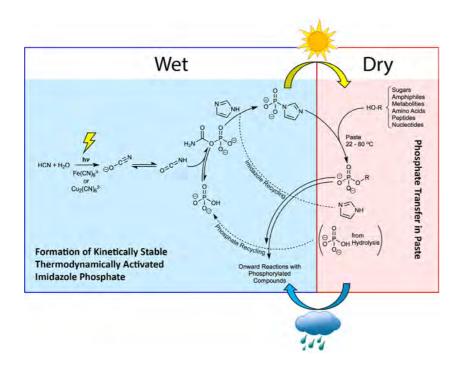
The prevalent presence of a single chiral variant of molecules in live organisms is one of the most distinctive signs of life as a global phenomenon. One of the greatest ambitions of Biochemistry and Astrobiology is to provide an explanation of this predominance. Several mechanisms were proposed in the past, based on the "propagation" of chirality from a homo-chiral substrate and the amplification of effects associated with electro-weak interaction. Here, a different scenario is proposed: anomalous fluctuations associated with a self-replication scenario can lead to selective extinction of one of the two variants<sup>1</sup>. These fluctuations arise spontaneously when a global (not a local one) feedback acts. The idea is based on two key-points: a) the simulation of prebiotic processes as a "chessboard play"<sup>2</sup>; b) the presence of great fluctuations during an autocatalytic process<sup>3</sup>. In order to demonstrate this mechanism, a computational model is developed, describing the "struggle for life" of two different kinds of primordial replicators in a n×m chessboard, with a periodic contour; each replicator employs catalyzers of different chirality but on a non-chiral substrate, thereby with no selective

advantage. The replication occurs randomly and with a fixed probability, providing that a sufficient amount of chemical energy is locally available. Results clearly show that strong fluctuations in the number of individuals of each species and a subsequent selective extinction of one of the two are observed. These studies may contribute to shed light on a most mysterious phase transition occurred during the biochemical evolution of our planet.

1 Longo, S., & Coppola, C. M. (2013) Rendiconti Lincei 24(3), 277-281. 2 Eigen. M., & Winkler, R. (1993). Laws of the game: how the principles of nature govern chance (Vol. 10). Princeton University Press

3 Prigogine I (1981) From being to becoming, time and complexity in the physical

sciences. W. H. Freeman & Co, New York



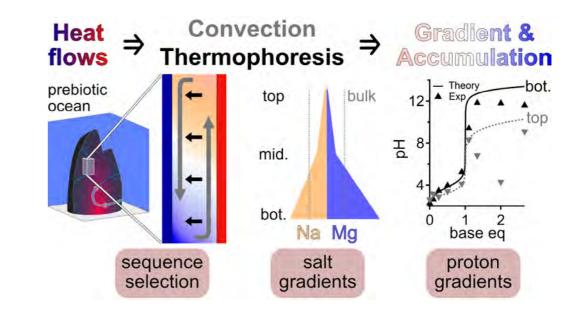
# CYCLING OF ORTHOPHOSPHATE UNDER MILD PREBIOTICALLY PLAUSIBLE CONDITIONS



Oliver R. Maguire, Iris B. A. Smokers, Wilhelm T. S. Huck Radboud University Nijmegen, Institute for Molecules and Materials, Hevendaalseweg 135, 6525 AJ Nijmegen, The Netherlands

Phosphate-based chemistry is crucial to Life and extant Life is able to continuously activate, transfer and recycle orthophosphate in order to drive cellular biochemistry. However, in prebiotic chemistry the activation of orthophosphate from geochemical sources and subsequent transfer to prebiotically important organic compounds under mild conditions remains an outstanding fundamental challenge. Considering the paucity of prebiotically plausible phosphorylating reagents and reaction conditions that can directly activate and transfer phosphate in a single reaction step, we sought an alternative primordial scenario whereby the phosphate activation step and phosphate transfer step are separated. Conversion of an activated form of phosphate into a kinetically stable thermodynamically activated molecule would enable the accumulation of an activated form of phosphate in solution. Subsequent drying down of the solution into a paste could then enables the transfer of the phosphate.

Here, we demonstrate that we can use isocyanate to activate orthophosphate and store the energy in the phosphoramidate imidazole phosphate. Initiation of a wet/dry cycle enables transfer of phosphate to a diverse range of prebiotically important organic compounds. Upon re-wetting, orthophosphate can be reactivated, and the cycle repeated. This approach enables orthophosphate to be continuously recycled and converted back into an activated form under mild conditions thus establishing a prebiotic analogue of the system by which extant Life cycles orthophosphate.



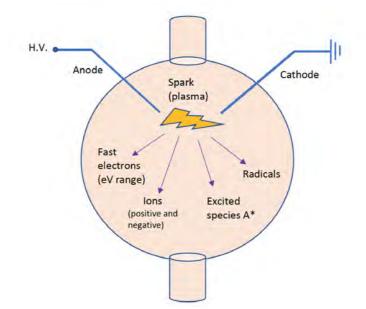
# HEAT FLOWS SHIFT CHEMICAL EQUILIBRIA BY SELECTIVE ACCUMULATION



Thomas Matreux<sup>1</sup>, Alexandra Kühnlein<sup>1</sup>, Johannes Raith<sup>2</sup>. Kristian Le Vav<sup>3</sup>. Dieter Braun<sup>1</sup>, Christof B. Mast<sup>1</sup> Systems Biophysics, LMU Munich, Amalienstraße 54, 80799 München <sup>2</sup>Gerland group, TU München, James-Franck-Straße 1, 85748 Garching <sup>3</sup>MPI of Biochemistry, Martinsried (Munich)

and their constituent phases with a feedstock of simple molecules. Our aim is to combine this scenario with thermal non-equilibrium and bring together geomaterials, chemistry and microfluidics in a realistic environment. The reaction chambers are sandwiched between highly heat conducting sapphire plates ensuring complete thermal control including possible thermal gradients. Microfluidic structures are made from FEP, which lets us focus on the interactions between the molecules. Ions leached from prebiotically plausible mineral samples are selectively accumulated by thermal gradients and permit enzymatic activity. Thermal nonequilibrium boundary conditions drive concentration gradients, enabling chemical reactions and generating and controlling pH gradients in a plausible prebiotic scenario. Local gradients driven by heat fluxes will offer unique opportunities to enable molecular selection and evolution at the origins of life.

The first steps in the emergence of life on Earth occurred on rocks



### NEW INSIGHTS ON PREBIOTIC CHEMISTRY FROM PLASMA KINETICS



### Gaia Micca Longo<sup>1,2</sup>, Vincenzo Laporta<sup>2</sup>, Savino Longo<sup>1,2</sup>

Department of Chemistry - Università degli Studi di Bari Aldo Moro – Via Orabona 4 – 70125 Bari, Italy <sup>2</sup> Istituto per la Scienza e Tecnologia dei Plasmi – Consiglio Nazionale delle Ricerche, Bari Section – Via Amendola 122/D - 70125 Bari, Italy

The famous Miller-Urey experiment<sup>1</sup>, which provides essential insight on the prebiotic synthesis of the molecules of life, still has many obscure points. Although the theoretical studies on Miller's experiment are very advanced<sup>2</sup>, one aspect of these studies is still rather primitive, namely the role of electrons in the plasma in producing the first excited and radical species and ions, which later enter several chemical channels to form prebiotic molecules. Here, we want to suggest a way of possible future progress: framing the experience of Miller and Urey in the context of the kinetics of ionized gas, or plasma. In this context, effective and versatile theoretical tools, based on quantum mechanics and chemical kinetics, make it possible to look, in a new way, at the elementary processes that lead to the formation of excited species and ions, at the origin of the cascade of subsequent reactions.

Two new elements may produce a synergistic push towards further progresses: a) the awareness that the primordial atmosphere was not at all the strongly reducing mixture believed in Miller's times; b) the development of new methods in the context of computer modeling of the kinetics of plasmas. The communication between the two communities of plasma kinetics and astrobiology can therefore help, in the future, to attain a better understanding and new insights on the chemical kinetics of an historical experiment, which has changed our ideas on the genesis of prebiotic molecules on the primordial Earth.

1 Miller, S.L., and Urey, H.C. (1959) Science 130 (3370), 245-251. 2 Saitta, A.M., and Saija, F. (2014) Proceedings of the National Academy of Sciences 111(38), 13768 - 13773.[5] Schwarz, R.-J.; Richert, C., Nanoscale 9, 7047-7054 (2017).

### INDEX - FLOOR 8 Aligns with meeting schedule SAIBAL MITRA The first step from molecules to life: Formation of large random molecules acting as microenvironments MAITANE MUÑOZ BASAGOITI Physics and evolution of catalysts ATIDA NASUFOVSKA Prebiotic chemical energy flux JOANA PEREIRA On the origins of the protein world: A large-scale computational approach to study the emergence of the first autonomously folding proteins **BENEDIKT PETER & ELIA SALIBI** Freeze-thaw driven proliferation of RNA protocells MARTINA PREINER The ambivalent role of water at the origins of life KHOSROW RAHIMI Amphiphilic comb-polymers solve the dilemma of polymer-based cell-mimetic membranes MEHRNOUSH RAHIMZADEH Structural design of amphiphilic comb polymers to self-assemble into faceted membrane protocells ANA MARIA RESTREPO SIERRA An evolutionary approach for building a synthetic cell PAULA CATALINA RODRIGUEZ RAMIREZ Linking microbial diversity to carbon cycling in subseafloor sediments from the Namibian continental shelf

110

111

112

113

114

115

116

117

118

119

120 CESAR RODRIGUEZ-EMMENEGGER Superselectivity in synthetic protocells



# THE FIRST STEP FROM MOLECULES TO LIFE: FORMATION OF LARGE RANDOM MOLECULES ACTING AS MICRO-**ENVIRONMENTS**

Saibal Mitra

The processes that led to life had to circumvent the limit on the complexity of low fidelity replicating systems<sup>1</sup>. I have proposed a three step process to address thus problem<sup>2</sup>. The first step takes place in a space environment where in ice grains large random molecules form under the influence of UV radiation and cosmic radiation. The structure of these molecules are those of 3 dimensional percolation clusters which are known to have a fractal structure<sup>3,4</sup>. Such molecules of a typical size of 100 nm have a porous structure that are permeable to small molecules.

The second step involves the formation of loosely bound aggregates of such random molecules on proto-planets in the early solar system. The third step involves biochemical processes taking place inside these aggregates. One can then consider conventional models of the origin of life inside the micro-environment within such aggregates instead of the raw outside environment. The fidelity problem is then addressed by the effective compartmentalization and fixed surface structures having a large effect on the biochemical due to the small size and the fractal structure of the environment.

The loosely bound aggregate will gradually erode, which will cause the micro-environment to gradually resemble the outside environment, providing for a mechanism for the biochemical system to gradually adapt to the outside environment.

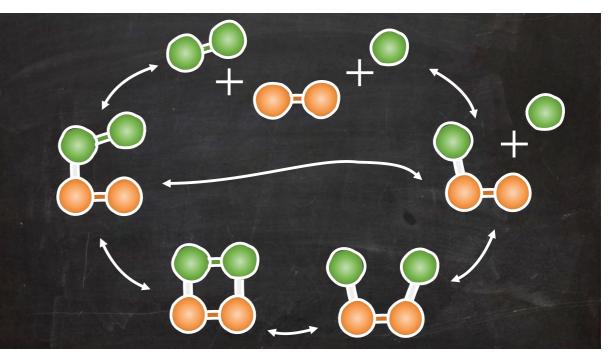
1 Eigen, M., 1971, Self-organization of matter and the evolution of biological macromolecules. Naturwissenschaften 58, 465-523

2 Mitra. S., 2018. Percolation clusters of organics in interstellar ice grains as the incubators of life, Progress in Biophysics and Molecular Biology 149, 33-38.

3 Wang, J., Zhou, Z., Zhang, W., Garoni, T.M., Deng, Y., 2013. Bond and site percolation in three dimensions. Phys. Rev. E 87, 052107.

4 Wang, J., Zhou, Z., Zhang, W., Garoni, T.M., Deng, Y., 2014. Erratum: bond and site percolation in three dimensions. Phys. Rev. E 89, 069907.

### FLOOR 8



### PHYSICS AND EVOLUTION OF CATALYST



Maitane Muñoz Basagoiti<sup>1</sup>, Olivier Rivoire<sup>2</sup>, Zorana Zeravcic<sup>1</sup> <sup>1</sup> Gulliver Lab UMR 7083, ESPCI PSL Research University, Paris, France <sup>2</sup>Center for Interdisciplinary Research in Biology (CIRB), Collège de France, CNRS, INSERM, PSL Research University, Paris, France

Enzymes are the unsurpassed catalysts of Nature, operating under mild conditions to produce reaction rate enhancements of several orders of magnitude<sup>1</sup>. Their remarkable catalytic activity is attributed to the high complementarity between the enzyme's active site and the transition state of the catalyzed reaction, a feature that has inspired the design of artificial enzymes and transition state analogs<sup>2</sup>. Despite outstanding progress in the field, the driving forces underlying catalysis still remain unclear<sup>3</sup>. Advances in singlemolecule enzymology provide unprecedented information of enzyme dynamics and catalytic mechanisms<sup>4,5</sup>, making it timely to analyze the design principles and fundamental constraints of catalysts.

In our work, we search for the geometrical and physical constraints necessary for the emergence of catalytic activity in a system of DNAcoated colloids. We use coarse-grained computer simulations to build structures of increasing complexity with spherical colloids. Our goal is to identify the simplest structure with the potential to catalytically cleave a bond. 2D simulations show that a rigid dimer structure can provide a bond-breaking mechanism by binding strongly to the substrate and forming a rhomboidal configuration. We explore the optimal trade-off between strong binding and product release achieved in catalysts<sup>3</sup>.

1 Wolfenden, R. et al., Acc. Chem. Res. 2001, 34, 938-945 2 Breslow, R., Acc. Chem. Res. 1995, 28, 146 3 Swiegers, G. et al., Chem. Eur. J. 2009, 15. 4746-4759 4 Lu, H. P. et al., Science 1998, 282, 1877-1882 5 Min. W. et al., Acc. Chem. Res. 2005, 38, 923-931

### PREBIOTIC CHEMICAL ENERGY FLUX



A. Nasufovska<sup>1</sup>, F. Meyer<sup>2,3</sup>, U. Diederichsen<sup>\*1,3</sup>, H.-J. Fritz<sup>1,3</sup>

\*Institute of Organic and Biomolecular Chemistry, Georg-August University Göttingen

<sup>2</sup>Institute of Inorganic Chemistry, Geora-August University Göttingen <sup>3</sup>Academy of Science and Humanities, Göttingen \* Tammannstr. 2, D-37077 Göttingen

Life on planet Earth may have started at sites of volcanic activity like deep sea hydrothermal vents, feeding on chemical energy of ejected matter in disequilibrium.<sup>1,2</sup> The project presented here explores flux of chemical energy under modelled primordial conditions which may have laid the foundation for biological energy metabolism prior to cellular life. In particular, we study energetic ramifications of the NiS-catalyzed carbonylation of thiomethanol yielding acetic acid, a possible prebiotic carbon fixation reaction.<sup>2</sup> It is hypothesized that energy-rich intermediates (acetyl nickel species, methylthioacetate and possibly others) are evolutionary precursors of analogous intermediates in still prevailing metabolic pathways.<sup>3,4</sup> The poster illustrates experiments to determine formation and hydrolysis rates of key intermediates that are likely candidates for having served as gateways to the "thioester world"4 and/or the establishment of a versatile biological "energy currency" like ATP or evolutionary precursors thereof.

1 J.B. CORLISS, J. DYMOND, L. I. GORDON, J. M. EDMOND, R. P. VON HERZEN, R. D. BALLARD, K. GREEN, D. WILLIAMS, A. BAINBRIDGE, K. CRANE, T. H. VAN ANDEL, Science, 1979, 203, 1073. 2 C. HUBER, G. WÄCHTERSHÄUSER, Science, 1997, 276, 245-247. 3 M. CAN, F. A. ARMSTRONG, S. W. RAGSDALE, Chemical Reviews, 2014, 114 (8). 4149-4174. 4 SOUSA, F.L., THIERGART, T., LANDAN, G., NELSON-SATHI, S., PEREIRA, I.A., ALLEN, J.F., LANE, N. AND MARTIN, W.F. (2013) Early bioenergetic evolution. Philosophical Transactions of the Royal Society of London, Series B. Biological Sciences 368, 20130088. 5 CH. DE DUVE, Blueprint for a Cell - The Nature and Origin of Life, Neil Patterson Publishers, Ed. 1 1991 275

### FLOOR 8

# ON THE ORIGINS OF THE **PROTEIN WORLD: A LARGE** -SCALE COMPUTATIONAL APPROACH TO STUDY THE EMERGENCE OF THE FIRST AUTONOMOUSLY FOLDING PROTEINS

### Joana Pereira, Andrei N. Lupas

Department for Protein Evolution. Max Planck Institute for Developmental Biology, Max-Planck-ring 5, 72076 Tübingen, Germany

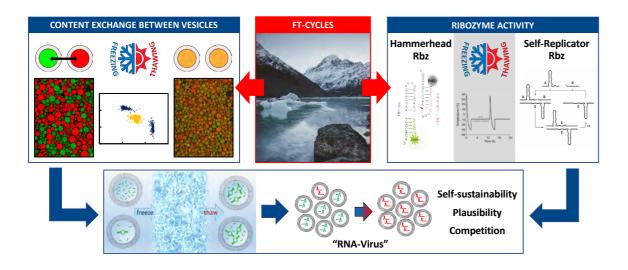
folding proteins.

Since the time of the Last Universal Common Ancestor (LUCA), proteins have been the fundamental catalysts of life. For their activity they must assume three-dimensional structures by a complex, easily disrupted, process of folding. However, it is still unclear how the first folded proteins emerged and how life came to rely so extensively on their ability to fold. Our hypothesis is that the first folded proteins resulted from the increased complexity of peptides in the "RNA-peptide world" that preceded LUCA, possibly by three mechanisms<sup>1,2</sup>: repetition, accretion, and recombination. While repetition is one of the most common mecha-nisms for the emergence of new folded proteins3, and accretion could already be traced to a few ancient folds<sup>4</sup>, recombination is a mechanism harder to trace.

Instead of searching for examples of folds that could have their origins in the recombination of at least two ancient fragments, we followed a large-scale computational approach to study whether two such fragments could generate a folded protein when recombined and excluded from their original scaffolds. Using the ribosome as a model of the primordial "RNA-peptide world"1, we collected a set of ribosomal peptide fragments, which are only folded in the context of the ribosome, and followed an all-against-all molecular docking approach to evaluate their propensity to establish geometrically and energetically compatible inter-faces that would allow the formation of stable, globular, recombinant folds in the absence of the RNA. As a result, we identified multiple ribosomal peptide frag-ment pairs that can recreate not only frequent protein folds but also novel fold topologies and further optimised some of these folds by exploring the sequences of their parent fragments in different organisms. From these, we selected two pairs that are now being experimentally characterised, opening a door to a better understanding of the emergence of the first autonomously

1 A. N. Lupas and V. Alva, J. Struct. Biol. 48, 103-109 (2017) 2 V. Alva and A. N. Lupas, Curr. Opin. Struct. Biol. 48, 103-109 (2018) 3 H. Zhou et al. El ife 5, 551-560 (2016) 4 J. Pereira and A. N. Lupas, Bioinformatics 34(23), 3961-3965 (2018)

0



# FREEZE-THAW DRIVEN PROLIFERATION OF **RNA PROTOCELLS**

### Elia Salibi, Benedikt Peter, Hannes Mutschler, Petra Schwille Max Planck Institute of Biochemistry,

Martinsried (Munich), Germany

early protocells able to grow and proliferate without the sophisticated protein machineries found in modern forms of life? Based on previous studies of genetic exchange between giant unilamellar vesicles (GUVs), we seek to apply repeated freeze-thaw (FT) cycles as a physicochemical driver for the expansion of encapsulated selfreplicating ssRNA enzymes (ribozymes). This system will serve as a model for the growth and proliferation of RNA protocells in the plausible geochemical environment of early Earth exhibiting diurnal freezing of water to ice.

With the advent of compartmentalization in the RNA world, how were

Litschel T, Ganzinger KA, Movinkel Torgeir, Heymann M, Robinson T, Mutschler H, Schwille P (2018) New J Phys 20:055008 Paudel BP, Fiorini E, Börner R, Sigel RKO, Rueda DS (2018) PNAS 115:11917-11922

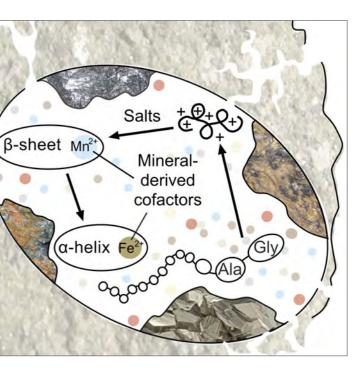




Andrey do Nascimento Vieira<sup>1</sup>, Karl Kleinermanns<sup>2</sup>, William F. Martin<sup>1</sup>, Martina Preiner<sup>1</sup>

<sup>1</sup> Institute for Molecular Evolution, University of Düsseldorf, 40225 Düsseldorf, Germany <sup>2</sup> Institute for Physical Chemistry, University of Düsseldorf, 40225 Düsseldorf, Germany

self-organisation5.



Without water as a solvent and reactant, life as we know it would not exist. However, water molecules can also counteract the formation of essential organic molecules due to hydrolysis. This conundrum constitutes one of the central issues in origin of life research<sup>1</sup>. Hydrolysis is an important part of energy metabolism but only because inside a cell, it is a controlled reaction. How could hydrolysis have been regulated under prebiotic settings? Lower water activities possibly provide an answer. Geochemical sites with less free and more bound water can supply the necessary conditions for protometabolic reactions. Such conditions occur in serpentinizing systems, hydrothermal sites that synthesise hydrogen gas via rock-water interactions<sup>2,3</sup>. We summarise the parallels between biotic and abiotic means of controlling hydrolysis in order to narrow the gap between biochemical and geochemical reactions<sup>4</sup>, and outline how hydrolysis could even have played a constructive role at the origin of molecular

1 Westall F & Brack A (2018) The importance of water for life. Space Sci. Rev. 214, 1-23. 2 Lamadrid HM, Rimstidt JD, Schwarzenbach EM, Klein F, Ulrich S, Dolocan A & Bodnar RJ (2017) Effect of water activity on rates of serpentinization of olivine. Nat. Commun. 8, 16107. 3 Preiner M, Xavier J, Sousa F, Zimorski V, Neubeck A, Lang S, Greenwell H, Kleinermanns K,

Tüysüz H, McCollom T, Holm N & Martin W (2018) Serpentinization: connecting geochemistry, ancient metabolism and industrial hydrogenation. Life 8, 41. 4 Preiner, M., Igarashi, K., Muchowska, K. B., et al. (2020) 'A hydrogen dependent geochemical

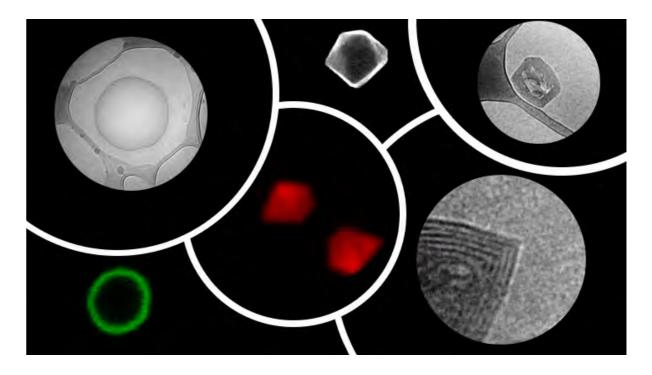
analogue of primordial carbon and energy metabolism', Nat. Ecol. Evol. 4, 534–542. 5 do Nascimento Vieira, A., Kleinermanns, K., Martin, W. F., Preiner, M. (2020) The ambivalent role

of water at the origins of life, FEBS Letters, in press.

# AMPHIPHILIC COMB-POLYMERS SOLVE THE **DILEMMA OF POLYMER-**BASED CELL-MIMETIC MEMBRANES

Khosrow Rahimi, Mehrnoush Rahimzadeh, Jan Tenbusch, Nina Kostina, Cesar Rodriguez-Emmenegger DWI-Leibniz Institute for Interactive Material, Aachen, Germany

Natural membranes achieve an incredibly rich functionality by the selfassembly of different components at an almost invariable thickness (5±1 nm). Remarkably, in spite of their minute thickness and flexibility, natural membranes are incredibly stable. The combination of these seemingly antagonistic properties makes membranes a key for life to exist. The thickness and flexibility has been mimic by assembly of lipids into synthetic vesicles call liposomes. Nonetheless, liposome lack stability to environmental conditions, severely limiting their use for advance functions. Polymersomes from amphiphilic block copolymers display a much enhanced mechanical stability, but at the expense of thickness well above the natural ones and an almost complete stall of the dynamics compared to biological membranes. Furthermore, the mismatch between the membrane thickness and the size of transmembrane proteins has been the main obstacle hampering the integration of natural bioreceptors in polymersomes. Here, I present our advances in cell mimetic membranes based on amphiphilic comb polymer. The polymers consist of a hydrophilic highly flexible backbone to which fatty-acid-like side groups are appended. The latter drive the zipping of the polymer chains into bilayers. We developed an accelerated iterative combinatorial synthesis to generate a library with systematic structural variation. This allowed us to elucidate how to program the thickness, stability and flexibility in the molecular structure and topology of the polymer and in this way solve the dilemma of combine stability with extreme flexibility and biomimetic thickness. Contrary to block copolymer, no entanglement of hydrophobic domains occurs, thus the thickness and flexibility of our membrane mimic closely matches those of their natural counterparts. This is demonstrated by structural analysis of vesicles as well as by the insertion of transmembrane proteins. Our model hold promise for the design of interactive protocells.



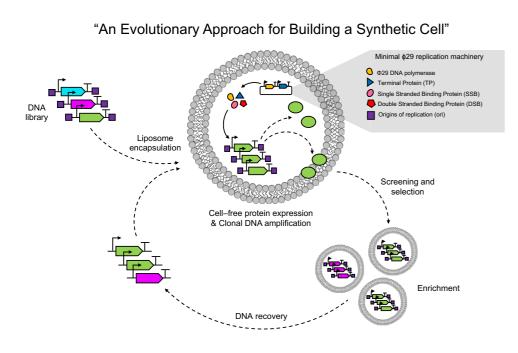
### STRUCTURAL DESIGN OF AMPHIPHILIC COMB POLYMERS TO SELF-ASSEMBLE INTO FACETED MEMBRANE PROTOCELLS



Mehrnoush Rahimzadeh, Jan Tenbusch, Khosrow Rahimi, Nina Kostina, Cesar Rodriguez-Emmenegger DWI-Leibniz Institute for Interactive Materials and Institute of Technical and Macromolecular Chemistry. rodriguez@ dwi.rwth-aachen.de; rodriguezemmenegger-lab.com

nature.

Compartmentalization by self-assembly of lipids into membrane is a key element to the origin of life. Various synthetic and hybrid mimics have been developed based on the self-assembly of lipids or amphiphilic block copolymers aiming at recapitulating some properties of spherical membranes. However, other more complex morphologies are also present in nature. Particularly interesting are faceted membranes, in which the Gaussian and mean curvature of the membrane are zero like in Haloquadratum archaea, where cells are perfectly prismatic. Recapitulating such membranes requires a fine balance between surface energy, conformational entropy, and order, as the formation of sharp edges is extremely unfavorable in soft matter. To mimic this morphology, we developed a new family of amphiphilic comb-copolymers consisting of a poly(N-vinylacetamide) hydrophilic backbone to which we grafted hydrophobic side alkyl chains. However, to achieve faceted vesicle it is necessary to device a two-step selfassembly procedure. At temperatures above crystallization temperature of the side chains, the segregation between the backbone and side chains drives the assembly into bilayers of biomimetic thickness. By cooling the sample, the crystallization of side chains under the confinements of the membrane, forces a shape transformation from the spherical into polyhedral vesicles. The thermodynamic conditions for this transition are encoded in the side groups. Polymers with long alkyl chains (≥C18) in their extended and frozen state crystallize inside the bilayers in a planar hexagonal lattice. Since this packing is incompatible with spherical shape, it results in topological defects of various folds forming multiple facets. Indeed, comb polymers with low degree of substitution form faceted vesicles if the crystallization forces are strong enough (e.g. for C20). Our system hold promises to further elucidate the mechanisms behind the formation of complex living membranes in



# AN EVOLUTIONARY **APPROACH FOR BUILDING A SYNTHETIC** CELL

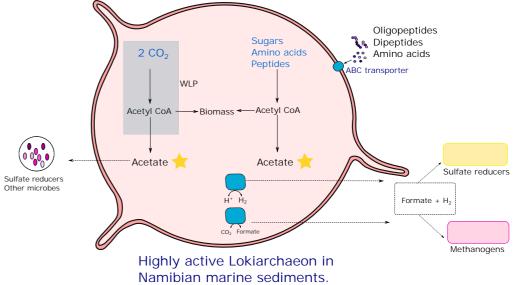


### Ana M. Restrepo, Zhanar Abil, **Christophe Danelon**

Department of Bionanoscience, Kavli Institute of Nanoscience, Delft University of Technology, van der Maasweg 9, Delft 2629 H7 The Netherlands

The goal of bottom-up synthetic biology culminates in the construction and engineering of a 'living' artificial cell. This grand challenge can be broken down into the reconstitution of essential cellular functions into an enclosing membrane, such that autonomous life-like properties as growth or division, will emerge. We propose to complement traditional strategies with an in vitro directed evolution approach, as an optimization scheme to implement the different fundamental biological modules. Specifically, in-liposome cell-free gene expression coupled with isothermal DNA amplification forms the basic unit for our evolutionary experiments. The PURE system is used as a minimal transcription-translation apparatus, while protein-primed DNA replication is performed by the Phi29 machinery. We established conditions for driving the function of the encoded protein when coupled to orthogonal DNA replication. Cases of study include auto fluorescent proteins, phospholipid-synthesizing enzymes and Phi29 replication proteins. We believe that when combined with the compartmentalized expression of a gene library, our directed evolution approach will accelerate the engineering of complex biological functions and, ultimately, of a synthetic cell.

Van Nies, P. et al. Self-replication of DNA by its encoded proteins in liposome-based synthetic cells, Nat Commun 9, (2018).



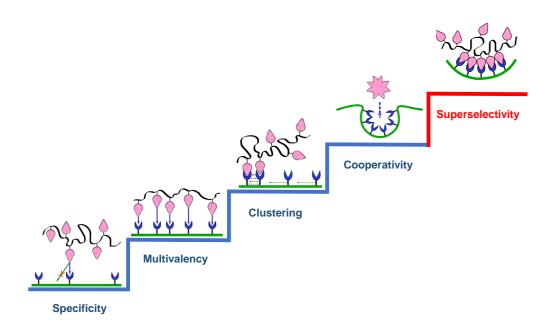
### LINKING MICROBIAL **DIVERSITY TO CARBON** CYCLING IN SUBSEAFLOOR SEDIMENTS FROM THE NAMIBIAN CONTINENTAL SHELF

### Paula Rodríguez<sup>1</sup>, Ömer K. Coskun<sup>2</sup>, Aurèle Vuillemin<sup>2</sup>, William D. Orsi2,3

<sup>1</sup> Department of Earth Sciences, Geobiology Group, Swiss Federal Institute of Technology Zürich, 8092, Zürich Switzerland

<sup>2</sup> Department of Earth and Environmental Sciences, Paleontology & Geobiology, Ludwig- Maximilians-Universität München, 80333 Munich, Germany. <sup>3</sup> GeoBio-CenterLMU. Ludwig-Maximilians-Universität München, 80333 Munich, Germany.

The Benguela Upwelling System (BUS) is one of the most productive ecosystems in the world (0.37 Gt carbon/year). This oceanographic feature exhibits a wind-driven, seasonal primary production that peaks during the austral winter when cold, nutrient-rich waters are injected into the photic zone. The increased primary productivity leads to a high export rate of organic carbon to the marine seafloor, where remineralization occurs, mainly driven by microbial metabolic processes such as sulfate reduction, denitrification, methanogenesis, methanotrophy and fermentation. Thus, carbon turnover in Namibian marine sediments plays a key role in large-scale nutrient losses and gains, oceanic N:P ratio dynamics and production of greenhouse gases such as CH, N<sub>2</sub>O and H<sub>2</sub>S. Here, a novel experimental approach known as quantitative Stable Isotope Probing (qSIP) was used to quantify 13C incorporation into bacterial and archaeal 16S rDNA from subseafloor sediments of the Namibian Continental Shelf. Incubations from 28 cm depth carbonate-rich, sulfidic sediments were set up for ten days under anoxic conditions with [13C] labeled bicarbonate and [13C] labeled diatomaceous extracellular polymeric substances (dEPS). Experimental data shows DIC assimilation by a total of 1676 microbial operational taxonomic units (OTUs) primarily affiliated to Gamma and Deltaproteobacteria, Chloroflexi, Planctomycetes, Latescibacteria and Acidobacteria. Archaeal OTUs belonging to Bathyarchaeota, Euryarchaeota and Asgardaeota accounted for 52% of the [13C] bicarbonate labeled populations. Lower levels for [13C] dESP incorporation were measured, with a total of 329 enriched microbial OTUs mainly from the same taxonomic groups found enriched in the autotrophic incubation, likely indicating a bias towards consumption and cycling of organic matter produced in dark carbon fixation. This study gives insights into the ecological features of several uncultured groups and their metabolic activity. Furthermore, it sheds light on the taxonomic identity of key microbial populations performing biochemical processes linked to carbon turnover in Namibian Continental Shelf anoxic sediments.



# **SUPERSELECTIVITY** IN SYNTHETIC PROTOCELLS



Cesar Rodriguez-Emmenegger DWI - Leibniz Institute for Interactive Materials.

I address the question: How can artificial superselectivity be accomplished in synthetic cell membrane mimics? Not only is this important to expand the understanding of biological systems, but also to develop synthetic protocells with life-like function. I will introduce our concept for superselectivity in synthetic cell membranes which requires the integration of (i) specificity, (ii) multivalency (to enhance binding but retain reversibility), (iii) 2D dimensional organization of receptors and (iv) concepts of cooperativity in binding. To tackle this my team has designed and synthesized new families of amphiphiles -comb-polymers and Janus dendrimers- that self-assemble into cell-mimetic vesicles. Although, these molecules do not exist in nature, the vesicles formed closely mimic the thickness, flexibility, and lateral 2D organization of cell membranes. These properties are precisely encoded in the chemical structure, architecture and topology of the macromolecular building blocks of the membrane. As an example, I will show our recent work where we discovered that the reactivity of sugar receptors towards lectins is enhanced by the 2D organization of sugars into nanoarrays (clustering) and raft-mimics (cooperativity) on the periphery of protocells.<sup>1</sup> Furthermore, this talk will show how to introduce life-like functions such as endocytosis of living bacteria without active cell machinery.<sup>2</sup>

1 C. Rodriguez-Emmenegger, et al, Proc Natl Acad Sci U S A 2019, 116, 5376-5382. 2 N. Kostina, K. Rahimi, Q. Xiao, T. Haraszti, S. Dedisch, J. P. Spatz, U. Schwaneberg, M. L. Klein, V. Percec, M. Moeller, C. Rodriguez-Emmenegger, Nano Lett 2019, 19, 5732-5738.

# INDEX - FLOOR 9

Aligns with meeting schedule

122	SAROJ KUMAR ROUT On the amyloid world hypothesis
123	ALEXANDER RUF The challenging detection of nucleobases from
124	ANNALENA SALDITT A thermal habitat that triggers the retention ar
125	<b>FABIAN SCHMIDT</b> The Origin of Interstellar CO <sub>2</sub>
126	CHRISTIAN SCHMITT RNA library screening for self-aminoacylating
127	TOBIAS SCHNITZER Catalytic peptides – Potential precursors of er
128	PETER RICHARD SCHREINER Carbohydrate formation in the absence of bios
129	<b>PHILIPP SCHWINTEK</b> Thermal gradient driven formation of homoc polynucleotide mixtures
130	ADRIANA SERRAO Understanding the genetic code from affinities
131	DOMINIK SÖDER Sugars program the hierarchical self-assembl
132	EMILIE SONG Probing RNA stability, formation, and catalysis and in Space





m preaccretional astrophysical ice analogs

nd RNA-catalyzed replication of RNA

tRNA precursors

nzymes?

synthesis

chiral domains in hydrogels starting from racemic

of aminoacyl adenylates to pre-transfer RNA motifs

ly in onion glycodendrimers

in simulated prebiotic environments on the early Earth



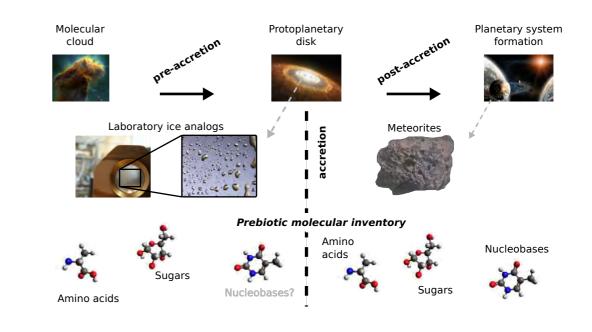
# ON THE AMYLOID WORLD ΗΥΡΟΤΗΕδΙδ



Saroj K. Rout, Radoslaw Bomba, Michael P. Friedmann, Witold Kwiatkowski, Jason Greenwald & Roland Riek

Laboratory of Physical Chemistry, Department of Chemistry and Applied Biosciences, ETH Zürich

How can simple molecules become organized into complex systems capable of supporting life? To address this question we have turned to small peptides that are capable of aggregating into beta-structured amyloids. We hypothesize that amyloids could have helped bridge the complexity gap that lies between small <200 Da prebiotic molecules and ordered macroscopic structures of the kind that can support catalytic and self-replicative functions. The projects described in this poster have revealed numerous activities and characteristics of amyloids, ranging from their feasibility as prebiotic entities and their effect on the chiral amplification of polymerizing amino acids to their ability to catalyze reactions and template their own replication. The cooperative interactions between amyloids and other (pre)biological molecules such as vesicle-forming fatty acids have also been investigated, providing insights into the formation of early membranes. Work by others on the cooperative assembly of amyloid and nucleic acid suggest that amyloids are capable of acting as a template for other polymers, suggesting a possible early connection between the peptide and RNA pre-biotic worlds. Amyloids, with their uniquely repetitive and templating structure, exceptional stability, chiro-selectivity, catalytic ability could have played important roles in the dynamic processes on the prebiotic earth that led to the increased complexity, organization and compartmentalization of key molecules and, eventually life.



### THE CHALLENGING DETECTION OF NUCLEOBASES FROM PREACCRETIONAL ASTROPHYSICAL ICE ANALOGS



Alexander Ruf<sup>1</sup>, Justin Lange<sup>2</sup>, Balkis Eddhif<sup>2</sup>, Claude Geffroy<sup>2</sup>, Louis Le Sergeant d'Hendecourt<sup>1</sup>, Pauline Poinot<sup>2</sup>, and Grégoire Danger<sup>1</sup> <sup>1</sup> Université Aix-Marseille, UMR CNRS 7345, Laboratoire de Physique des

Interactions loniques et Moléculaires (PIIM), Marseille, France <sup>2</sup> University of Poitiers, UMR CNRS 7285, Institut de Chimie des Milieux et Matériaux de Poitiers (IC2MP), E.BiCoM Team, 4 rue Michel-Brunet, TSA 51106, F- 86073 Poitiers cedex 9, France

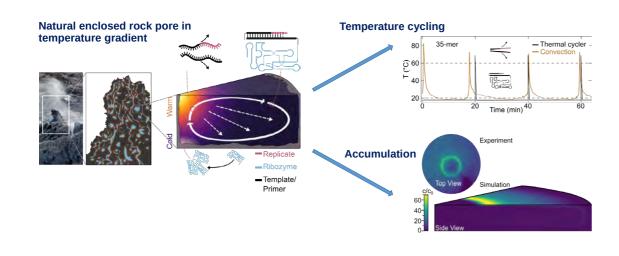
Amino acids, sugars, and nucleobases are considered as the so-called molecular bricks of life, the major subunits of proteins and genetic materials<sup>1</sup>. All three chemical families have been previously detected in meteorites<sup>2</sup>. In dense molecular cloud ice analogs, the formation of a large set of amino acids and sugars (+derivatives) has been observed<sup>3,4</sup>. In this contribution, we demonstrate that similar ices (H<sub>2</sub>O:<sup>13</sup>CH<sub>2</sub>OH:NH<sub>2</sub> ices, 2:1:1) can also lead to the formation of nucleobases<sup>5</sup>. Using combined UPLC-Orbitrap mass spectrometric and UPLC-SRM-triple quadrupole mass spectrometric analyses, we have unambiguously detected cytosine in these primitive, realistic astrophysical ice analogs. Additionally, a huge variety of nucleobase isomers was observed. These results indicate that all central subunits of biochemical materials may have already been present at early stages of chemical evolution of the protosolar nebula, before accretion toward planetesimals. Consequently, the formation of amino acids, sugars, and nucleobases does not necessarily require secondary alteration processes inside meteoritic parent bodies. They might have been supplied from dense molecular cloud ices toward post-accretional objects, such as nonaqueously modified comets, and subsequently delivered onto the early Earth's surface, potentially triggering the emergence of prebiotic chemistry leading to the first living systems.

1 Ruiz-Mirazo, K., Briones, C., & de la Escosura, A. (2014). Chemical Reviews, 114(1), 285-366. 2 Ruf, A., d'Hendecourt, L. L., & Schmitt-Kopplin, P. (2018). Life, 8(2), 18.

3 Caro, G. M., Meierhenrich, U. J., Schutte, W. A., Barbier, B., Segovia, A. A., Rosenbauer, H., ... & Greenberg, J. M. (2002). Nature, 416(6879), 403-406.

4 Meinert, C., Myrgorodska, I., De Marcellus, P., Buhse, T., Nahon, L., Hoffmann, S.V., ... & Meierhenrich, U. J. (2016). Science, 352(6282), 208-212.

5 Ruf, A., Lange, J., Eddhif, B., Geffroy, C., d'Hendecourt, L. L. S., Poinot, P., & Danger, G. (2019). The Astrophysical Journal Letters, 887(2), L31.



# A THERMAL HABITAT THAT TRIGGERS THE **RETENTION AND RNA-**CATALYZED REPLICATION OF RNA



Annalena Salditt<sup>1</sup>, David P. Horning<sup>2</sup>, Lorenz M. R. Keil<sup>1</sup>, Christof B. Mast<sup>1</sup>, Gerald F. Joyce<sup>2</sup> & Dieter Braun<sup>1</sup> Systems Biophysics, Physics Department, Center for Nanoscience, Ludwig-Maximilians-Universität München, 80799 Munich, Germany <sup>2</sup> The Salk Institute, 10010 N. Torrey Pines Road, La Jolla, CA 92037

Early replication in the RNA world is assumed to be an RNA-catalyzed process. In vitro evolution provided ever better RNA-based polymerases, but the required strand separation as well as the emergence of these complex ~200-base catalytic sequences are unsolved questions for the Origin of Life.

Here, we present a microfluidic reaction compartment with a pointed heat source, that both protected and drove laminar thermal convection in aqueous solution and allowed the autonomous, exponential RNA amplification by the RNA strand separation. The reaction proceeded despite the instability of RNA at elevated temperatures under the required salt conditions and offered replication kinetics comparable to explicit thermal cycling.

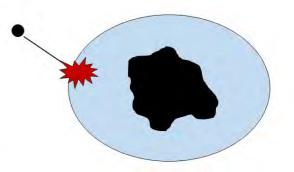
Accumulation experiments with fluorescently labeled nucleic acids revealed a ring like accumulation pattern for the long functional RNA-polymerase as well as its DNA complement, whereas similar experiments for dsDNA of similar length showed the expected central accumulation. Imaging the reaction mixture at higher resolution revealed the formation of micrometer sized conglomerates that depended on presence of PEG, introduced as crowding agent in the buffer. By including a diffusiophoretic term, the experimental accumulation behavior could be matched by simultaneously simulating the accumulation of the conglomerates and PEG with a commercial finite element simulation code

### THE ORIGIN OF INTERSTELLAR CO<sub>2</sub>



Fabian Schmidt, Jan Hendrik Bredehöft Universität Bremen, Institut für Angewandte und Physikalische Chemie

(2013)



Carbon monoxide (CO) and carbon dioxide (CO<sub>2</sub>) are some of the most abundant molecules in the icy mantles around interstellar dust grains<sup>1</sup> Unlike many other abundant molecules, CO<sub>2</sub> cannot be formed by gas phase reactions, as the newly formed bond immediately dissociates without a third body to remove excess energy<sup>2</sup>. While it is commonly accepted, that CO<sub>2</sub> forms from energetic processing of CO and water (H<sub>2</sub>O) in the ice phase, the exact mechanism of the reaction is less well known. One major point of contention is the role of the intermediate hydrocarboxyl radical (HOCO') whose presence in the reaction mixture has been clearly shown by infrared spectroscopy<sup>3</sup>. Theoretical models predict, however, that HOCO' should not be able to form CO, by dissociation of its O-H bond<sup>4</sup>, for energetic reasons.

It is known, that energetic processing of a H<sub>2</sub>0:CO mixture does not only produce CO<sub>2</sub>, but also formic acid, formaldehyde and methanol<sup>3</sup>. In the present poster, we present results of a further experimental investigation, looking at the H<sub>2</sub>0:CO system under energetic processing with low-energy electrons (2-20 eV)<sup>5</sup>. By looking at the dependence of product yield on electron energy, it is possible to unravel what kind of primary electron-molecule interaction starts a reaction sequence and which products are formed from specific intermediates. We observe that HOCO is indeed an important intermediate, just not on the reaction path to CO<sub>2</sub>, but as a precursor to formic acid. CO<sub>2</sub>, on the other hand is formed by dissociation of H<sub>2</sub>O into neutral O atoms and H<sub>2</sub>.

5 Schmidt, F., Swiderek, P. and Bredehöft, J.H. ACS Earth Space Chem 3:1974-1986 (2019).

<sup>1</sup> Hama, T. and Watanabe, N. Chem. Rev. 113:8783-8839 (2013).

<sup>2</sup> Shortridge, R.G., Lin, M.C. J. Chem. Phys. 64:4076-4085 (1976).

<sup>3</sup> Bennett, C.J., Hama, T., Kim, Y.S., Kawasaki, M., Kaiser, B.I. Astrophys. J. 727:27 (2011). 4 Arasa, C., van Hemert, M.C., van Dishoeck, E.F., Kroes, G.J. J. Phys. Chem. A 117:7064-7074

CATALYTIC PEPTIDES -

POTENTIAL PRECURSORS

<sup>1</sup> ETH Zürich, Laboratory of Organic Chemistry, Vladimir-

Complex Molecular Systems, Het Kranenveld, 5600 MB

<sup>2</sup> Eindhoven University of Technology, Institute for

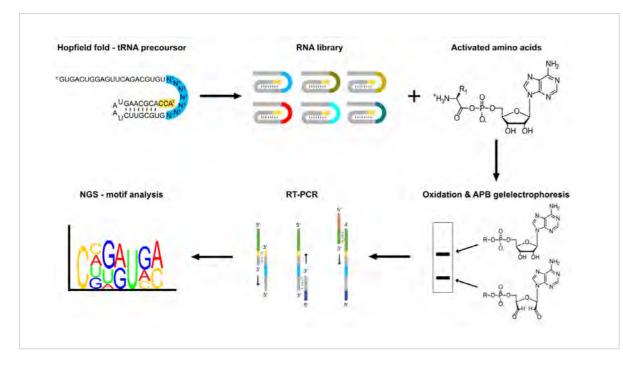
OF ENZYMES?

Eindhoven. The Netherlands.

T. Schnitzer<sup>1,2</sup>, J. W. Rackl<sup>1</sup>, H. Wennemers<sup>1</sup>

Prelog-Weg 5, 8093 Zürich, Switzerland

140 10829 5877



# RNA LIBRARY SCREENING FOR SELF-AMINOACYLATING TRNA PRECURSORS



Christian Schmitt, Andres Jäschke Institute of Pharmacy and Molecular Biotechnology, Heidelberg University

In every modern organism, protein biosynthesis follows the same principle: codons, which consist of three bases of genetic information encode one specific amino acid. This genetic code is basically identical throughout all kingdoms of life. Transfer RNAs (tRNAs) carry complementary anticodons which facilitate the translation of these codons into chains of amino acids i.e. proteins. But proteins are also needed to load the tRNAs with their corresponding amino acid. This resembles a chicken or egg problem for the beginning of life and even though this problem is well known to the scientific community, there is little experimental data available. This project specifically addresses the 'testable Hypothesis' formulated by J. J. Hopfield in 1978<sup>1</sup>. His postulated Hopfield folds resemble tRNA precursors in which the anticodon can directly interact with the amino acid attachment site. Unlike modern tRNAs should these precursors be able to bind specific amino acids on their own depending on the sequence of their anticodon. In this project, the hypothesis will be tested through kinetic sequencing of RNA libraries. The libraries consist of Hopfield folds with randomized anticodons and will be mixed with activated amino acids. Upon oxidation with periodate, APB gel electrophoresis, reverse transcription and reduced cycle PCR, DNA libraries are being created which are sequenced by next generation sequencing techniques. The sequencing data may show reproducible enrichments of certain sequences depending on which amino acid was used. This will not only show if such a precursor was practically possible but also how the genetic code itself was initially defined.

1 Hopfield, J. J. (1978). Origin of the genetic code: a testable hypothesis based on tRNA structure, sequence, and kinetic proofreading. Proceedings of the National Academy of Sciences, 75(9), 4334-4338.

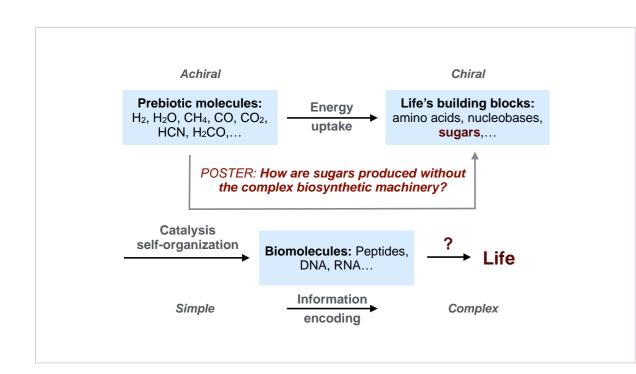
Nature uses enzymes to catalyze reactions essential for life. The complexity of enzymes is thereby crucial for their reactivity, stereo- and chemoselectivity. In recent years numerous small peptidic catalysts were developed that have high levels of reactivity and stereoselectivity.<sup>1</sup> These peptides are smaller and less complex than enzymes but show features reminiscent of enzymes. Moreover, peptides and enzymes are buildup of the same building blocks: amino acids. This provokes the question of a potential role of catalytic peptides in the evolution of enzymes. Based on a combinatorial library screening with more than 3000 peptides, our group developed tripeptidic catalysts of the H-Pro-Pro-Xaa type (Xaa: any amine). These peptides are highly reactive and stereoselective catalysts for aldol and conjugate addition reactions in organic solvents and water.<sup>2</sup> Underlying reaction mechanisms as well as conformational features of the peptides are well understood.<sup>3</sup> Hence, H-Pro-Pro-Xaa type catalysts are ideal model systems to explore a potential role of catalytic peptides in the evolution of life. Here, we address two related questions: 1) There is a general notion that the bigger and more complex peptidic catalysts are, the more enzyme-like and better they should become. But is this really true?<sup>4</sup> 2) A prerequisite for a potential role of peptides in the evolution of enzymes is high chemoselectivity. Yet, are peptidic catalysts robust enough to perform reactions in complex multi-component environments?5

1 E. A. Colby Davie, S. M. Mennen, Y. Xu, S. J. Miller, Chem. Rev. 2007, 107, 5759. 2 a) P. Krattiger, R. Kovasy, J. D. Revell, S. Ivan, H. Wennemers, Org. Lett. 2005, 7, 1101. b) M. Wiesner, J. D. Revell, H. Wennemers, Angew. Chem. Int. Ed. 2008, 47, 1871, c) C. E. Grünenfelder. J. K. Kisunzu, H. Wennemers, Angew. Chem. Int. Ed. 2016, 55, 8571. d) T. Schnitzer, A. Budinská, H. Wennemers, Nat. Catal. 2020. 3. 143.

3 a) T. Schnitzer, H. Wennemers, J. Am. Chem. Soc. 2017, 139, 15356. b) C. Rigling, J. K. Kisunzu, J. Duschmalé, D. Häussinger, M. Wiesner, M.-O. Ebert, H. Wennemers, J. Am. Chem. Soc. 2018,

4 T. Schnitzer, M. Wiesner, P. Krattiger, J. D. Revell, H. Wennemers, Org. Biomol. Chem. 2017, 15,

5 T. Schnitzer, J. W. Rackl, H. Wennemers, manuscript submittee



# CARBOHYDRATE FORMATION IN THE ABSENCE OF ΒΙΟδΥΝΤΗΕδΙδ

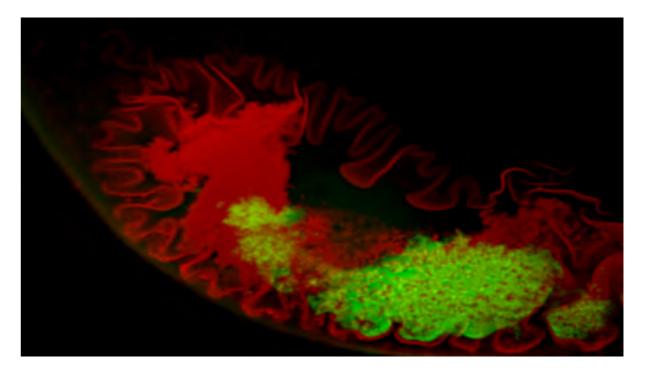
### Peter R. Schreiner

Institute of Organic Chemistry, Justus Liebig University, Heinrich-Buff-Ring 17, 35392 Giessen, Germany, prs@unigiessen.de, ORCID: 0000-0002-3608-5515

Although the simplest sugar, glycolaldehyde (HOCH, CHO), has been generated in the lab from its constituents1 and is suggested to occur in the formose<sup>2</sup> (Butlerow<sup>3</sup>) reaction, the mechanism for the dimerization of two H2CO molecules to glycolaldehyde and on to higher sugars is a riddle to date; the finding of "glycolaldehyde autocatalysis" does not explain the fundamental chemistry, requires the presence of liquid water, a strong base, high reactant concentrations, and ambient temperatures - all conditions unlikely to be present on early Earth or in extraterrestrial environments.<sup>4</sup> We focus on non-aqueous reactions, ideally starting directly from the photoreaction of CO and H, to give hydroxymethylene (HCOH)5: Under appropriate conditions H<sub>2</sub>CO and HCOH react to glycolaldehyde and glyceraldehyde.<sup>6</sup> Similarly, we demonstrate that glycolaldehyde and H<sub>2</sub>CO form a new 1,3-dioxolane that may well be the photostable storage form of these two key molecules.7 Finally, with "time-compression experiments" we demonstrate the formation of glyoxylic acid from the HCO and HOCO radicals under conditions mimicking those of interstellar water ices doped with CO.8

1 D. T. Halfen, A. J. Apponi, N. Woolf, R. Polt, L. M. Ziurys, The Astrophysical Journal 2006, 639, 237. 2 R. Breslow, Tetrahedron Lett. 1959, 22. 3 A. Butlerow, Liebigs Ann. Chem. 1861, 120, 295. 4 R. Shapiro, Origins Life Evol, Biosphere 1988, 18, 71. 5 a) P. B. Schreiner, H. P. Beisenauer, F. C. Pickard IV, A. C. Simmonett, W. D. Allen, F. Mátyus, A. G. wCsászár, Nature 2008, 453, 906; b) P. R. Schreiner, H. P. Reisenauer, D. Ley, D. Gerbig, C.-H Wu, W. D. Allen, Science 2011, 332, 1300. 6 A. K. Eckhardt, M. M. Linden, R. C. Wende, B. Bernhardt, P. R. Schreiner, Nat. Chem. 2018, 10, 1141.

[7] A. K. Eckhardt, R. C. Wende, P. R. Schreiner, J. Am. Chem. Soc. 2018, 140, 12333. [8] A. K. Eckhardt, A. Bergantini, S. K. Singh, P. R. Schreiner, R. I. Kaiser, Angew. Chem. Int. Ed. 2019, 58, 5663.



THERMAL GRADIENT DRIVEN FORMATION OF HOMOCHIRAL DOMAINS IN HYDROGELS STARTING FROM RACEMIC POLYNUCLEOTIDE MIXTURES



Philipp Schwintek, M. Heinlein, C. Mast and Dieter Braun

Systems Biophysics, Physics Department, Center for Nanoscience, Ludwig-Maximilians-Universität München, 80799 Munich, Germany

philipp.schwintek@physik.uni-muenchen.de

molecules in situ.

The origin of homochirality of DNA still remains an unresolved puzzle for the origin of life research<sup>1</sup>. In previous work, the formation of DNAhydrogels inside porous rock of hydrothermal vents has proven to be sequence specific<sup>2</sup>. This work investigates the selectivity of such hydrogels towards DNA-backbone enantiomers, forming homochiral domains starting from racemic solutions. Mimicking boundary conditions similar to hydrothermal pores as well as using real time fluorescent microscopy enables us to monitor the accumulation of DNA

By screening parameters such as salt concentrations, temperatures, DNA-sequences and pore-geometries, we investigate the phaseseparation into hydrogels as a darwinistic selection pressure that could have led towards homochirality of biopolymers.

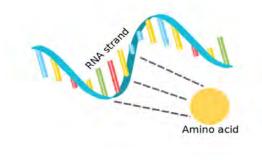
1 D. Blackmond Cold Spring Harbor perspectives in biology.2010, 2. Jg., Nr. 5, S. a002147. 2 M. Morasch, D. Braun, and C. Mast. Angewandte Chemie 128.23 2016: 6788-6791.

# SUGARS PROGRAM THE HIERARCHICAL SELF-ASSEMBLY IN ONION **GLYCODENDRIMERS**

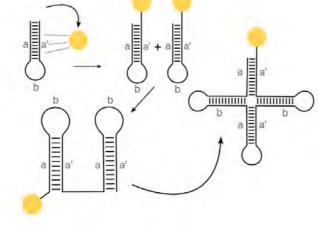
### D. Söder<sup>1</sup>, N. Yu. Kostina<sup>1</sup>, T. Haraszti<sup>1</sup>, K. Rahimi<sup>1</sup>, Q. Xiao<sup>2,3</sup>, M. L. Klein<sup>3</sup>, V. Percec<sup>2</sup>, C. Rodriguez-Emmenegger

<sup>1</sup> DWI - Leibniz Institute for Interactive Materials, Forckenbeckstraße 50, 52074 Aachen, Germany. <sup>2</sup> Roy & Diana Vagelos Laboratories, Department of Chemistry, University of Pennsylvania, Philadelphia, Pennsylvania 19104-6323, United States <sup>3</sup>Institute of Computational Molecular Science, Temple University, Philadelphia, Pennsylvania 19122, United States

Stereochemical theory



# Formation of pre-tRNA



UNDERSTANDING THE **GENETIC CODE FROM** AFFINITIES OF AMINOACYL ADENYLATES TO PRE-TRANSFER RNA MOTIFS



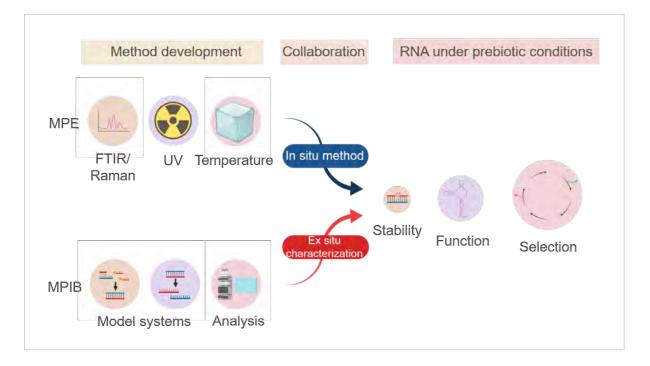
Adriana Serrao, Dieter Braun Systems Biophysics, Functional Nanosystems, Ludwig-Maximilian University Munich, Amalienstr. 54, 80799 Munich, Germany

The patterns present in the genetic code and its wide conservation across species suggest that it was not a random accident. According to the stereochemical theory, assignments between codons and amino acids originated from intrinsic interactions between RNA and amino acids or small peptides. RNA/Amino acid complexes could have played a role in translation from the beginning, or could have initially acted as cofactors for ribozymes or as nucleic acid stabilizers.

In 1978, J.J Hopfield proposed a potential primordial secondary structure for the tRNA in which the anticodon loop is placed in direct vicinity of the CCA end, which is a strong argument in favor of the stereochemical theory. As a starting point for the experimental validation of this theory, we are testing double stranded RNA complexes, based on the pre-tRNA structure proposed by Hopfield, and measuring their binding affinities to AMP and activated amino acid analogs. The binding affinity constants of the designed constructs to AMP are on the milimolar range. The next step consists of measuring the binding affinities of the corresponding activated amino acid analogs. We are interested in finding specific binding of the amino acid moieties to the constructs 'anticodon' overhang, which would reflect on a stronger binding. This could shed a light on the origin of the present day correspondence between codons and amino acids.

Natural cell membranes contain nano- and micro-organized domains formed by non-random association of lipids. Located at the cellular interface these functional domains contain glycoproteins and glycolipids that control functions such as communication, division, cargo trafficking, and signal transduction. The sugar moieties provide the structural basis for affinity in protein recognition and the formation of molecular patterns on the surface. But can these patterns be controlled by the sugar type? And how do these nanoarrays affect the binding of lectins, the cell-binding proteins? In this poster I tackle these questions with the help of Janus glycodendrimers (JGDs). Recently, we reported the self-assembly of JGDs into cell membranemimics in water, named glycodendrimersomes (GDSs).<sup>1</sup> The resulting uni- and multilamellar vesicles closely resemble the thickness, flexibility and lateral 2D organization of natural cell membranes.<sup>2</sup> These properties stem from the chemical structure, architecture, and topology of the dendrimer. For this study, the JGDs are decorated with the monosaccharide mannose (Man). Man is highly biologically relevant. We demonstrated an increase in the biological activity of sugars when they formed nanoarrays. We discovered that the coassembly of judiciously selected Janus dendrimers (JDs) with JGDs led to multiscale membrane architectures. AFM allowed observing how dendrimers segregated in micrometer-size raft-like domains with the Man moieties nano-assembled in lamellar or hexagonal patterns. These periodic arrays of Man resulted in a new ligand with enhanced reactivity to Concanavalin A as determined by surface plasmon resonance. Thus, these experiments provide a powerful example in which structure determines function, in particular how different supramolecular assemblies encode biological recognition.

1 C. Rodriguez-Emmenegger, et al., Proc. Natl. Acad. Sci. U.S.A. 2019, 116, 5376-5382. 2 N. Kostina, et al. Nano Lett. 2019, 19, 8, 5732-5738



PROBING RNA STABILITY. FORMATION, & CATALYSIS IN SIMULATED PREBIOTIC ENVIRONMENTS ON THE EARLY EARTH AND IN **SPACE** 



Germany

Emilie Y. Song<sup>1</sup>, Max Winkler<sup>2</sup>, Paola Caselli<sup>2</sup>, Hannes Mutschler<sup>1</sup> Max Planck Institute of Biochemistry, Am Klopferspitz 18, Martinsried 82152,

<sup>2</sup> Max Planck Institute for Extraterrestrial Physics, Giessenbachstrasse 1, Garching 85748, Germany

Can life based on nucleic acids survive and evolve in extreme, precellular conditions provided by early Earth or even space? Given the lack of spatiotemporal data of abiogenesis, a wide range of conditions provide possible prebiotic environments including hydrothermal vents<sup>1</sup>, impact craters<sup>2</sup>, ice<sup>3</sup>, and warm ponds<sup>4</sup>. We aim to reduce the number of these possible environments by systematically narrowing down the range of physicochemical parameters that allow the sustained emergence and/or existence of biopolymers. Specifically, we focus on ribonucleic acid (RNA) polymers and building blocks, which are thought to be central key players in early chemical and Darwinian evolution5. We employ a powerful reaction setup capable of simulating relevant prebiotic environmental conditions such as the young Sun UV spectrum. Using this setup, we characterize RNA survival, synthesis, and catalysis under realistic conditions using methods such as Raman spectroscopy, HPLC, electrophoresis, and deep sequencing to explore to what extent RNA molecules are capable of survival, adaptation and evolution in extreme environments.

1 Sojo, V. et al. (2016) The Origin of Life in Alkaline Hydrothermal Vents. Astrobiology 16. 2 Cockell, C. S. (2006). The origin and emergence of life under impact bombardment. Philosophical Transactions of the Royal Society of London B: Biological Sciences. 3 Price, P. B. (2006). Microbial life in glacial ice and implications for a cold origin of life. FEMS Microbiology Ecology.

4 Pearce, B. K. et al. (2017). Origin of the RNA world: The fate of nucleobases in warm little ponds. PNAS.

5 Leslie E, O. (2004). Prebiotic chemistry and the origin of the RNA world. Critical reviews in biochemistry and molecular biology

# INDEX - FLOOR 10

Aligns with meeting schedule

134	CHRISTINA SPRINGSKLEE Prebiotic synthesis in volcanic discharges:
135	<b>TOMISLAV STOLAR</b> The solid-state as a reaction medium for pre- pairing of model nucleobases
136	JAN TENBUSCH Super-flexible biomimetic vesicles
137	BEATRIZ VON DER ESCH Prebiotic pathway to DNA nucleosides
138	MARIIA VOROBII Building membrane machines to endocytize l
139	ANNA MARIA WAGNER How does spontaneous curvature induce the
140	CRAIG WALTON Phosphorous mineralogy on the Early Earth
141	KARL WIENAND Public outreach on 'Emergence of Life'
142	MAX WINKLER UV resistance of nucleosides - an experimen
143	SREEKAR WUNNAVA Acid-catalyzed polymerization of cyclic GMP
144	<b>NOEL YEH MARTIN</b> Out-of-equilibrium cellular mimics driven by



An experimental approach

biotic chemical selection: dry heating enables selective

living bacteria: the battle between adhesion and flexibility

morphogenesis of dendrimersome vesicles?

tal approach

thermal gradients



# PREBIOTIC SYNTHESIS IN **VOLCANIC DISCHARGES**: AN EXPERIMENTAL **APPROACH**



Christina Springsklee<sup>1</sup>, Thomas Steiner<sup>2</sup>. Thomas Geisberger<sup>2</sup>. Bettina Scheu<sup>1</sup>, Claudia Huber<sup>2</sup>, Wolfgang Eisenreich<sup>2</sup>, Corrado Cimarelli<sup>1</sup>, and Donald Bruce Dingwell<sup>1</sup>

<sup>1</sup> Ludwig Maximilian University of Munich, Munich, Germany <sup>2</sup> Technical University of Munich, Munich, Germany

The formation of primitive organic molecules is a key question in the enigmatic debate on the emergence of life on Early Earth. Several iconic experiments already explored potential processes which lead to the formation of organic molecules. One of these experiments are the discharge experiments performed by Miller and Urey, which corroborates the conclusion that amino acids can be produced by lightning under reducing atmospheric conditions<sup>1</sup>.

This project aims to combine prebiotic discharge experiments with a geological relevant setting for Early Earth: active volcanism. For this purpose, a shock tube apparatus was developed to perform discharge experiments in varying atmospheres. The discharges are generated by the eruption of the volcanic ash itself by triboelectrification and fracto-emission, a process which is frequently observed in nature and described as volcanic lightning. In the experiment the total magnitude of electric discharge can be adjusted by changing the amount of mass of ash, the proportion of fines and eruptive conditions<sup>2</sup>. The development of the new experimental setup allows to probe the ejected sample, the gas atmosphere, present before and after the experiment, as well as to quantify the magnitude of total discharge generated during the experiment. Special focus is given to the role of ash, offering variable porosity, high surface area and significant surface reactivity. This project aims to determine and quantify the impact of volcanic ash on its environmental setting as a catalyst and a container in the creation and accumulation of first organic molecules.

1 Miller, S.L. (1953). A production of amino acids under possible primitive earth conditions. Science.117.528-529. 2 Gaudin, D. and Cimarelli, C. (2019). The electrification of volcanic jets and controlling parameters: A laboratory study. EPSL,513,69-80.

THE SOLID-STATE AS A **REACTION MEDIUM FOR** PREBIOTIC CHEMICAL **SELECTION: DRY** 

Dry environment as a reaction medium for

chemical selection?

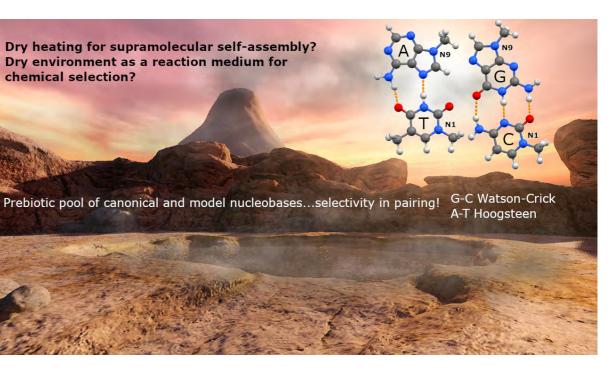
Ruđer Bošković Institute, Bijenička c. 54, 10000 Zagreb, Croatia <sup>2</sup> Deutsches Elektronen-Synchrotron (DESY), 22607 Hamburg, Germany <sup>3</sup> Xellia Pharmaceuticals, Slavonska avenija 24/6, 10000 Zagreb, Croatia

**SELECTIVE PAIRING OF** MODEL NUCLEOBASES Tomislav Stolar<sup>1</sup>, Martin Etter<sup>2</sup>,

Krunoslav Užarević<sup>1</sup>, Ivan Halasz<sup>1</sup>, Ernest Meštrović<sup>3</sup>

**HEATING ENABLES** 





From the perspective of prebiotic DNA assembly, it seems unlikely that the specific pairs of nucleobases would have been coupled into DNA if they were unwilling to selectively and specifically self-assemble beforehand<sup>1</sup>. Base pairing of canonical nucleobases, or their simple derivatives, is known to be notoriously difficult to achieve in aqueous media, due to solvation issues and competition for hydrogen bonding<sup>2</sup>. Here, we show that pairing of methylated guanine and methylated cytosine is readily achievable by dry heating of their solid mixtures<sup>3</sup>. In the G-C cocrystal, molecules self-assemble in the Watson-Crick hydrogen-bonded motif. Furthermore, selectivity in the solid-state selfassembly is DNA-specific. Dry heating of a four-component mixture of methylated adenine, guanine, cytosine, and thymine provided only A-T and G-C pairing. Equivalent experiments with canonical nucleobases failed to yield pairing, and there seems to be a strong influence of the methyl group at the glycosidic nitrogen atom. The findings of this study suggest that chemical processes occurring in the solid-state could have had an important role in the prebiotic chemistry context. For our future endeavors, we will include a larger pool of proto-nucleobases<sup>4,5</sup>.

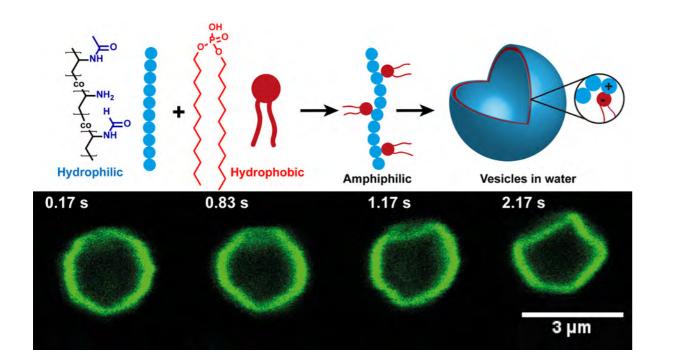
1 N. V. Hud. S. S. Jain, X. Li, D. G. Lynn, Chem. Biodivers, 2007, 4, 768–783.

2 P. Cieplak and P. A. Kollman, J. Am. Chem. Soc. 1988, 110, 3734-3739.

3 T. Stolar, S. Lukin, M. Rajić Linarić, M. Etter, K. Užarević, I. Halasz, E. Meštrović, ChemRxiv 2019, DOI: 10.26434/chemrxiv.8327162.v2

4 B. J. Cafferty and N. V. Hud. Isr. J. Chem. 2015, 55, 891–905.

5 A. C. Rios and Y. Tor, Isr. J. Chem. 2013, 53, 1-15.



### SUPER-FLEXIBLE **BIOMIMETIC VESICLES**

Materials e.V. Aachen



Jan Tenbusch, Pilar Bologna, Nina Kostina, Mehrnoush Rahimzadeh, Khosrow Rahimi, Cesar Rodriguez-Emmenegger DWI - Leibniz Institute for interactive

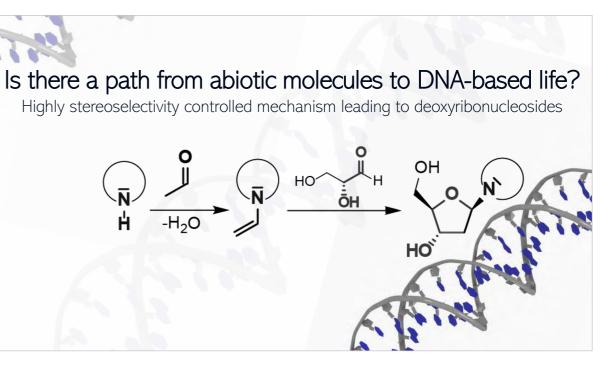
Compartmentalization is essential to life. As protocellular research and the study of abiogenesis - was driven forward in the last couple decades, researchers had to tackle different obstacles. Stability and longevity in cell membrane models are achieved by either utilizing cholesterol in high molar ratios when working with liposomes, or polymersomes assembled from block copolymers. Both approaches present their own disadvantages such as a low 2D mobility and storage problems for the former and no biomimetic thickness as well as a low flexibility and 2D mobility for the latter. In this poster we will present a new class of amphiphiles that are able to self-assemble into super-flexible biomimetic vesicles. We designed and synthesized non-covalently linked amphiphilic comb-shaped oligomers which amalgamate the stability of polymersomes with the biomimetic properties of liposomes while increasing the flexibility many times over. A flexible intrinsic hydrophilic oligomer backbone is utilized to which alkyl chains are attached by ionic interactions. The length of the alkyl chains is selected to represent the biomimetic thickness of the natural cell membrane (5±1 nm). We control the degree of flexibility by encoding the grafting density of the alkyl chains along the hydrophilic backbone. These molecules create new possibilities for applications where high dynamics of the membrane are necessary.

### PREBIOTIC PATHWAY TO DNA NUCLEOSIDES



Kai Kohler, Beatriz von der Esch, Florian Kruse, Christian Ochsenfeld, Oliver Trapp University of Munich (LMU), Butenandtstr. 7, 81377 Munich, Germany

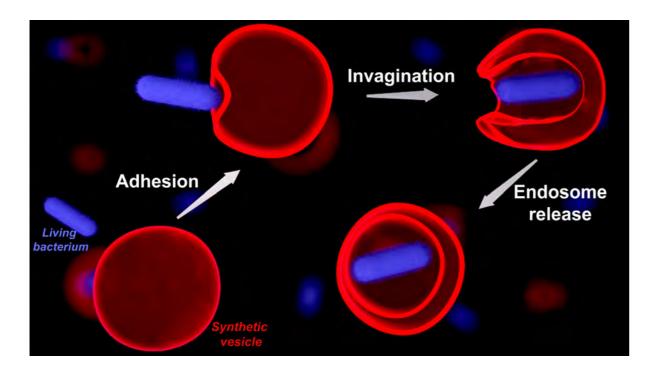
 $-H_2O$ 



So far it is assumed that RNA played a key role in the origin of life. However, the transition from ribonucleotides to deoxyribonucleotides remains unknown.<sup>1</sup> Therefore, the question remains: Is there a path from abiotic molecules to DNA-based life?

We are exploring the formation of deoxyribonucleosides under relevant prebiotic conditions in water in high regio- and stereoselectivity from adenine, guanine, cytosine, and thymine. By condensation of the canonical bases with acetaldehyde and sugar-forming precursors we found a continuous path to deoxyribonucleotides, starting from prebiotically available molecules.<sup>2,3</sup> The experimental studies are complimented with computational simulations to elucidate aspects which are experimentally inaccessible.

1 G. F. Joyce, Nature 2002, 418, 214. 2 J. S. Teichert, F. M. Kruse, and O. Trapp, Angew. Chem. Int. Ed. 2019, 58, 9944-9947. 3 A. M. Poole, D. T. Logan, B.-M. Sjöberg, J. Mol. Evol. 2002, 55, 180-196



# **BUILDING MEMBRANE** MACHINES TO ENDOCYTIZE LIVING **BACTERIA: THE BATTLE** BETWEEN ADHESION AND FLEXIBILITY



Marija Vorobij, Nina Kostina, Jan Tenbusch, Tamas Haraszti, Mehrnoush Rahimzadeh, Dominik Söder, Virgil Percec, Cesar Rodriguez-Emmenegger DWI - Leibniz Institute for Interactive Materials e.V., RWTH Aachen UniversityDepartment of Physics, LMU Munich

There is much interest in developing vesicular microcompartments from natural and synthetic amphiphiles, enabling programmable interactions with living matter. Of particular interest is the development of vesicles capable of endocytosis of living bacteria. Despite the complexity of this process, theoretical studies predict that the endocytosis of prolate micro-objects is possible without the need of active cell machinery if the energy released upon bacterial adhesion to the membrane surpasses the energy required to bend the membrane. Nonetheless, natural liposomes and synthetic polymersomes fail to sufficiently recapitulate membrane properties to perform this advanced function. Here we report the engulfment of living bacteria into endosomes by cell-like dendrimersomes assembled from Janus dendrimers.<sup>1</sup> Full engulfment occurred in less than a minute after contact. The process is driven by the adhesion of the bacterium to the dendrimersome's membrane by ultraweak interactions, comparable to those utilized by nature. The key to success relies on the combination of high flexibility and stability of the dendrimersomes. The key properties of the dendrimersomes are programmed into the molecular structures of their building blocks. The ability to support endocytosis highlights opportunities for the design and programming of dendrimersomes in biomedical research.

1 Kostina, N. Y.; Bahimi, K.; Xiao, O.; Haraszti, T.; Dedisch, S.; Spatz, J. P.; Schwaneberg, U.; Klein, M. L.; Percec, V.; Moller, M.; Rodriguez-Emmenegger, C., Membrane-Mimetic Dendrimersomes Engulf Living Bacteria via Endocytosis. Nano Lett 2019, 19 (8), 5732-5738.

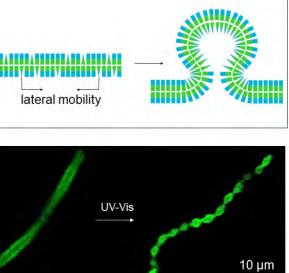
trigger Dendrimersomes o-assembled from Janus Dendrimers UV-Vis UV-Vis

### **ΗΟΨ DOES SPONTANEOUS** CURVATURE INDUCE THE MORPHOGENESIS OF DENDRIMERSOME **VESICLES?**

A. M. Wagner, N. Y. Kostina, T. Haraszti, K. Rahimi and C. Rodriguez-Emmenegger

DWI - Leibniz Institute for Interactive Materials, Forckenbeckstraße 50, 52074 Aachen, Germany. vesicles.

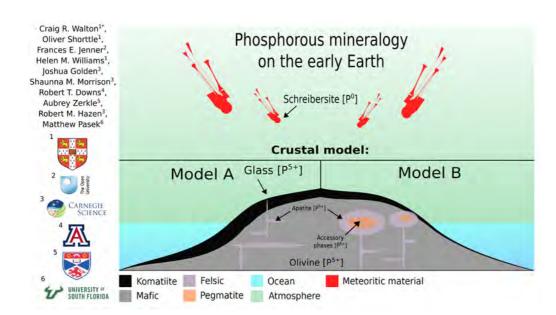
nanolett 9b02349



The vital functions of cell membranes require their ability to guickly change shape to perform complex tasks such as motion, division, endocytosis, and apoptosis. Membrane curvature in cells is modulated by very complex processes such as changes in lipid composition, the oligomerization of curvature-scaffolding proteins and the reversible insertion of protein regions that act like wedges in the membrane. But, could a much simpler mechanism support membrane shape transformation? In this poster, I will show how the change of the amphiphile topology (shape) in the bilayer can drive the morphogenesis of cell membrane models. To tackle this, we have designed and synthesized a new type of amphiphiles -Janus dendrimers- that selfassemble into uni- or multilamellar vesicles.1 Although these molecules do not exist in nature, the vesicles formed closely mimic the thickness, flexibility and the lateral 2D organization of cell membranes. These properties are precisely encoded in the chemical structure, architecture, and topology of the macromolecular building blocks of the membrane.<sup>2</sup> For these studies, we synthesized Janus dendrimers containing a photo-labile bond that upon UV-irradiation cleave losing a part of the hydrophilic dendron. This leads to a change from a cylindrical to a wedge-shaped amphiphile. The high mobility of these dendrimers allows for the concentration of the wedge-shaped amphiphiles and the generation of local spontaneous curvature. The concentration of the wedges and their rate of segregation allowed controlling the budding and generation of structures such as tubules, starfish, and high genus

1 C. Rodriguez-Emmenegger, et al, Proc Natl Acad Sci U S A 2019, 116, 5376-5382. 2 N. Kostina, K. Rahimi, O. Xiao, T. Haraszti, S. Dedisch, J. P. Spatz, U. Schwaneberg, M L. Klein, V. Percec, M. Moeller, C. Rodriguez-Emmenegger, Nano Lett 2019, 10.1021/acs.





# PHOSPHOROUS MINERALOGY ON THE EARLY EARTH



### Craig R. Walton<sup>1</sup>,Oliver

Shorttle<sup>1,2</sup>, Frances E. Jenner<sup>3</sup>, Helen M. Williams¹, Joshua Golden⁴, Shaunna M. Morrison<sup>5</sup>, Robert T. Downs<sup>4</sup>, Aubrey Zerkle<sup>6</sup>, Robert M. Hazen<sup>4</sup>, Matthew Pasek<sup>7</sup>

<sup>1</sup> Department of Earth Sciences, University of Cambridge, Downing Street, Cambridge CB2 3EQ, UK <sup>2</sup> Institute of Astronomy, University of Cambridge, Madingley Road, Cambridge, CB3 OHA, UK <sup>3</sup> School of Environment, Earth and Ecosystem Sciences. The Open University, Walton Hall, Milton Keynes MK7 6AA, UK <sup>4</sup> Geophysical Laboratory, Carnegie Institution for Science, 5251 Broad Branch Road, NW, Washington, DC 20015, UK <sup>5</sup> Department of Geosciences, University of Arizona, 1040 E. 4th Street, Tucson, AZ 85721, USA <sup>6</sup> School of Earth & Environmental Sciences, University of St Andrews, Irvine Building, KY16 9AL <sup>7</sup> School of Geoscience, University of South Florida, 4202 E Fowler Ave, Tampa FL 33620, USA

It has long been held that the early crustal phosphorous (P) reservoir was dominated overwhelmingly by apatite (Ca(PO<sub>4</sub>)<sub>2</sub>OH,Cl,F). The relative insolubility of apatite under ambient surface conditions today presents a challenge to the early availability of phosphate with which to drive prebiotic chemistry and sustain early life. However, stark differences in P mineralogy are found between mafic and felsic rocks. Given that global crustal compositions have evolved towards more felsic compositions over time, we might expect a contemporaneous evolution in crustal P mineralogy. We critically examine the evidence for a such a shift, beginning with a review of the processes that lead to the formation of the Earth, moving through the delivery of exogeneous P to the Earth's surface, and exploring the possible makeup of P mineral reservoirs in the early emergent crust. We conclude that exogeneous schreibersite, along with crustal silicate-hosted P (olivine, pyroxene, and glass), and terrestrial apatite were dominant on the early Earth. However, apatite was likely a much more minor phase in the early crust, in particular during the Prebiotic Era, than has been previously assumed (< 50 %, potentially as low as < 1 %). Silicate weathering may have played a vital role in supplying prebiotic chemistry and early life with available phosphate.

### PUBLIC OUTREACH ON EMERGENCE OF LIFE



Karl Wienand, Wolfgang M. Heckl1, CRC 235 Collaboration Deutsches Museum, Museumsinsel 1, 80538 Munich, Germany



Research on the emergence of life has the potential to captivate a large public, but is often little known. Involving scientists, especially junior scientists, could improve the communication and the public understanding of this important field of research<sup>1</sup>.

The Collaborative Research Center 235 Emergence of Life<sup>2</sup> puts a strong emphasis on outreach. Its planned effort includes a number of initiatives-from public talks to events in schools. It will culminate in the realization of a museum exhibition about the science and research on emergence of life. Participating researchers (especially PhD students) actively participate in ideating, designing, and realizing the exhibition and all other outreach activities.

In this poster, I will present some contents currently in the works, both for the exhibition and for more general outreach.

1 D. Battachary. Survey of Factors Affecting Science Communication: Conclusions, Recommendations and Actions. Royal Society, 2006. 2 https://www.emergence-of-life.de

# UV RESISTANCE OF NUCLEOSIDES - AN

Max Winkler, Barbara Michela Giuliano, Paola Caselli Max Plank Institute for Extraterrestrial Physics

EXPERIMENTAL APPROACH

The emergence of life on Earth is a highly discussed, but still unsolved question. Different hypotheses have been proposed. Current research underlines the importance of environments within close proximity to the Earth's surface as they can solve long standing problems like polymerisation of nucleotides1 and phosphorylation of nucleosides2. However, surface-near settings, e.g. ponds or ice shields, are prone to UV irradiation. We investigated the photosensitivity of uracil, uridine, adenosine, cytidine and guanosine by using Raman microscopy. DMSO was used as a solvent to improve the signal to noise ratio. The samples were irradiated by a UV source with 150 mW/cm<sup>2</sup> for 10 minutes. Uracil and uridine showed the highest photosensitivity, while adenosine, cytidine and guanosine remained stable. This stands in contradiction with previews works using ab initio quantum calculations<sup>3</sup>. These studies concluded that uracil is more photostable than the other canonical nucleobases. However, theoretical investigations suffer from limitations and simplifications, e.g. intermolecular interaction or their photo cross sections, due to high computational costs. This could explain the observed lower UV resistance of uracil and uridine. Understanding the survivability of these important biomolecules is important to understand the prebiotic chemistry of the early Earth. Environmental stresses could have served as drivers towards a rise in chemical complexity.

1 Mutschler, H., Wochner, A. & Holliger, P. Freeze-thaw cycles as drivers of complex ribozyme assembly. Nat. Chem. 7, 502-508 (2015). 2 Toner, J. D. & Catling, D. C. A carbonate-rich lake solution to the phosphate problem of the origin of life. Proc. Natl. Acad. Sci. 117, 883-888 (2020). 3 Beckstead, A. A., Zhang, Y., de Vries, M. S. & Kohler, B. Life in the light: nucleic acid photoproperties as a legacy of chemical evolution. Phys. Chem. Chem. Phys. 18, 24228-24238 (2016)

### **FLOOR 10**

## ACID-CATALYZED POLYMERIZATION OF CYCLIC GMP

### Sreekar Wunnava<sup>1</sup>, Judit Šponer<sup>2</sup> and Dieter Braun<sup>1</sup>

<sup>1</sup> Physics Department, NanoSystems Initiative Munich and Center for Nanoscience Ludwig-Maximilians-Universität München, Amalienstrasse 54, 80799 München, Germany.

<sup>2</sup> Institute of Biophysics, Academy of Sciences of the Czech Republic, Královopolská 135, Brno 61265, Czech Republic

Different prebiotic pathways have been explored for the synthesis of nucleosides and nucleotides, yet our current understanding lacks in deciphering a prebiotically plausible route for their polymerization to form long linear oligonucleotides. Methods involving activation chemistry of nucleotides in aqueous solutions are not only challenging due to different side reactions, but gives rise to the question whether such activating agents existed on prebiotic Earth. Cyclic nucleotides have been shown to be a viable precursor in prebiotic polymerization<sup>1</sup>. It has been demonstrated previously that 3'-5' cGMP, in its free acid form, can spontaneously polymerize under hot and dry conditions by transphosphorylation of the phospohodiester bond<sup>2</sup>. We demonstrate that the Na-salt of cGMP, which does not polymerize upon drying in alkaline conditions, can be polymerized under acidic conditions. Therefore, in addition to previously described base-catalyzed reaction mechanism<sup>3,4</sup>, we propose and motivate an alternative acid-catalyzed mechanism for the ring opening polymerization of cGMP.

(2015): 2979-2989.

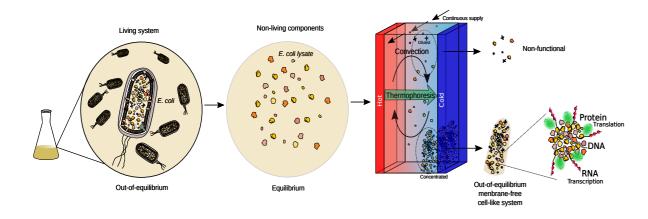
3 Sponer, Judit E., et al. "Untemplated nonenzymatic polymerization of 3', 5' cGMP. a plausible route to 3', 5'-linked oligonucleotides in primordia." The Journal of Physical Chemistry B 119.7

4 Costanzo, Giovanna, et al. "Generation of RNA Molecules by a Base-Catalysed Click-Like Reaction." ChemBioChem 13.7 (2012): 999-1008.



<sup>1</sup> Renz, M et al. "Catalysts for the polymerization of adenosine cyclic 2',3'-phosphate on a poly (U) template." Biochimica et biophysica acta vol. 240,4 (1971): 463-71.

<sup>2</sup> Morasch, Matthias, et al. "Dry Polymerization of 3', 5'-Cyclic GMP to Long Strands of RNA." ChemBioChem 15.6 (2014): 879-883.



## **OUT-OF-EQUILIBRIUM CELLULAR MIMICS** DRIVEN BY THERMAL GRADIENTS

Noël Yeh Martín<sup>1</sup>, Laura Weise<sup>2</sup>, Hannes Mutschler<sup>2</sup>, Sheref Mansy<sup>3,4</sup>, Christof Mast<sup>1</sup> & Dieter Braun<sup>1</sup>. <sup>1</sup> System Biophysics, Ludwig - Maximilian Universität, Amalienstr 54. 80799 Munich, Germany. <sup>2</sup> Max Planck Institute of Biochemistry, Am Klopferspitz 18, 82152 Martinsried, Germany. <sup>3</sup> Department of CIBIO, University of Trento, via Sommarive 9, 38123 Povo, Italy. <sup>4</sup> Department of Chemistry, University of Alberta, 11227 Saskatchewan Drive, Edmonton AB T6G 2G2, Canada.

Thermal gradients applied at the microscale have been shown to accumulate diluted simple mixtures of molecules to high local concentrations<sup>1-3</sup>. It is unclear, however, whether thermal gradients could also drive complex biochemical reactions. In other words, it is yet to be assessed whether vastly different biomolecules in complex mixtures can be accumulated together into a cooperative and functioning system that could display the properties of a living cell. Therefore, we are currently exploring the feasibility of exploiting thermal gradients to generate a disequilibrium system reminiscent of natural living cells by accumulating the hundreds of components of an E. coli cell lysate<sup>4</sup> and the purified components of PURE system<sup>5</sup> needed to reconstitute RNA and protein synthesis in the absence of a membrane comparment. Furthermore, we are aiming at sustaining the activity of our system over long periods of time by a continuous supply of feedstock molecules and removal of waste toxic products that would lead to a long lasting cell like system operating under disequilibrium. Ultimately, we hope that our approach will help us gain insight of the fundamental processes and the set of conditions needed to sustain cellular life.

1 Kreysing, M. et al. Heat flux across an open pore enables the continuous replication and selection of oligonucleotides towards increasing length. Nat. Chem. 7, 203-208 (2015). 2 Mast, C. B. et al. Escalation of polymerization in a thermal gradient. Proc. Natl. Acad. Sci. U. S. A. 110, 8030-8035 (2013).

3 Keil, L. et al. Proton gradients and pH oscillations emerge from heat flow at the microscale. Nat. Commun. 8, 1897 (2017).

4 Shimizu, Y. et al. Cell-free translation reconstituted with purified components. Nat. Biotechnol. 19.751-755 (2001)

5 Sun, Z. Z. et al. Protocols for Implementing an Escherichia coli based TX-TL Cell-Free

Expression System for Synthetic Biology. J. Vis. Exp. e50762-e50762 (2013).

#### INDEX - FLOOR 11 (UTC -04:00 TO -07:00)

146	ZACHARY COHEN Fatty acid membranes are stable in carbonat
147	SAURJA DASGUPTA Chemistry and catalysis join forces in RNA liga
148	HADI FARES Impact of wet-dry cycling on the phase and co
149	JAY FORSYTHE Proline incorporation in model prebiotic deps
150	MCCAULEY MEYER Nucleotide-level resolution of RNA folding inte
151	TRISHOOL NAMANI Role of amino acids on nonenzymatic clay-pro
152	ARTASH NATH Using machine learning to improve prediction of
153	ALINE NOVAIS Exoplanetary atmospheric retrieval using ma
154	FATMA PIR CAKMAK Prebiotically-relevant low polyion multivalency compartments
155	RAGHAV POUDYAL RNA world inside compartments: Activation of
156	MICHAEL L WONG & STUART BARTLETT Defining lyfe in the universe: From three privile
157	WEN ZHANG Deciphering nonenzymatic RNA polymerization



Visit at late hours of the event or during poster session

te-rich, prebiotic lake environments

ation

ompartmentalization behaviors of complex coacervates

sipeptides

eractions within peptide-based complex coacervates

omoted oligomerization of activated nucleotide

of chemical composition of exoplanetary atmospheres

achine learning

y might improve functionality of membraneless

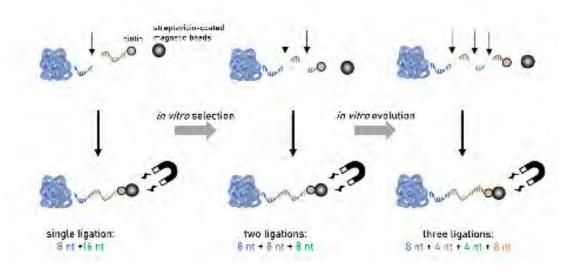
RNA catalysis by complex coacervates

eged functions to four pillars

on through crystallography

#### **FLOOR 11**

Evolving ribozymes to favor ligations of shorter substrates - Short to long RNAs



### CHEMISTRY AND CATALYSIS JOIN FORCES IN RNA LIGATION



S.DasGupta<sup>1,2</sup>, T. Walton<sup>1,2</sup>, D. Duzdevich<sup>1,2</sup>, S.S.Oh<sup>3</sup> and J. W. Szostak<sup>1,2</sup>

<sup>1</sup> Center for Computational and Integrative Biology and Department of Molecular Biology, Massachusetts General Hospital, <sup>2</sup> Department of Genetics. Harvard Medical School. <sup>3</sup>Pohang University of Science and Technology (POSTECH), South Korea.

2020, 117, 5741-5478,

## FATTY ACID MEMBRANES ARE STABLE IN CARBONATE-RICH. PREBIOTIC LAKE **ENVIRONMENTS**



Zachary R. Cohen<sup>1</sup>, Caitlin Cornell<sup>1</sup>, David Catling<sup>2</sup>, Roy Black<sup>1</sup>, Sarah Keller<sup>1</sup>

Department of Chemistry <sup>2</sup> Department of Earth and Space Science, University of Washington, Seattle WA 98195

Prebiotic membrane compartments were likely composed of fatty acids. Saturated fatty acids are widely considered to be available in the early Earth environment due to their presence on carbonaceous meteorites<sup>1</sup> and potential abiotic synthesis via Fischer-Tropsch reactions<sup>2</sup>. Here I will show that membrane formation is possible in shallow. carbonate-rich lake environments. These carbonate-rich lakes are attractive sites for prebiotic chemistry for multiple reasons: high carbonate concentrations can help solubilize phosphate minerals<sup>3</sup> and sequester ferrocyanide salts<sup>4</sup>, and periodic drying and wetting of shallow water can drive condensation reactions5. However, high concentrations of salts have been shown to disrupt formation of pure fatty acid membranes<sup>6</sup>. I have tested mixtures of salts (HCO<sub>2</sub><sup>-</sup>, PO<sub>4</sub><sup>-</sup> , Cl- anions) that are likely components of shallow, early Earth lake environments. These experiments show that 2:1 mixtures of decanoic acid and decanol (both putative prebiotic amphiphiles) can form stable compartments in the presence of .5M prebiotic salts. My results strengthen the claim that shallow lake environments are attractive sites for prebiotic chemistry.

٠

1 J. C.-Y. Lai, B. K. D. Pearce, R. E. Pudritz, and D. Lee, "Meteoritic abundances of fatty acids and potential reaction pathways in planetesimals," Icarus, vol. 319, pp. 685-700, Feb. 2019, doi: 10.1016/j.icarus.2018.09.028.

2 D. W. NOONER and J. ORO, "Synthesis of Fatty Acids by a Closed System Fischer-Tropsch Process," in Hydrocarbon Synthesis from Carbon Monoxide and Hydrogen, vol. 178, 0 vols. AMERICAN CHEMICAL SOCIETY, 1979, pp. 159-171.

3 J. D. Toner and D. C. Catling, "A carbonate-rich lake solution to the phosphate problem of the origin of life," Proc. Natl. Acad. Sci., vol. 117, no. 2, pp. 883-888, Jan. 2020, doi: 10.1073/ pnas.1916109117.

4 J. D. Toner and D. C. Catling, "Alkaline lake settings for concentrated prebiotic cyanide and the origin of life," Geochim. Cosmochim. Acta, vol. 260, pp. 124-132, Sep. 2019, doi: 10.1016/j. gca.2019.06.031

5 J. G. Forsythe et al., "Ester-Mediated Amide Bond Formation Driven by Wet-Dry Cycles: A Possible Path to Polypeptides on the Prebiotic Earth," Angew. Chem. Int. Ed., vol. 54, no. 34, pp. 9871-9875, 2015, doi: 10.1002/anie.201503792.

6 C. E. Cornell et al., "Prebiotic amino acids bind to and stabilize prebiotic fatty acid membranes," Proc. Natl. Acad. Sci., vol. 116, no. 35, pp. 17239-17244, Aug. 2019, doi: 10.1073/ pnas.1900275116.

#### GO Weblink to the Poster Room zrcohen2@uw.edu

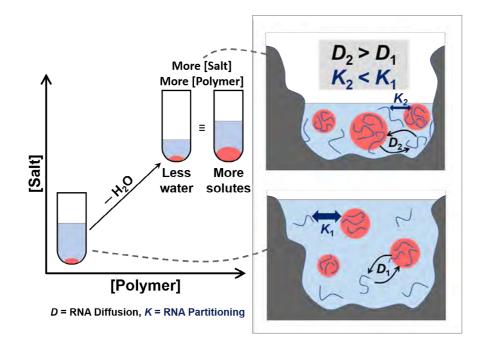
The ability of RNA to function as carriers of heritable information and enzymes has made it central to the origin and evolution of life on earth. Since the emergence and advancement of life required propagation of genetic information contained within RNA, in the absence of proteins, we are developing model systems to demonstrate protein-free RNA copying. Non-enzymatic, template-directed polymerization/ligation of intrinsically reactive, potentially prebiotic 2-aminoimidazolide (2AI) monomers/oligomers generate RNA sequences of varying complexity; however, these processes are inefficient<sup>1</sup>. Therefore, the emergence of ribozymes that use these prebiotically-relevant building blocks to assemble complex RNAs more efficiently was perhaps an essential step in the transition from chemistry to biology.

We used in vitro selection to identify ligase ribozymes that utilize 2AIactivated substrates<sup>2</sup>. These ribozymes achieved rate-accelerations of up to 1000-fold. Unfortunately, ligation of short substrates was inefficient. We, therefore, evolved ribozymes that catalyze multiple ligations joining substrates as short as 4 nt long. We also identified ligase ribozymes that function at low Mg2+, making them compatible with vesicular encapsulation by fatty acids. This presents an exciting stride toward achieving compartmentalized RNA-catalyzed RNA synthesis - a major step in the origin of life.

1 Joyce G. F. and Szostak J. W. (2018) Cold Spring Harbor Perspectives in Biology 10 (9) pii: a034801, doi: 10.1101/cshperspect.a034801

2 Walton T., DasGupta S., Duzdevich D., Oh S. S. and Szostak J. W. Proc. Natl. Acad. Sci. U. S. A.,

#### FLOOR 11



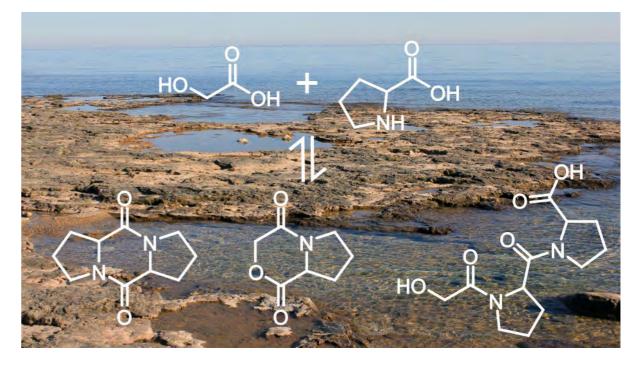
### IMPACT OF WET-DRY CYCLING ON THE PHASE & COMPARTMENTALIZATION **BEHAVIORS OF COMPLEX** COACERVATES



Hadi M.Fares<sup>1,2</sup>, Alexander E. Marras<sup>3,4</sup>, Jeffrey M. Ting<sup>3,4</sup>, Matthew V. Tirrell<sup>3,4</sup>, and Christine D. Keating<sup>1</sup> Department of Chemistry, The

Pennsylvania State University, University Park, PA 16802, USA; <sup>2</sup>NASA Postdoctoral Program, Universities Space Research Association, Columbia, MD 21046, USA; <sup>3</sup> Pritzker School of Molecular Engineering, University of Chicago, Chicago, IL 60637, USA; <sup>4</sup> Center for Molecular Engineering and Materials Science Division, Argonne National Laboratory, Lemont, IL 60439, USA

Wet-dry cycling is an environmental scenario proposed for the increase in molecular complexity on the early Earth. Alternating periods of dehydration-hydration have been shown to promote the production of molecular building blocks of life such as peptides1 and nucleosides,2 among others. Studies of the cycle's impact on the assembly of protocells and their functions have been limited. Here, we dehydrated and rehydrated model membraneless compartments made with poly(diallyldimethylammonium)/ poly(acrylic acid). We found that the process led to formation or disassembly of droplets, depending on starting concentrations of the components. The preference of an RNA oligomer to partition within the coacervate phase decreased during tenfold dehydration as its concentration remained constant inside the compartments while increasing tenfold globally. An increase in ionic strength, caused by drying, promoted faster RNA diffusion between the coacervate and its surroundings. After full dehydration, rehydration to the original volume allowed a recapture of original compartments morphology and behavior. Composition mimics, which reproduce the coacervate components concentrations at different steps of dehydration, helped in connecting the drying to linear compositional changes on the coacervate phase diagram. The results emphasize the importance of carefully considering the environment in studies of membraneless coacervate protocells as small alterations can significantly impact their compartmentalization and structural properties.



### PROLINE **INCORPORATION IN** MODEL PREBIOTIC **DEPSIPEPTIDES**

Jabbarrius N. Ervin<sup>1,2</sup>. Marcos Bouza<sup>2,3</sup>. Facundo M. Fernández<sup>2,3</sup>, and Jay G. Forsythe<sup>1,2</sup> <sup>1</sup> Department of Chemistry and Biochemistry, College of Charleston, Charleston, SC 29424, USA <sup>2</sup> NSF/NASA Center for Chemical Evolution <sup>3</sup> School of Chemistry and Biochemistry, Georgia Institute of Technology, Atlanta, GA 30332, USA

The chemical evolution of amino acids to peptides with structure and function is of great interest to origins-of-life researchers. Amino acid condensation is unfavorable in water, so various approaches have been explored which polymerize amino acids in model prebiotic scenarios<sup>1,2</sup>. Previously, we demonstrated an approach in which alpha-hydroxy acids condense into oligoesters upon heating and subsequently react with amino acids to form depsipeptides, or amino acid and hydroxy acid copolymers<sup>3,4</sup>. Here, we made mixtures of depsipeptides and compared the incorporation of proline to that of three other prebiotically-plausible amino acids: glycine, alanine, and valine. Oligomers were characterized by mass spectrometry and infrared spectroscopy. Proline was found to incorporate efficiently into linear oligomers, yet it also was detected in small cyclic byproducts. Proline is known to induce unique structural properties in biological peptides and warrants further study in the context of proto-peptide evolution.

1 Danger et al. Chem Soc Rev 2012, 41, 5416-5429;

2 Frenkel-Pinter et al. Chem Rev 2020. DOI: 10.1021/acs.chemrev.9b00664:

3 Forsythe, Yu et al. Angew Chem Int Ed 2015, 54, 9871-9875; [4] Forsythe et al. Proc Natl Acad Sci USA 2017, 114, E7652-E7659

<sup>1</sup> Rodriguez-Garcia, M.; Surman, A. J.; Cooper, G. J. T.; Suarez-Marina, I.; Hosni, Z.; Lee, M. P.; Cronin, L., Formation of oligopeptides in high yield under simple programmable conditions. Nature Communications 2015. 6

<sup>2</sup> Becker, S.; Schneider, C.; Okamura, H.; Crisp, A.; Amatov, T.; Dejmek, M.; Carell, T., Wet-dry cycles enable the parallel origin of canonical and non-canonical nucleosides by continuous synthesis. Nature Communications 2018. 9.

### NUCLEOTIDE-LEVEL RESOLUTION OF RNA FOLDING INTERACTIONS WITHIN PEPTIDE-BASED COMPLEX COACERVATES

McCauley Meyer<sup>1,2</sup>, Saehyun Choi<sup>3</sup>, Fatma Pir Cakmak<sup>3</sup>, Philip C. Bevilacqua<sup>1,2,3</sup> and Christine D. Keating<sup>3</sup> <sup>1</sup> Department of Biochemistry and Molecular Biology, Pennsylvania State University, University Park, PA 16802 <sup>2</sup> Center for RNA Molecular Biology, Pennsylvania State University, University Park, PA 16802 <sup>3</sup> Department of Chemistry, Pennsylvania State University, University Park, PA 16802 The RNA World Hypothesis states that RNA or an RNA-like polymer may have acted as both the initial genetic material and the catalyst for the reactions of life. In the 1980s, the first ribozymes were discovered, demonstrating that RNA could act as a catalyst. Since then, it has become apparent that RNA folding is integral to function in a way similar to protein enzyme folding. Because of this, it is important to try to understand RNA folding under prebiotically relevant conditions. On the early Earth, a problem that would have been faced by the first enzymes was the scarcity of organic material. To overcome this issue, organic material would need to be localized and concentrated on either a mineral surface or in some type of compartment, like a protocell. An ideal protocell candidate should partition molecules required for catalysis such as: Mg2+, nucleotides, RNAs, amino acids, and peptides. A model protocell that is able to do this is complex coacervates. Herein, I describe RNA folding studies within complex coacervate droplets made out of Lys, -Asp,, and Lys, -ATP. A model functional RNA with a well-defined three-dimensional structure, tRNA<sup>phe</sup> from S. cerevisiae, was used for these initial RNA folding studies. tRNAphe was subjected to in-line probing (ILP) under the following conditions (0.5 mM Mg<sup>2+</sup>, 15 mM KCl, and 10 mM Tris, pH 8.3) initially to determine its native fold. Then, tRNA<sup>phe</sup> was placed inside of Lys, -Asp, and Lys, -ATP coacervates where we found that under all of these coacervate conditions, the tRNA had lost its tertiary contacts and the acceptor stem was unfolded. Upon changing Mg2+ conditions and charge-ratio of polyanions to polycations, more native folding of tRNAphe was observed. Future studies will focus on evaluating if similar trends will be seen for ribozymes under the above conditions.

#### FLOOR 11

Τŀ

H  $h_{1}$   $h_{2}$   $h_{2}$  $h_{2}$ 

### ROLE OF AMINO ACIDS ON NONENZYMATIC CLAY-PROMOTED OLIGOMERIZATION OF ACTIVATED NUCLEOTIDE



Trishool Namani<sup>1</sup>, Savannah Snyder<sup>2</sup>, James Eagan<sup>1</sup>, Philip C. Bevilacqua<sup>3</sup>, Chrys Wesdemiotis<sup>2</sup>, and Nita Sahai<sup>1</sup> <sup>1</sup> Department of Polymer Science, 170 University Ave., University of Akron, Akron, OH 44325

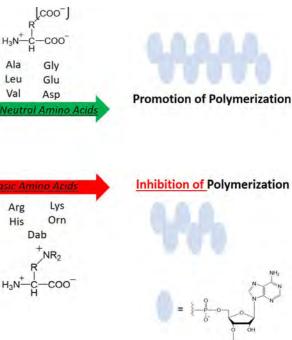
<sup>2</sup> Department of Chemistry, 190 E. Buchtel Ave., University of Akron, Akron, OH 44325

<sup>a</sup>Departments of Chemistry and of Biochemistry, Microbiology and Molecular Biology, and Center for RNA Molecular Biology, Pennsylvania State University, University Park, Pennsylvania 16802, USA.

> 1 Ferris, J. P.; E 5'-Phosphorin 2 Rajamani, S. Synthesis of R 38, 57-74 3 Kaddour, H.; RNA Oligomer Polymerization

oligomers.

mxm1357@psu.edu 💿 Weblink to the Poster Room

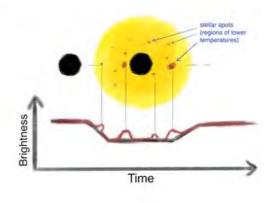


RNA biopolymer synthesis from prebiotically possible molecules is one of the important steps in the early evolution of life on Earth, hence numerous studies have reported on oligonucleotide synthesis under various experimental conditions<sup>1,2</sup>. Montmorillonite clay was extensively studied as a reaction promoter for activated nucleotides forming RNA polymer<sup>1,3</sup>. In the present study, we have examined a multicomponent system in which amino acid and montmorillonite clay both influence the polymerization of activated mononucleotide, adenosine 5'-phospho-2-methylimidazolide (2-MeImPA) polymerization at neutral pH. The polymerization of 2-MelmPA is further promoted in the presence of acidic and neutral nonpolar amino acids, which results in longer oligomers than the clay system alone. Positively charged amino acids, by comparison, exhibit an inhibitory effect and create shorter oligomers. To understand the impact of amino acids on 2-MeImPA polymerization, we hypothesize: 1) the acidic and non-polar amino acids are anchored to the negatively charged clay surface sites through their alpha ammonium group, while the alpha carboxylate interacts with the methylimidazolium ring of the activated nucleotide providing an ideal conformation to convert the activated nucleotide into a better leaving group, thus creating longer oligomers, 2) the basic amino acids bind with their basic side chain to the montmorillonite surface and compete with the mononucleotide for adsorption to the clay surface, thus resulting in shortened

1 Ferris, J. P.; Ertem, G. Oligomerization of Ribonucleotides on Montmorillonite: Reaction of the 5'-Phosphorimidazolide of Adenosine. Science (1992) 257, 1387–1389 2 Rajamani, S.; Vlassov, A.; Benner, S.; Coombs, A.; Olasagasti, F.; Deamer, D. Lipid-Assisted Synthesis of RNA-Like Polymers from Mononucleotides. Origins Life Evol. Biospheres (2008)

3 Kaddour, H.; Gerislioglu, S.; Dalai, P.; Miyoshi, T.; Wesdemiotis, C.; Sahai, N. Non-Enzymatic RNA Oligomerization at the Mineral-Water Interface: An Insight into the Adsorption-Polymerization Relationship. J. Phys. Chem. C (2018) 122, 29386–29397

Using Machine Learning to **Improve Prediction of Chemical Composition of Exoplanetary Atmospheres** 



### USING MACHINE LEARNING TO IMPROVE PREDICTION OF CHEMICAL COMPOSITION OF EXOPLANETARY **ATMOSPHERES**



Artash Nath Co-founder, HotPopRobot.com Humankind does not currently possess instruments to detect life on exoplanets - planets outside the solar system. But efforts are ongoing to come up with newer ground and space-based telescopes that could help us learn more about atmos-pheres of these exoplanets. When an exoplanet transits in front of its parent star, its main body blocks out some light of the star. This causes a dip in the light received from the star. If the exoplanet has an atmosphere around it, then the atmosphere will also absorb some of this light. How much light is absorbed by the atmosphere depends on its thickness and gases present.

Different gases absorb different wavelengths of light to different degrees. If we plot the transit of an exoplanet in different wavelengths, we will get light curves of different depths. Studying transit light curves of exoplanets in different wave-lengths could help us predict the chemical composition of their atmospheres.

However, the parent star of the exoplanet may have stellar spots that are cooler than the surrounding surface. This adds noise in the data We must isolate depth in light curves caused by the exoplanetary atmosphere from those caused by the stellar spots. The current approach is to remove this noise manually which is time consuming and prone to errors

Applying machine learning to exoplanetary data may help remove the noise of star-spots in data on transiting exoplanets' atmospheres received by space tele-scopes. I created a Hybrid Machine Learning model using Long-Short Term Memory (LSTM) - a form of Recurrent Neural Network (RNN) to reduce this noise. My model was able to accurately predict the exoplanet-star radius ratio in 55 wavelengths with a mean square error of 0.001. The Algorithm leads to elimination of noise and may lead to improved and accu-rate prediction of chemical composition of exoplanetary atmospheres.

The simulated dataset I used for my project can be accessed from the ARIEL Space Telescope Machine Learning Challenge website at: https://ariel-datachallenge.azurewebsites.net/ML

FLOOR 11

### EXOPLANETARY **ATMOSPHERIC** RETRIEVAL USING MACHINE LEARNING

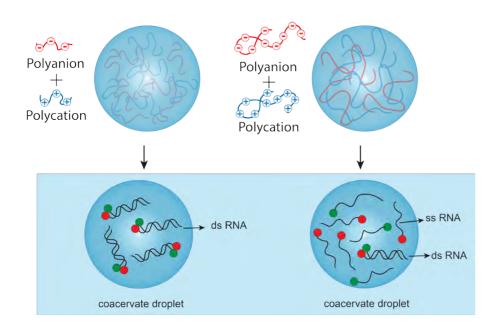
#### Aline Novais<sup>1</sup>, Luan Ghezzi<sup>1</sup>, Kevin Heng<sup>2</sup>

<sup>1</sup> Valongo Observatory, Federal University of Rio de Janeiro, Brazil <sup>2</sup> Center for Space and Habitability, University of Bern, Switzerland

Although the analysis of exoplanet atmospheres has become one of the most pertinent topics within planetary science, characterizing these objects directly from their spectra might still be a challenge. To interpret the spectrum of an exo-atmosphere, one can apply an inverse technique known as atmospheric retrieval, which is the use of an observed transmission spectrum to infer planetary properties, such as its temperature profile, chemical composition and atmospheric circulation. This work establishes if the stellar radius' and gravity of the exoplanet's uncertainties cause non-negligible effects in currently available HST Wide Field Camera 3 (WFC3) and Space Telescope Imaging Spectrograph (STIS) spectra and in future spectra measured by JWST. We intend to determine when the effects of critical spectral resolution and wavelength coverage are important, so as to guide future procurement of exo-atmospheric spectra. Consequently, we will establish the conditions under which the analytical formula for isothermal, isobaric transit chords breaks down and a full numerical treatment is needed instead.

1 Fisher, C., & Heng, K. 2018, , 481, 4698, doi:10.1093/mnras/sty2550 2 Heng, K., & Kitzmann, D. 2017, , 470, 2972, doi:10.1093/mnras/stx1453 3 Kreidberg, L., Line, M. B., Bean, J. L., et al. 2015, 1814, 66. doi:10.1088/0004-637X/814/1/66 4 Márguez-Neila, P., Fisher, C., Sznitman, R., & Heng, K. 2018, Nature As-tronomy, 2, 719, doi:10.1038/s41550-018-0504-2





## PREBIOTICALLY-**RELEVANT LOW POLYION** MULTIVALENCY MIGHT IMPROVE FUNCTIONALITY OF MEMBRANELESS COMPARTMENTS



Fatma Pir Cakmak<sup>1</sup>, Saehyun Choi<sup>1</sup>, McCauley O. Meyer2,3, Philip C. Bevilacqua<sup>1,2,3</sup> and Christine D. Keating<sup>1</sup> Department of Chemistry, The Pennsylvania State University, University Park, Pennsylvania 16802, USA <sup>2</sup>Center for RNA Molecular Biology, The Pennsylvania State University, University Park, Pennsylvania 16802, USA <sup>3</sup>Department of Biochemistry and Molecular Biology, The Pennsylvania State University, University Park, PA

16802, USA

Multivalent polyions can undergo complex coacervation, producing membraneless compartments that accumulate ribozymes and enhance catalysis, and offering a mechanism for functional prebiotic compartmentalization in the origins of life. Here, we evaluated the impact of low, prebiotically-relevant polyion multivalency in coacervate performance as functional compartments. As model polyions, we used positively and negatively charged homopeptides with one to 100 residues, and adenosine mono-, di-, and triphosphate nucleotides. Polycation/polyanion pairs were tested for coacervation, and resulting membraneless compartments were analyzed for salt resistance, ability to provide a distinct internal microenvironment (apparent local pH, RNA partitioning), and effect on RNA structure formation. We find that coacervates formed by phase separation of the relatively shorter polyions more effectively generated distinct pH microenvironments, accumulated RNA, and preserved duplexes. Hence, reduced multivalency polyions are not only viable as functional compartments for prebiotic chemistries, but they can offer advantages over higher molecular weight analogues.

doi: https://doi.org/10.1101/2020.02.23.961920

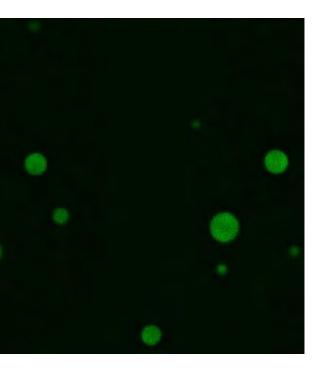
### RNA WORLD INSIDE COMPARTMENTS: ACTIVATION OF RNA CATALYSIS BY COMPLEX COACERVATES



Raghav R. Poudyal<sup>1</sup>, Rebecca M. Guth-Metzler<sup>3\*</sup>, Andrew J. Veenis<sup>1</sup>, Erica A. Frankel<sup>1#</sup>, Christine D. Keating<sup>1,2</sup>, Philip C. Bevilacqua<sup>1,2,3</sup> <sup>1</sup> Department of Chemistry, The Pennsylvania State University, University Park, Pennsylvania 16802, USA <sup>2</sup>Center for RNA Molecular Biology, The Pennsylvania State University, University

Park, Pennsylvania 16802, USA <sup>3</sup>Department of Biochemistry and Molecular Biology, The Pennsylvania State University, University Park, PA 16802, USA

\*Present address: School of Chemistry and Biochemistry, Georgia Institute of Technology, Atlanta, GA, 30332, USA #Present address: The Dow Chemical Company, 400 Arcola Road, Collegeville, PA, 19426, USA L



Complex coacervates are non-membranous compartments (NMCs) formed by associative phase separation of oppositely-charged polyelectrolytes. Owing to their ability to encapsulate biomolecules, they have been postulated as alternative prebiotic compartments. In this study, we report that NMCs of complex coacervates can concentrate RNA oligonucleotides and activate catalysis of multiple nucleic acid enzymes, including a deoxyribozyme. Furthermore, we reveal that polyanions can tune microenvironments of NMCs to further enhance RNA catalysis. By competing for unproductive RNA- polycation interactions, short polyanions enhance ribozyme reactions more than 12-fold. Productive RNA interactions in NMCs that are otherwise inaccessible in dilute solutions reveal potential roles for these compartments in the context of origin of life and extant biological intracellular condensates.



### DEFINING LYFE IN THE UNIVERSE: FROM THREE PRIVILEGED FUNCTIONS TO FOUR PILLARS



Stuart Bartlett<sup>1,2</sup> & Michael L. Wong<sup>3,4</sup> Division of Geological & Planetary Sciences, California Institute of Technology, Pasadena, CA 91125, USA <sup>2</sup>Earth-Life Science Institute, Tokyo Institute of Technology, Tokyo 152-8550, Japan



<sup>3</sup> Department of Astronomy & Astrobiology Program, University of Washington, Seattle, WA 98195, USA <sup>4</sup>NASA Nexus for Exoplanet System Science's Virtual Planetary Laboratory University of Washington, Seattle, WA, 98195, USA

Motivated by the need to paint a more general picture of what life is-and could be-with respect to the rest of the phenomena of the universe, we propose a new vocabulary for astrobiological research. Lyfe is defined as any system that fulfills all four processes of the living state, namely: dissipation, autocatalysis, homeostasis, and learning (see Figure)<sup>1</sup>. Life is defined as the instance of lyfe that we are familiar with on Earth, one that uses a specific organometallic molecular toolbox to record information about its environment and achieve dynamical order by dissipating certain planetary disequilibria. This new classification system allows the astrobiological community to more clearly define the questions that propel their research-e.g., whether they are developing a historical narrative to explain the origin of life (on Earth), or a universal narrative for the emergence of lyfe, or whether they are seeking signs of life specifically, or lyfe at large across the universe. While the concept of "life as we don't know it" is not new, the four pillars of lyfe offer a novel perspective on the living state that is indifferent to the particular components that might produce it.

1 Bartlett, S and Wong, M L (accepted) Defining Lyfe in the Universe: From Three Privileged Functions to Four Pillars, Life

#### FLOOR 11

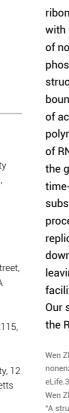
### DECIPHERING NONENZYMATIC RNA POLYMERIZATION THROUGH CRYSTALLOGRAPHY



Wen Zhang<sup>1</sup>, Jack W. Szostak<sup>2,3,4</sup> <sup>1</sup> Department of Biochemistry and Molecular Biology, Indiana University School of Medicine, 635 Barnhill Dr., Indianapolis, IN 46202, USA <sup>2</sup>Howard Hughes Medical Institute, Department of Molecular Biology and Center for Computational and Integrative Biology, Massachusetts General Hospital, 185 Cambridge Street, Boston, Massachusetts 02114, USA <sup>3</sup>Department of Genetics, Harvard Medical School, 77 Avenue Louis Pasteur, Boston, Massachusetts 02115, LISA

<sup>4</sup>Department of Chemistry and Chemical Biology, Harvard University, 12 Oxford St., Cambridge, Massachusetts 02138, USA

2017, 114(29), 7659-7664.



Many high resolution crystal structures have contributed to our understanding of the reaction pathway for catalysis by DNA and RNA polymerases, but the structural basis of nonenzymatic templatedirected RNA replication has not been studied in comparable detail. Here we present crystallographic studies of the binding of ribonucleotide monomers to RNA primer-template complexes, with the goal of improving our understanding of the mechanism of nonenzymatic RNA copying. Using our synthetic unreactive phosphonate-linked analog of activated monomer, we obtained the structures of RNA primer-template complexes with the monomers bound. Our structures demonstrate the versatile binding motifs of activated mononucleotide substrate in nonenzymatic RNA polymerization, which could significantly influence the rate and fidelity of RNA replication, and they also illustrate the structural rationale of the great catalytic function of the downstream helpers. In addition, our time-resolved structures successfully integrate several static RNAsubstrate structures into a molecular "movie" following the reaction process, and it clearly reveals the mechanism of RNA nonenzymatic

replication. Against the traditional opinion that the catalysis of downstream activation is based on the noncovalent leaving groupleaving group effect, our time-resolved structures demonstrate that the facilitation is from the formation of imidazolium-bridged intermediate. Our structures provide the powerful tool and fundamental evidence for the RNA self-replication mechanistic studies.

Wen Zhang, Travis Walton, Li Li and Jack W. Szostak\*, "Crystallographic observation of nonenzymatic RNA primer extension", eLife, 2018, 7:e36422. DOI: https://doi.org/10.7554/ eLife.36422. (Highlighted by Nature Chemical Biology News, 2018, 14, 745.) Wen Zhang, Chun Pong Tam, Lijun Zhou, Seung Soo Oh, Jiawei Wang, and Jack W. Szostak\*, "A structural rationale for the enhanced catalysis of nonenzymatic BNA primer extension by a

downstream oligonucleotide", Journal of American Chemical Society, 2018, 140(8), 2829-2840. Wen Zhang, Chun Pong Tam, Travis Walton, Gabriel Birrane, and Jack W. Szostak\*, "Insight into the mechanism of nonenzymatic RNA primer extension from the structure of an RNA-GpppG complex", Proceedings of the National Academy of Sciences of the United States of America,



#### INDEX - FLOOR 12 (UTC -04:00 TO -07:00)

Visit at late hours of the event or during poster session

- 159 MARION ZULEMA ARMAS VAZQUEZ Computational study of physicochemical properties of an adenine synthesis route under UV radiation
- 160 **JULIAN CORZO** Endolithic culturable bacteria in minerals from geologic samplings in Colombia
- 161 **ROMULO LEONCIO** Chemical oscillations in a theoretical system of dinitrosyl iron complex (DNIC) with thiol-containing ligands
- 162 LUIS DELAYE Was LUCA a hyperthermophilic prokaryote? The impact-bottleneck hypothesis revisited
- 163 FRANKY DJUTANTA Producing cell-like structures from oil films residing on ocean water by raindrop impacts
- 165 **GIOVANNA GHIRLANDA** Membraneless organelles by design
- 166 ROSA REYES What about the origin of death?
- 167 DAVID RODRIGUEZ Origin of life on ice. Is Mars a potential habitable planet?
- 168 ALFREDO RODRÍGUEZ ARTEAGA Extremophilic proteins and their resistance secrets
- 169 LUIS MIGUEL RODRÍGUEZ TORRES Contrast between the main life's origins abiogenic models
- 170 TOVILLA QUESADA RUBÉN DE JESÚS The importance of laboratory practices on the origin of life at the upper-middle level

#### **FLOOR 12**

## COMPUTATIONAL STUDY OF PHYSICOCHEMICAL **PROPERTIES OF AN** ADENINE SYNTHESIS ROUTE UNDER UV RADIATION

#### M. Armas<sup>1,2</sup>, A. Segura<sup>2</sup>, A. Heredia<sup>2</sup>

<sup>1</sup> Posgrado en Astrofísica, Universidad Nacional Autónoma de México, México. <sup>2</sup> Instituto de Ciencias Nucleares. Universidad Nacional Autónoma de México. México.

One of the most important topics of the origin of life studies is the searchfor plausible routes of prebiotic synthesis of fundamental molecules for living beings. Since the experiments carried out by Miller up to date, many environments have been simulated for the conditions of the Earth 3.8 Ga ago. They have managed to synthesize nucleotides, amino acids, among other essential components for life. All these studies have found that in order to trigger the synthesis reactions of molecules of prebiotic importance, it is necessary to supply some type of energy such as temperature, electric discharges, radiation, etc(Barks etal. 2010)). The surface of the Earth during the Archean period (approximately 3.8 Ga) was exposed to higher doses of UVC radiation (200-280 nm) and UVB (280-315 nm) in comparison with the radiation that arrives to the Earth today (Sagan, 1973; Cockell, 1998). The reason for thishigh UV irradiation was thelack of layer of ozone and higher emissionof the Sun in this wavelength range. In this sense, UV radiation might have played animportant role in the evolution of prebiotic chemistry. In an astrobiological context, all stars emit UV radiation; particularly dwarf M-type stars (Hunt-Walker et al., 2012). TheUV flux on a planetary surface of a potentially habitable hypothetical planetdependson its atmospheric composition and the spectral type of the hosting star. This fact might thus change the potential chemical reactivity and synthesis of some organic compounds on the planet (Rugheimer et al., 2015). For our work we analyzed a known route of adenine prebiotic synthesis from formamide using the HyperChem software.We analyzed some physicochemical properties (oscillator strength, energy difference between the HOMO-LUMO orbitals and dipolar moment among others) of intermediate molecules in this path under the interaction with an UV photon. Here, we present the preliminary results on how UV radiation affects these molecules of prebiotic importance and the possible triggering effect of UV radiation to promote physicochemical reactions for its production.



### ENDOLITHIC CULTURABLE **BACTERIA IN MINERALS** (QUARTZ. K-FELDESPAR. CALCITE) FROM **GEOLOGIC SAMPLING** IN VILLA DE LEYVA. BOYACA & PESCADERO. SANTANDER (COLOMBIA)

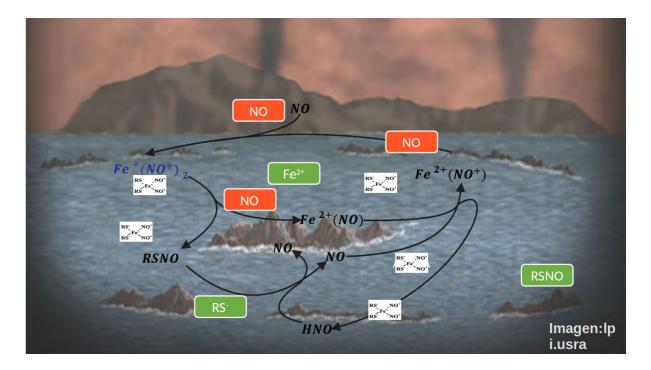


Corzo-Acosta, J.<sup>1</sup>, Tchegliakova, N.<sup>1</sup>, Corzo, J.<sup>2</sup>

Geosciences Department, National University of Colombia Statistics Department, National University of Colombia

Endolithic microbial communities have been reported in arid and hyperarid zones around the world. Also, it has been found that bacteria use as microhabitat different lithologies, whose chemical elements may act as a nutritional source. This study focuses on the relation between culturable endolithic microbes and three rock-forming minerals (quartz, k-feldspar and calcite) in order to determine the potential of these minerals to serve both as a microhabitat and as a source of nutrients. Geological data collected in two localities in Colombia: Villa de Leyva, Boyacá (5°36' 24.12" N, 73° 31' 31.32" W) and Aratoca, Santander (6°49' 0.62" N, 73°0' 20.3" W) along with microbiological data obtained in the laboratory provide information about the affinity between native endolithic microbial communities and the three mentioned minerals. In order to determine geochemical characteristics of the minerals X-Ray Fluorescence (XRF), environmental scanning electron microscopy (ESEM), and Petrographic analysis were carried out. Additionally, microbiological procedures [culture techniques, gram stain and physicochemical profile] were performed to characterize the colonies of native endolithic culturable microorganisms along with climatological data from the study areas. Multiple Correspondence Analysis (MCA) was used to discover whether there are relationships between microbes and some abiotic conditions. In this study we discuss that rock-forming minerals calcite, K-feldspar and quartz act as a potential microhabitat for microbes and that its bio-receptivity could be useful for the search of extraterrestrial life on Mars surface.

#### **FLOOR 12**



CHEMICAL OSCILLATIONS IN A THEORETICAL SYSTEM OF DINITROSYL IRON COMPLEX (DNIC) WITH THIOL-CONTAINING LIGANDS



Romulo Cruz-Simbron<sup>1,2</sup>, Gino Picasso<sup>1</sup>, Yannick de Decker

Laboratory of Physical Chemistry Research, Faculty of Sciences, National University of Engineer-ing, Tupac Amaru 210 Av., Rimac, Lima, Peru <sup>2</sup> Scientific Society of Astrobiology of Peru (SCAP), Lima, Lima <sup>3</sup>Nonlinear Physical Chemistry Unit and Center for Nonlinear Phenomena and Complex Systems (CENOLI), Université libre de Bruxelles, Boulevard du Triomphe. C.P. 231, B-1050 Brussels. Belgium

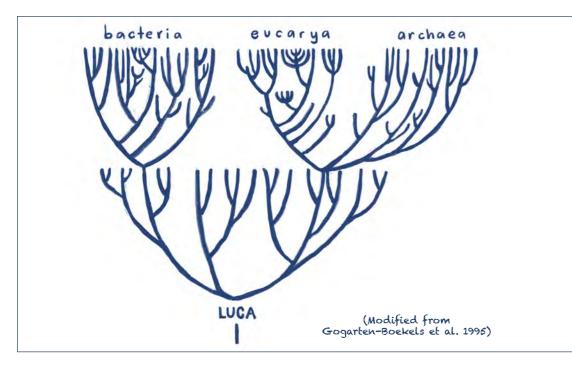
studied. 9-15

Anatoly Vanin and his research group have been involved for more than 50 years in an extensive study on the preponderant role of the iron dinitrosil complexes (DNICs) in biological systems. Owing to its simple structures, it constitutes a work-ing form of endogenous nitric oxide in alive systems. The involvement of nitric oxide as a universal regulator present in all known life forms led to the important conclusion regarding the leading role of that have played the DNICs both at the present and in the past in the definition of the metabolic processes of living be-ings. Ferrous ion and nitrosothiols are the building blocks through an enriched chemistry stand out over other regulatory mechanisms. Vanin's reaction be-tween the ferrous ion and nitrosothiols have shown attenuated fluctuations in the DNIC concentration in a time interval of 500 to 2500 s. Considering the high pos-sibility of real oscillations in the Vanin's work, the present study has reanalyzed the kinetic model proposed by Dr. Vanin in his original work<sup>1</sup> and investigated other possible mechanisms linking the traditional Vanin model with chemical os-cillations. The simplicity of this mechanism, and the possibility that it does not involve advanced biochemical species such as enzymes, allow us to infer that it is very likely that this is a mechanism of temporal-spatial regulation<sup>2</sup> is very old and associated with each domain of life, even early life3. As Epstein has indicated4 if the experiments and the mechanism agree, we can maintain the mechanism and try some more experiments; for this reason, this work seeks to motivate to continue investigating experimentally the oscillations that can oc-cur in this type of systems that Vanin has

3 Ducluzeau, A. L., Van Lis, R., Duval, S., Schoepp-Cothenet, B., Russell, M. J., & Nitschke, W. (2009). Was nitric oxide the first deep electron sink?. Trends in biochemical sciences, 34(1),

4 Epstein, I. R., & Pojman, J. A. (1998). An introduction to nonlinear chemical dynamics: oscillations, waves, patterns, and chaos, Oxford University Press,

<sup>1</sup> Vanin, AF, Papina, AA, Serezhenkov, VA, & Koppenol, WH (2004). The S-nitrosothiol Mechanisms of decomposition catalyzed by iron. Nitric oxide, 10 (2), 60-73. 2 Vanin, A. F., Mikoyan, V. D., Rubtsov, N. M., & Kubrina, L. N. (2010c). Autowave distribution of nitric oxide and its endogenous derivatives in biosystems strongly enhances their biological effects: A working hypothesis. Nitric Oxide, 23(3), 175-180.



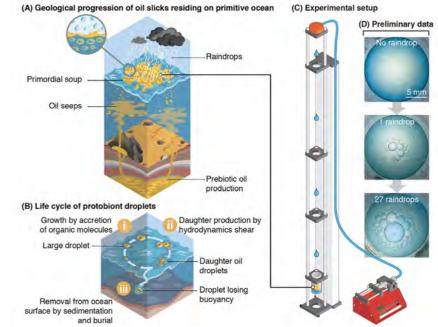
### WAS LUCA A **HYPERTHERMOPHILIC PROKARYOTE? THE IMPACT-BOTTLENECK** HYPOTHESIS REVISITED



Gilberto P. Morales and Luis Delave Departamento de Ingeniería Genética, Cinvestav-Irapuato, Km. 9.6 Libramiento Norte Carretera Irapuato-León, 36821, Irapuato, Guanajuato, Mexico

In the Origin of Species, Darwin wrote "The affinities of all the beings of the same class have sometimes been represented by a great tree. I believe this simile largely speaks the truth." Modern comparative genomics has revealed that the intuition of Darwin was correct. A set of highly conserved genes and cellular functions indicate that all life on Earth is related by common ancestry. These genes were inherited from the Last Universal Common Ancestor or LUCA. The functions coded by these genes suggest that LUCA was a rather complex cell already endowed with a genetic code and a protein translation apparatus. One of the questions regarding the nature of LUCA is whether it was a hyperthermophile. Here, we review recent evidence derived from the molecular fossil record on the temperature preferences of LUCA. We suggest that current evidence on the nature of LUCA and its immediate predecessors are compatible with the impact-bottleneck hypothesis the proposal that during the early evolution of life a meteoritic impact eliminated all life on Earth except for prokaryotes capable of living at high temperatures. If our interpretation of the data is correct, it would indicate that early life was resilient to the rough environmental conditions of the Archean. A relevant result from the point of view of astrobiology because it would exemplify the persistence of life in harsh environments.

**FLOOR 12** 



### PRODUCING CELL-LIKE STRUCTURES FROM OIL FILMS RESIDING ON **OCEAN WATER BY** RAINDROP IMPACTS

#### Franky Djutanta<sup>1,2</sup>, Rachael Kha<sup>1,2</sup>, Bernard Yurke<sup>3</sup>, and Rizal F. Hariadi<sup>1,4</sup>

<sup>1</sup> Biodesign Center for Molecular Design and Biomimetics (at the Biodesign Institute) at Arizona State University, Tempe, AZ85287, USA

<sup>2</sup>School for Engineering of Matter, Transport and Energy, Arizona State University, Tempe, AZ 85287, USA <sup>3</sup> College of Engineering, Boise State University, Boise, ID 83725. USA

<sup>4</sup>Department of Physics, Arizona State University, Tempe

Fig. 1. Raindrop impacting prebiotic soup of oil film residing on ocean water produces protocell "droplets"

(A) Schematic of proposed geological progression of oil slick production on primitive ocean followed by action from raindrop impact forming oil droplets and cell-like structure droplets

(B) Life cycle of the proposed protobionts of oil droplets.

(C) Experimental setup to generate raindrop impacting on oil film residing on ocean

(D) Preliminary data of droplet formation at indicated raindrop numbers

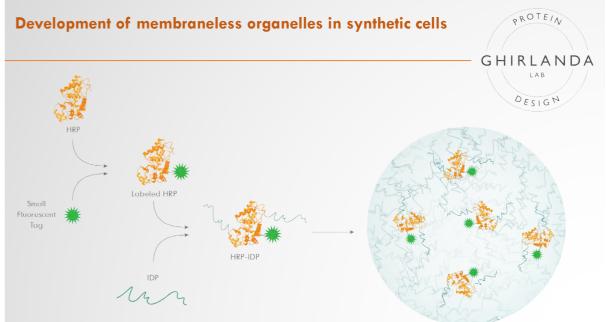
Progress in the bottom-up construction of synthetic cells<sup>1</sup> has given insights into how biological organisms function and on the origins of life. Despite advances in the last decades, a demonstration of an open-ended in-vitro evolution of synthetic cells that is simple enough to serve as a plausible model of the prebiotic/biotic transition has yet to be demonstrated. Here, we develop a synthetic cell platform based on geophysical considerations. We model the protocell as oil-inwater droplets. While there have been several works demonstrating oil-in-water droplets exhibiting the behaviors that protocell may encounter,<sup>2,3</sup> there have yet a complete demonstration of oil-in-water droplet that can undergo a complete life cycle involving birth, growth, replication, and death, within a plausibly prebiotic environment. The proposed model centers on the ubiguitous mechanical processes in the dispersal of natural and manmade oil slicks in modern oceans. In the prebiotic world, the organic matter, primarily delivered to the earth's surface by micrometeorites, would have naturally accumulated to ocean surface as oil slicks.<sup>4</sup> Additionally, these organic compounds were, perhaps, prevented from reaching shore by entrapment in ocean gyres. Hydrodynamics forces such as raindrops or breaking waves on these thick organic films, resembling a primordial soup, would have produced droplets (Fig. 1A). Similar notion was proposed by Oparin where hydrodynamic forces play an important role on the origins of life by enabling Darwinian evolution in the fragmentation of coacervates through hydrodynamic shear.<sup>5</sup> Comparably, the oil droplets could have undergone growth via accumulation of organic compounds (Fig. 1B-i), fission via locally intense hydrodynamic shear produced by rainfall or breaking waves (Fig. 1B-ii), and eventually death by burial via the loss of buoyancy through mineral accumulation (Fig. 1B-iii). In short, these droplets could have functioned as simple selfreplicators that natural selection could act to select droplets possessing more fit chemical and structural composition, and ultimately giving rise to the protobionts. Finally, we will present results from preliminary laboratory experiments that explore the rich fluid-mechanics phenomena involved in the production of oil droplets from hydrodynamics forces acting on oil films residing on water surfaces and in the fissioning of oil droplets (Fig.

1C). In particular, raindrop impacting oil film residing on water also produces water-in-oil-in-water (w/o/w) droplets that resemble lipid vesicles in that a volume of water is surrounded by film of oil (Fig. 1D). Such droplets may have facilitated the prebiotic/biotic transition. Stability test reveals that the droplets can maintain its identity up to three days if PEG-octyl-ether is used as the surfactant. In contrast, the droplets are only stable on a time scale of minutes for PEG600-cholesterol surfactant. Further experimental development may lead to a bench-top system in which oil droplets undergo growth and fission, and are

maintained at a steady state concentration through continual removal of oil droplets which is analogous to a death process in natural environment. The population of oil droplets maintained by such a system could serve as a platform for in-vitro evolution.

- 1. Blain JC, Szostak JW (2014) Progress toward synthetic cells. Annual review of biochemistry 83:615-640.
- 2. Lach S, Yoon SM, Grzybowski BA (2016) Tactic, reactive, and functional droplets outside of equilibrium. Chemical Society Reviews 45(17):4766-4796.
- 3. Dzieciol AJ, Mann S (2012) Designs for life: protocell models in the laboratory. Chemical Society Reviews 41(1):79-85.
- 4 Maurette M Brack A (2006) Cometary petroleum in badean time? Meteoritics and Planetary Science 41:5247
- 5. Oparin AI (1924) Proiskhozhdenie zhizny (The origin of life, Ann. Synge. Trans.).

#### **FLOOR 12**



#### MEMBRANELESS ORGANELLES BY DESIGN



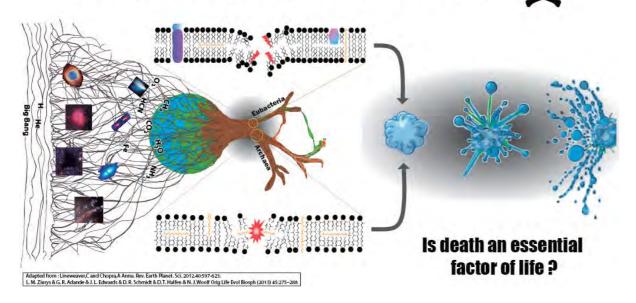
Miichele Costantino, Saul Favila, Matthias Heyden, Sara Vaiana, and Giovanna Ghirlanda

School of Molecular Sciences, Arizona State University, Tempe, AZ<sup>2</sup> 2209 Mill Race Drive, Chapel Hill, NC 27514, USA equilibrium.

Compartmentalization is at the basis of cellular organization. Traditional organelles are delineated from the cytoplasm with a semipermeable lipid bilayer and are considered permanent structures. In addition, cells comprise functional membraneless organelles such as the Cajal body and the nucleolus, in both the nucleus and the cytoplasm, that contain high concentrations of proteins. It has been recently established that these structures form by liquid liquid phase separation of nucleic acids and/or proteins, and carry out diverse functions in the cell. They form and disperse in response to the cellular environment and, without a membrane, are accessible to the surroundings through

Our group is designing functional membraneless organelles formed by liquid-liquid phase separation as compartments for synthetic cells. Here, we use fused in sarcoma (FUS) and a DEAD-box helicase (Ddx4), which form LLPS droplets in certain conditions, as elements to drive the localization of enzymes into an artificial organelle. Specifically, the sequences corresponding to wild-type FUS (residues 1-214, from here on defined as N-FUS), and to wild-type Ddx4 (residues 1-236, N-Ddx4), were conjugated to horseradish peroxidase. The phase diagram describing the spinodal and binodal transition to form LLPS droplets is experimentally determined by DLS, both before and after conjugation. Fluorescent microscopy was used to confirm the presence of conjugated enzymes within the concentrated droplets. We are currently investigating the activity of enzymes within the droplets. Finally we will test the effectiveness of LLPS droplets in organizing and enhancing the activity of enzymatic cascades, starting with two well-known model systems: the two-enzyme GOx/HRP system, and the more challenging three enzyme cascade that forms the methanol dehydrogenase pathway (ADH/ALDH/FDH). Future work will address the enzymatic efficiency of the system through absorbance of the final product in kinetic assays.

#### From the origin of life to the origin of 🧔



### WHAT ABOUT THE **ORIGIN OF DEATH?**



#### Rosa Reves

Grupo de Ciencias Planetarias y Astrobiología, Departamento de Biología Departamento de Geociencias, Universidad Nacional de Colombia. Carrera 45 # 26 - 85 Edificio Uriel Gutiérrez, Bogotá D.C., Colombia.

The emergence of life implies the origin of an archaic form of death. From our understanding of primitive cells, the death state must have been based on thermodynamic/material starvation<sup>1</sup>, a phenomenon understood as the irreversible transition of a system from the dynamic kinetic stability status, of the replicative world, to the thermodynamic world<sup>2</sup>.

The adaptative responses that early life set towards different agents that compromise cell vitality, such as: temperature shock, oxidative stress, DNA damage, and pathogens, constituted the evolutionary origin of senescence<sup>3</sup>. Nowadays, when modern prokaryotic cells accumulate endogenous damage, they enter a non-proliferating state ruled by many molecular alterations that could have been established during the conformation of LUCA4.

Additionally, collapse and depolarization of the membrane generates a decline in the ATP levels and the release of free radicals, a classic sequence that triggers programmed cell death<sup>5</sup>. However, LUCA's membrane type, respiratory complexes and ATP-synthase status remains a subject of discussion between two main approaches for life's site of origin: the sub-areal hot springs and the hydrothermal vent hypothesis. Environmental conditions on these sites, proton permeability, and membrane antiporters nature would have evolved different properties, implying distinct responses to agents that may cause the first death of a living cell.

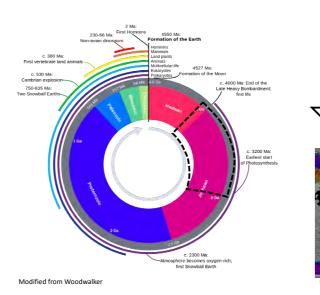
1 Popa, R. (2007). Between Necessity and Probability: Searching for the Definition and Origin of Life. I.J. of Astrobiology

2 Pross, A (2012). What is life? How Chemistry becomes Biology. Oxford University Press. 3 Golubev, A., Hanson, A. D., & Gladyshev, V. N. (2018). A Tale of Two Concepts: Harmonizing the Free Radical and Antagonistic Pleiotropy Theories of Aging. 1003-1017. A & R.S. 4 Pan, D. (2004). Topics in Current Genetics. Model Systems in Aging. Sprinter. 5 Lane, N. (2015). The Vital Question: Why is life the way it is? P. Books LTD.

#### **FLOOR 12**

#### Origin of life on ice. Is it Mars a potential habitable planet?

Tovar, D<sup>1,2</sup>., and Sánchez, M<sup>1</sup>. Grupo de Ciencias Planetarias y Astrobiología GCPA, Universidad Nacional de Colombia. Bogotá, Colombia 2. Facultad de Educación, Universidad de La Sabana. Chía, Colombia



## ORIGIN OF LIFE ON ICE. IS MARS A POTENTIAL HABITABLE PLANET?

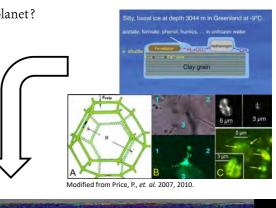


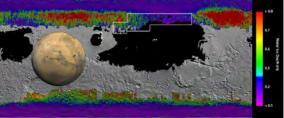
#### Tovar. D1, Sanchez. M2 <sup>1</sup> Grupo de

Ciencias Planetarias y Astrobiología, Universidad Nacional de Colombia. Carrera 45 # 26 – 85. Bogotá, Colombia. <sup>2</sup> Grupo de Ciencias Planetarias y Astrobiología, Universidad Nacional de Colombia, Carrera 45 # 26 - 85, Bogotá, Colombia.

on Mars currently<sup>4,5</sup> thrive<sup>2,5</sup>.

perspectives in biology and Space Sci. Microbiol. Ecol.





Modified from Piqueux, S., et. al. 2019.

Broad consensus among scientific community on the origin of life on Earth, suggests that it may have originated from hydrothermal underwater vents at temperatures close to 250 ° C1. However, low concentration of greenhouse gases such as CO2 and water vapor in the Archean, contribute to ubiguitous frozen oceans<sup>2,3</sup>.

Frozen oceans are usually associated in most cases with Earth's poles. An atmosphere generated by degassing due to asteroid impacts instead of continuous degassing, indicates that the presence of greenhouse gases is low at the primitive Earth<sup>2,3</sup>. During the Archaean, the relationship between the rate of microbial metabolism in ice and the rate of experimentally determined production of trapped gases of microbial origin, might explain the rate of methane production as a function of temperature from their habitat. This could be applied even

These reactions exhibit geophysical and geochemical characteristics that can be found at very low temperatures, since the interface of prebiotic activity with icy terrestrial environments is a dynamic environment where these microbes have been found to eventually

1 Macleod, G., McKeown, C., Hall, A. J. and Russell, M. J.: (1994). Hydrothermal and pH Conditions of Possible Relevance to the Origin of Life. Origins Life Evol. Biosphere 2 Bada, J., Bighma, C., and Miller, S.L. (1994). Impact melting of frozen oceans on the early Earth: Implications for the origin of life, Proc. Natl. Acad. Sci.

3 Zahnle, K., Schaefer, L., & Fegley, B. (2010). Earth's earliest atmospheres. Cold Spring Harbor

4 Price, P.B. (2010). Microbial life in martian ice. Abiotic origin of methane on Mars. Planetary

5 Price, P.B. (2007). Microbial life in glacial ice and implications for a cold origin of life. FEMS

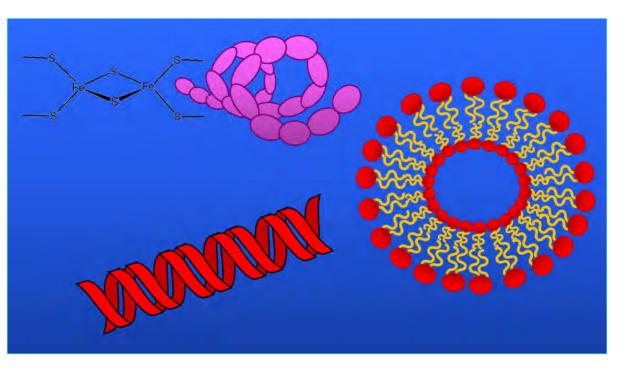
### EXTREMOPHILIC PROTEINS AND THEIR RESISTANCE **SECRETS**

Alfredo Rodríguez Arteaga Institute of Biotechnology of the UNAM, Mexico

All the cells of all the organisms we know have proteins that function as molecular factories to make copies of their genetic material and, of course, have ways to repair it from almost any damage they are exposed to on earth. Like the cases of extremophiles that survive in places where we cannot. The proteins of the extremophiles reveal to us their peculiarities and potential of applications. In this project I will talk about the use of X radiation to extract the secrets of a protein involved in the replication and repair of DNA from all terrestrial organisms. Its name is Nuclear Antigen of Proliferating Cells or PCNA for short. This protein has no enzymatic activity but plays the role of a sliding clamp. Enzymes from different families and functions such as DNA Ligase or DNA Polymerase are attached to PCNA.

The reasons that amaze us about this protein was that when irradiating the PCNA of Thermococcus gammatolerans with X-rays, it did not suffer any of the usual damages observed in other proteins. We bioinformatically explore the three-dimensional structure of this PCNA and analyze it evolutionarily. In other words, we explore their possible changes over time. Now we have a set of secrets that we steal from these proteins about their resistance. We are amazed at the results because they have enormous application potential to modify molecules of other organisms that cannot resist radiation but are indispensable in our daily lives, especially microorganisms. This represents a contribution to the nascent field of Synthetic Astrobiology that is the combination of modified organisms in the laboratory with there applications to astrobiology. Our dream for the future is the usage of this information to modify important organisms so that they survive extraterrestrial conditions and help our travel trough the stars.

**FLOOR 12** 



### CONTRAST BETWEEN THE MAIN LIFE'S **ORIGINS ABIOGENIC** MODELS



Rodríguez-Torres, L. M., Quintero-Luis, R., Carreño-López, R. Centro de Investigaciones en Ciencias Microbiológicas, BUAP, México.

experiments.

517-531

Currently, theories have been developed that seek to explain the origin of life from abiogenesis, however there are so many theories that they can be grouped into models. The 'genetics-first' model has the most accepted theory: RNA world, this molecule has important properties but it is too labile. Besides, the 'metabolism-first' model has a theory that has acquired importance: Iron-sulfur world, the energy released from the metal sulfides was available for the synthesis of organic molecules and the formation of polymers. Finally, the 'co-evolution of the three components' model takes some elements of the theories presented previously to generate a more complete and solid explanation but it has only one theory, which is based on peptide/oligonucleotide interdependence in relation with oil slick. This review contrasts these models and suggests the need to test them by well-designed

Gargaud, M., Martin, H., López-García, P., Montmerle, T., & Pascal, R. (2009). Young Sun, Early Earth and the Origins of Life. (S. Dunlop, Trans.) París, Francia: Springer. Griffith, R. W. (2009). A Specific Scenario for the Origin of Life and the Genetic Code Based on Peptide/Oligonucleotide Interdependence. Origins of Life and Evolution of the Biosphere, XXXIX,

Lazcano, A. (2010). El origen y la evolución temprana de la vida. In UNESCO, G. A. Lemarchand, & G. Tancredi (Eds.), Astrobiología: del Big Bang a las civilizaciones.

Montevideo, Uruguay: UNESCO.

Martin, W., & Russell, M. J. (2003). On the origins of cells: a hypothesis for the evolutionary transitions from abiotic geochemistry to chemoautotrophic prokaryotes, and from prokaryotes to nucleated cells. Philosophical Transactions of the Royal Society B, CCCLVIII, 59-85.

The importance of laboratory practices on the origin of life at the uppermiddle level

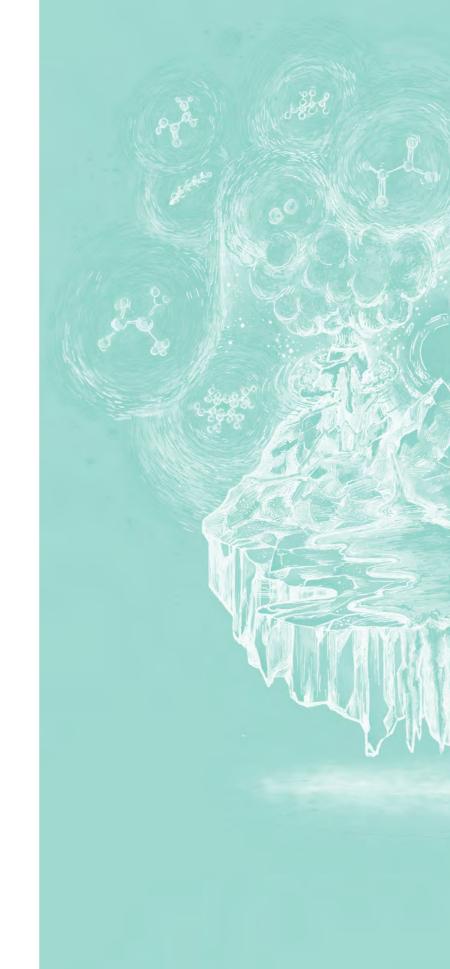
### THE IMPORTANCE OF LABORATORY PRACTICES ON THE ORIGIN OF LIFE AT THE UPPER-MIDDLE LEVEL



Tovilla Quesada Rubén de Jesús Instituto Politécnico Nacional The society in which the human being operates at present is a society characterized mainly by great scientific and technological advances and above all by the great dependence on technological devices, however, scientific illiteracy is just as broad, it is, therefore, from the schools, this scientific and technological knowledge of individuals at all educational levels must be worked on and struc-tured. In the school system corresponding to the upper secondary level in Mexico, there is a great diversity of subsystems which aim to satisfy the demand and coverage of the population in terms of access to education. Said subsystems approach biology in a similar way where the curriculum focuses mainly on systems theory with an emphasis on ecology and the environment, on average they take 3 to 5 hours/week/month, which leaves little time to deal with other topics of biological importance, this research aims to analyze how the topic of the Origin of Life is addressed as well as the laboratory practices that are implemented to reinforce the topic. Seven higher-level educational subsystems contained in four public educational systems were analyzed, which were:

Public Educational System	Subsystem
National Autonomous Uni. of Mexico (UNAM)	College of Science and Humanities (CCH)
	National High School (ENP)
National Polytechnic Institute (IPN)	Centers for Scientific and Technological
	Studies (CECyTs)
Undersecretary of Higher Middle Edu. (SEMS)	National College of Technical Professional
	Education (CONALEP)
Institute of Higher Middle Education (IEMS)	College of Baccalaureates (COLBACH)
	Directorate-General for Baccalaureates (DGE
	Mexico City High Schools

The results show that the topic of the Origin of life is approached in a theoretical way and some subsystems it does not exist in its study plan or it is approached superficially emphasizing the theoretical part where students are invited to de-bate the different theories of the origin of life integrating the social and cultural aspects of the historical moment when the theory was raised, in some subsystems a timeline of theories of the origin of life is requested. Regarding the labora-tory sessions, it was found that the experiment to be carried out is that of Francis-co Redi and the meat in jars. However, the results obtained by the students con-fuse them depending on the type of meat they use. Another practice found was the formation of coacervates with grenetine which interested the students, how-ever, the use of microscopes is necessary and some subsystems lack this re-source.



# PARTICIPANT DIRECTORY

S.No	Last Name	First Name	Affiliations	Country	Email Address	S.No	Last Name	First Name	Affiliations	Country	Email Address
1.	Abbas	Manzar	Radboud University	Netherlands	m.abbas@science.ru.nl	51.	Becker	Sidney	University of Cambridge	United Kingdom	sb2391@cam.ac.uk
2.	Abil	Zhanar	Delft University of Technology	Netherlands	z.abil@tudelft.nl	52.	Begka	Christina	Monash university	Australia	christina.begka@monash.edu
3.	Abrishamkar	Afshin	ESPCI Paris, CNRS, PSL Research University	France	afshin.abrishamkar@espci.fr	53.	Behmard	Aida	California Institution of Technology	United States	abehmard@caltech.edu
4.	Agudo-Canalejo	Jaime	Max Planck Institute for Dynamics and Self-Organization	Germany	jaime.agudo@ds.mpg.de	54.	Bektas	Onurcan	Middle East Technical University, Bilkent University	Turkey	onurcanbkts@gmail.com
5.	Aikkila	Paula	Ludwig Maximilian University, Munich	Germany	p.aikkila@physik.uni-muenchen.de	55.	Bell	Elizabeth	University of California, Los Angeles	United States	ebell21@ucla.edu
6.	Airapetian	Vladimir	NASA GSFC and American University, DC	United States	vladimir.airapetian@nasa.gov	56.	Bellini	Tommaso	Università degli studi di Milano	Italy	tommaso.bellini@unimi.it
7.	Aithal	Anuraag	Indian Institute of Science Education and Research (IISER), Pune	India	gv.anuraag@students.iiserpune.ac.in	57.	Bergmann	Alexander	Technical University of Munich	Germany	alexander.bergmann@tum.de
8.	Akbari	Zahra	University of Tehran	Iran, Islamic Republic of	f zakbary@ut.ac.ir	58.	Berry	Drew	Walter and Eliza Hall Institute of Medical Research	Australia	berry@wehi.edu.au
9.	Albayrak	Esra	Marmara University	Turkey	esraalbayrak@marun.edu.tr	59.	Bertram	Lauren	University of Edinburgh	United Kingdom	laurenbertram@btinternet.com
10.	Alić	Jasna	Ruđer Bošković Institute	Croatia	jasna.alic@irb.hr	60.	Bhoge	Bapurao	Indian Institute of Science Education and Research (IISER), Bhopal	India	bapub@iiserb.ac.in
11.	Alim	Karen	Technical University of Munich	Germany	k.alim@tum.de	61.	Biondi	Elisa	Foundation for Applied Molecular Evolution	United States	ebiondi@ffame.org
12.	Alımlı	Nimet	Middle East Technical University, Institute of Marine Sciences	Turkey	nimetalimli@gmail.com	62.	Biswas	Sudipta	National Institute of Science Education and Research	India	sudipta.biswas@niser.ac.in
13.	Alishayeva	Saudat	Bahcesehir University	Turkey	saudat.alishayeva@bahcesehir.edu.tr	63.	Bitaraf	Amirreza	Tarbiat Modares University	Iran, Islamic Republic of	a.bitaraf91@gmail.com
14.	Altaner	Bernhard	Technical University of Munich	Germany	bernhard.altaner@tum.de	64.	Black	Roy A.	University of Washington	United States	royblack@comcast.net
15.	Alvarez	Claudia	Georgia Institute of Technology	Mexico	ccarreno6@gatech.edu	65.	Blackmond	Donna G.	Scripps Research	United States	blackmon@scripps.edu
16.	Alves	Júlia	Valongo Observatory (Federal University of Rio de Janeiro)	Brazil	julia17@astro.ufrj.br	66.	Blokhuis	Alex	University of Groningen	Netherlands	a.w.p.blokhuis@rug.nl
17.	Ameta	Sandeep	Simons Centre, National Centre for Biological Sciences, Bangalore	India	sandeepameta@ncbs.res.in	67.	Boekhoven	Job	Technical University of Munich	Germany	job.boekhoven@tum.de
18.	Andrade	Jeffery	Max Planck Institute for Dynamics and Self-Organization	United States	jandrade01@alumni.harvard.edu	68.	Boeynaems	Steven	Stanford University	United States	sboeynae@stanford.edu
19.	Angerpointner	Severin	Ludwig Maximilian University, Munich	Germany	s.angerpointner@physik.lmu.de	69.	Bogucki	Ryan	University of Illinois at Urbana-Champaign	United States	bogucki2@illinois.edu
20.	Antonio Durán	Yadira Zulema	Universidad Autónoma de Nuevo León	Mexico	yadiraaduran@gmail.com	70.	Bomba	Radoslaw	ETH Zurich	Switzerland	radoslaw.bomba@phys.chem.ethz.ch
21.	Antony	Claude	CNRS Emeritus	France	claudeantony78@gmail.com	71.	Bonfio	Claudia	MRC Laboratory of Molecular Biology	United Kingdom	bonfio@mrc-Imb.cam.ac.uk
22.	Aonuma	Keito	Tokyo University of Science	Japan	aonuma-keito168@g.ecc.u-tokyo.ac.jp	72.	Bose	Rudrarup	Max Planck Institute for Molecular Cell Biology and Genetics	Germany	bose@mpi-cbg.de
23.	Arjomand-Fard	Hanieh	Ludwig Maximilian University, Munich	Germany	hanieh.arjomand-fard@lrz.uni-muenchen.de	73.	Bradley	James	Queen Mary University of London	United Kingdom	jbradley.earth@gmail.com
24.	Armas Vazquez	Marion Zulema	Universidad Nacional Autónoma de México, Instituto de Ciencias Nucleares	Mexico	mzulem@correo.nucleares.unam.mx	74.	Brau	Joanna	Ludwig Maximilian University, Munich	Germany	joanna.brau@min.uni-muenchen.de
25.	Arranz-Gibert	Pol	Yale University	USA	pol.arranzgibert@yale.edu	75.	Braun	Dieter	Ludwig Maximilians University, Munich	Germany	dieter.braun@lmu.de
26.	Ashkenasy	Gonen	Ben-Gurion University of the Negev	Israel	gonenash@bgu.ac.il	76.	Bredehöft	Jan Hendrik	University of Bremen	Germany	jhbredehoeft@uni-bremen.de
27.	Athavale	Soumitra	California Institute of Technology	United States	athavale@caltech.edu	77.	Briggs	Kyle	University of Ottawa	Canada	kbrig035@uottawa.ca
28.	Atri	Dimitra	New York University, Abu Dhabi	United Arab Emirates	atri@nyu.edu	78.	Brindley	John	University of Leeds	United Kingdom	john32brindley@gmail.com
29.	Attwater	James	MRC Laboratory of Molecular Biology	United Kingdom	jswa@mrc-Imb.cam.ac.uk	79.	Brouillet	Nathalie	Laboratoire d'astrophysique de Bordeaux	France	nathalie.brouillet@u-bordeaux.fr
30.	Augustine	Anusree	ESPCI Paris	France	anusree.augustine@espci.fr	80.	Broz	Jakub	Space Generation Advisory Council	Czechia	jakub.broz@spacegeneration.org
31.	Ayala	Dagoberto	Universidad Nacional de Ingenieria	Peru	dayalap@uni.pe	81.	Bunel	Louis	École Normale Supérieure	France	louis.bunel@ens.psl.eu
32.	В	Siddlingeshwar	M S Ramaiah Institute of Technology	India	sidduphysics@gmail.com	82.	Burkert	Andreas	Ludwig Maximilian University, Munich	Germany	burkert@usm.lmu.de
33.	Baba	Akiko	Tohoku University	Japan	baba@bio.phys.tohoku.ac.jp	83.	Cabañas	Noel	Universidad Nacional Autónoma de México	Mexico	ncabanas@iibiomedicas.unam.mx
34.	Bajaj	Lakshya	Harvard Medical School	United States	lakshya_bajaj@hms.harvard.edu	84.	Caimi	Federico	Università degli studi di Milano	Italy	federico.caimi@unimi.it
35.	Balcı	Uğur	Middle East Technical University	Turkey	ubalci@metu.edu.tr	85.	Caliskanoglu	Zeynep	Ludwig Maximilian University, Munich	Germany	caliskanoglu@itu.edu.tr
36.	Balga	Elizabeth	The Boeing Company	United States	elizabethbalga@gmail.com	86.	Calle	Demetrio	Consejería Educación y Ciencia Junta de Andalucía	Spain	cienciaydocencia@hotmail.com
37.	Ball	Rowena	Australian National University	Australia	rowena.ball@anu.edu.au	87.	Carlino	Roberto	NASA Ames	United States	rob.carlino47@gmail.com
38.	Bando	Hina	Kyoto University	Japan	trkhina@gmail.com	88.	Caro	Tristan	Colorado University Boulder	United States	tristan.caro@colorado.edu
39.	Banerjee	Abesh	Arizona State University	United States	abaner30@asu.edu	89.	Caselli	Paola	Max Planck Institute for Extraterrestrial Physics	Germany	caselli@mpe.mpg.de
40.	Bannan	Abdullah	Harvard College	United States	ambannan@college.harvard.edu	90.	Castillo	Gina	Universidad Nacional de Colombia	Colombia	gpcastillor@unal.edu.co
41.	Вао	Bin	Boston Children's Hospital	United States	bin.bao@childrens.harvard.edu	91.	Cavalcante	Larissa	KTH Royal Institute of Technology	Sweden	lalc@kth.se
42.	Barral	Antonio	Centro Nacional de Investigaciones Cardiovasculares	Spain	abarral@cnic.es	92.	Cerezuela Mora	Javier		Spain	javicmora@gmail.com
43.	Bartlett	Stuart	California Institute of Technology	United States	sjbart@caltech.edu	93.	Chakraborty	Nayan	National Centre for Biological Sciences, TIFR	India	nayanc@ncbs.res.in
44.	Bartolucci	Giacomo	Max Planck Institute for the Physics of Complex Systems	Germany	bartolucci@pks.mpg.de	94.	Chandrakanth Shett	y Sunidhi	Max Planck Institute of Colloids and Interfaces	Germany	sunidhi.shetty@mpikg.mpg.de
45.	Basak	Shibaji	University of Alberta	Canada	shibaji@ualberta.ca	95.	Chandrasekharan	Gaayatri	Indian Institute of Science Education and Research (IISER), Thiruvananthapu	ıra <b>m</b> dia	irtayaagaayatri@gmail.com
46.	Baudelaire	Fox	Brandeis University	United States	baudelaire@brandeis.edu	96.	Chandru	Kuhan	University of Chemistry and Technology, Prague	Czechia	nahuk82@gmail.com
47.	Bauer	Magnus	Ludwig Maximilian University, Munich	Germany	magnus.bauer@lmu.de	97.	Charron	Martin	University of Ottawa	Canada	mchar243@gmail.com
48.	Baum	Eric	TruthSift Inc.	United States	ebaum@fastmail.fm	98.	Chatterjee	Subhasish	Barnard College of Columbia University	United States	sc2952@columbia.edu
49.	Baum	Stefi	University of Manitoba	Canada	stefi.baum@umanitoba.ca	99.	Chávez	Fernanda	Benemérita Universidad Autónoma de Puebla	Mexico	fernanda-0298@hotmail.com
50.	Beatty	Meagan	University of Groningen	Netherlands	m.a.beatty@rug.nl	100.	Chelban	Andreea-Oana	Ludwig Maximilian University, Munich	Germany	oana.chelban@gmail.com 173

S.No	Last Name	First Name	Affiliations	Country	Email Address	S.No	Last Name	First Name	Affiliations
101.	Chen	Siyu	Max Planck Institute for Dynamics and Self-Organization	Germany	siyu.chen@ds.mpg.de	151.	Dogterom	Marileen	Delft University of Technology
102.	Chen	Xiaoyao	Technical University of Munich	Germany	xiaoyao.chen@tum.de	152.	Domingos de Souza Filho	Celso	CIIMAR
103.	Cheng	Gong	Harvard University	United States	chenggong24@gmail.com	153.	Domínguez	José	Universidad Autónoma de Chiapas
104.	Chiappino-Pepe	Anush	Harvard Medical School	United States	anush_chiappinopepe@hms.harvard.edu	154.	Donau	Carsten	Technical University of Munich
105.	Chiba	Toshikaze	Tohoku University	Japan	chiba@bio.phys.tohoku.ac.jp	155.	Doppleb	Olivia	University of Stuttgart
106.	Choi	Saehyun	Pennsylvania State University	United States	szc227@psu.edu	156.	Draper	Thomas	Harvard University
107.	Cimino	Roberto	LNF-INFN	Italy	roberto.cimino@Inf.infn.it	157.	Duda	Jan-Peter	University of Göttingen & Göttingen Academy of Sciences
108.	Civril	Filiz	CRC 235 - Emergence of Life	Germany	filiz.civril@lmu.de	158.	Duzdevich	Daniel	ННМІ
109.	Cleaves	Henderson	Tokyo Institute of Technology	Japan	henderson.cleaves@gmail.com	159.	Eiban	Ines	Ludwig Maximilian University, Munich
110.	Cleij	Céline	Delft University of Technology	Netherlands	celinecleij@hotmail.com	160.	Eicholt	Lars	Lund University
111.	Clifton	Bryce	Georgia Institute of Technology	United States	bclifton3@gatech.edu	161.	Eiler	John	California Institute of Technology
112.	Cohen	Trevor	University of California, Santa Barbara	United States	tecohen@ucsb.edu	162.	Eisenreich	Wolfgang	Technical University of Munich
113.	Cohen	Zachary	University of Washington	United States	zrcohen2@uw.edu	163.	Endesfelder	Ulrike	Carnegie Mellon University
114.	Colizzi	Enrico Sandro	Leiden University; Origins Center	Netherlands	e.s.colizzi@math.leidenuniv.nl	164.	Enright	Allison	University of New Brunswick
115.	Colomer	Ignacio	Universidad Autonoma de Madrid	Spain	ignacio.colomer@uam.es	165.	Erkamp	Nadia	Radboud University
116.	Colon-Santos	Stephanie	University of Wisconsin-Madison	United States	colonsantos@wisc.edu	166.	Escobar Rivera	Rolando	Instituto Tecnologico de Tuxtla Gutierrez
117.	Colville	Ben	University College London	United Kingdom	zccabwf@ucl.ac.uk	167.	Espada	Rocio	ESPCI Paris - PSL
118.	Corzo	Julian	Universidad Nacional de Colombia	Colombia	jacorzoa@unal.edu.co	168.	Espinosa Trujillo	Olivia	University of Sheffield
119.	Coskun	Ömer Kürsat	Ludwig Maximilian University, Munich	Germany	o.coskun@campus.lmu.de	169.	Esti	Mertcan	Middle East Technical University, Institute of Marine Scien
120.	Costa de Almeida	Ellen	Valongo Observatory (Federal University of Rio de Janeiro)	Brazil	dealmeida.ec@gmail.com	170.	Ettikkan	Nandha Kumar	University of Hyderabad
121.	Crisp	Antony	Ludwig Maximilian University, Munich	Germany	antony.crisp@cup.lmu.de	171.	Fairchild	Jasper	University College London
122.	Cross	Edward	King's College, London	United Kingdom	edward.cross@kcl.ac.uk	172.	Fakhretaha Aval	Sara	Georgia Institute of Technology
123.	Cruz Simbron	Romulo Leoncio	Universidad Nacional de Ingenieria/Sociedad Científica de Astrobiologia del P		romulo.cruz.s@uni.pe	173.	Fan	Yaxun	Chinese Academy of Sciences, Institute of Chemistry
124.	Cvjetan	Nemanja	ETH Zurich	Switzerland	nemanja.cvjetan@mat.ethz.ch	174.	Fares	Hadi	Pennsylvania State University/ NASA Postdoctoral Program
125.	Dağ	Berfin		Turkey	berfinddag@gmail.com	175.	Farfán-Ramos	Daniel	Universidad Nacional Autónoma de México
126.	Dagar	Shikha	Indian Institute of Science Education and Research (IISER), Pune	India	shikha.daqar@students.iiserpune.ac.in	176.	Faria	Vitor	University of Basel
127.	Dai	Kun	Technical University of Munich	Germany	daikun199@gmail.com	177.	Farias	Raquel	Federal University of Rio de Janeiro
128.	Daines	Elena	Radboud University	Netherlands	elenadaines@gmail.com	178.	Fathian	Maryam	University of Zanjan
129.	Danelon	Christophe	Delft University of Technology	Netherlands	c.j.a.danelon@tudelft.nl	179.	Feldmann	Jonas	Ludwig Maximilian University, Munich
130.	Dannenmann	Marie	Freie Universität Berlin	Germany	marie@dannenmann.net	180.	Feshangsaz	Niloofar	Utrecht University Medical Center
131.	Das	Souradeep	Indian Institute of Science Education and Research (IISER), Pune	India	das.souradeep@students.iiserpune.ac.in	181.	Filippidi	Emmanouela	Max Planck Institute for Molecular Cell Biology and Geneti
132.	DasGupta	Saurja	Harvard Medical School (Massachusetts General Hospital	United States	dasgupta@molbio.mgh.harvard.edu	182.	Flamm	Christoph	University of Vienna
133.	de Almeida Filho	Enezio E.		Brazil	neddybr@outlook.com	183.	Flommersfeld	Johannes	Ludwig Maximilian University, Munich
134.	de Boer	Carl	University of British Columbia	Canada	carl.deboer@ubc.ca	184.	Floroni	Alexander	Ludwig Maximilian University, Munich
135.	de la Escosura	Andres	Universidad Autonoma de Madrid	Spain	andres.delaescosura@uam.es	185.	Focil-Espinosa	Carlos	Universidad Autónoma de Yucatán
136.	De Micco	Claudia	Università degli studi di Bari Aldo Moro	Italy	c.demicco@studenti.uniba.it	186.	Forget	Selene	École Normale Supérieure
137.	de Vries	Mattanjah	University of California, Santa Barbara	United States	devries@ucsb.edu	187.	Forsythe	Jay	College of Charleston
138.	de Vries	Mattanjah	University of California, Santa Barbara	USA	devries@ucsb.edu	188.	Fracassi	Alessandro	ETH Zurich
139.	DeHaven	Baillie	Northwestern University	United States	baillie.dehaven@northwestern.edu	189.	Fraccia	Tommaso	ESPCI Paris
		Luis	Cinvestar Irapuato						Northwestern University
140.	Delaye		Universidad Nacional de Colombia	Mexico Colombia	luis.delaye@cinvestav.mx	190.	Fraser Frenkel-Pinter	Craig	
141.	Delgado Sarmiento	Isaac Augusto			iadelgados@unal.edu.co	191.		Moran	Georgia Institute of Technology
142.	Deshpande	Kshitij	Indian Institute of Science Education and Research (IISER), Pune	India	kshitij.ganeshdeshpande@students.iiserpune.ac.in	192.	Frey	Erwin	Ludwig Maximilians University, Munich
143.	Dev	Dharm	Ben-Gurion University of the Negev	Israel	dharmiitr@gmail.com	193.	Fuks	Elina	Ludwig Maximilian University, Munich
144.	Devanapally	Sindhuja	Columbia University	United States	sd3387@cumc.columbia.edu	194.	Furubayashi	Taro	CNRS
145.	Di Daniel	Simone		Italy	simone_didaniel@live.it	195.	Gagnon	Jean-Sebastien	Norwich University
146.	Di Meo	Thibault	ESPCI Paris	France	thibault.di-meo@espci.fr	196.	Gaizauskaite	Aukse	
147.	Diederich	Philippe	Helmholtz Zentrum München	Germany		197.	Gallardo Ramirez	Abel	Universidad Nacional Autónoma de México, Instituto de As
148.	Dingwell	Donald	Ludwig Maximilian University, Munich	Germany	dingwell@lmu.de	198.	Garcia	Amanda	University of Arizona
149.	Dirscherl	Christina Felicitas	Ludwig Maximilian University, Munich	Germany	christina.dirscherl@physik.uni-muenchen.de	199.	Garcia	Gabriel	West Virginia University
150.	Djutanta	Franky	Arizona State University	United States	fdjutant@asu.edu	200.	García Milian	Dulce Alejandra	Universidad Michoacana de San Nicolás de Hidalgo

	104	~
		Sec. 2
11 (1)		

Country

Email Address

	Netherlands	M.Dogterom@tudelft.nl
	Portugal	bio.celso.domingos@gmail.com
	Mexico	albertdomlop@gmail.com
	Germany	carsten.donau@tum.de
	Germany	olivia.doppleb@oc.uni-stuttgart.de
	United States	tdraper@fas.harvard.edu
nces and Humanities	Germany	jduda@gwdg.de
	United States	duzdevich@molbio.mgh.harvard.edu
	Germany	ines.eiban@lmu.de
	Sweden	la6340ei-s@student.lu.se
	United States	eiler@gps.caltech.edu
	Germany	wolfgang.eisenreich@ch.tum.de
	United States	uendesfelder@cmu.edu
	Canada	allison.enright@unb.ca
	Netherlands	nadia.erkamp@hotmail.com
	Mexico	escobar_bioquim99@hotmail.com
	France	rocio.espada@espci.fr
	United Kingdom	oespinosatrujillo1@sheffield.ac.uk
Sciences	Turkey	mertcanesti@gmail.com
	India	nandhakumar1997@gmail.com
	United Kingdom	uccajfa@ucl.ac.uk
	United States	sfa@gatech.edu
	China	yxfan@iccas.ac.cn
rogram	United States	hpf5094@psu.edu
	Mexico	d.farfan2003@gmail.com
	Switzerland	vitor.gouveiafaria@unibas.ch
	Brazil	raquelgomesgf@gmail.com
	Iran, Islamic Republic of	mfathian1987@gmail.com
	Germany	jonas.feldmann@cup.lmu.de
	Netherlands	n.feshangsaz@gmail.com
enetics	Germany	filippidi@mpi-cbg.de
	Austria	xtof@tbi.univie.ac.at
	Germany	johannes.flommersfeld@gmx.de
	Germany	alexander.floroni@campus.lmu.de
	Mexico	carlosfocil09@gmail.com
	France	selene.forget@ens.fr
	United States	forsythejg@cofc.edu
	Switzerland	afracassi@org.chem.ethz.ch
	France	tommaso.fraccia@espci.fr
	United States	craig.fraser@northwestern.edu
	United States	moranfp@gatech.edu
	Germany	frey@lmu.de
	Germany	elfuch@cup.uni-muenchen.de
	France	furubayashi.taro@gmail.com
	United States	jgagnon6@norwich.edu
	Lithuania	auksegaizauskaite@gmail.com
de Astronomia	Mexico	agallardo@astro.unam.mx
	United States	akgarcia@email.arizona.edu
	United States	patxos@yahoo.com
	Mexico	dulce_agm@outlook.com

S.No	Last Name	First Name	Affiliations	Country	Email Address	S.No	Last Name	First Name	Affiliations
201.	Gartner	Florian	Ludwig Maximilian University, Munich	Germany	f.gartner@physik.uni-muenchen.de	251.	Hlouchová	Klára	Charles University
202.	Gault	Stewart	University of Edinburgh	United Kingdom	s.a.gault@sms.ed.ac.uk	252.	Hoeijmakers	Jens	University of Bern
203.	Gaynor	Josselyn	IB	Mexico	josselyngaynor0@gmail.com	253.	Hogan	Michael	Independent Hospice Chaplain
204.	Gebauer	Stefanie	DLR - German Aerospace Center	Germany	stefanie.gebauer@dlr.de	254.	Holliger	Phil	MRC Laboratory of Molecular Biology
205.	Geisberger	Thomas	Technical University of Munich	Germany	thomas.geisberger@tum.de	255.	Honegger	Philipp	University of Vienna, Institute of Computational Biologica
206.	Gerland	Ulrich	Technical University of Munich	Germany	gerland@tum.de	256.	Honsa	Monique	Ludwig Maximilian University, Munich/ Institute for Molec
207.	Gezer	Eftal	Gebze Technical University	Turkey	egezer@gtu.edu.tr	257.	Huber	Laura	Ludwig Maximilian University, Munich
208.	Ghirlanda	Giovanna	Arizona State University	United States	gghirlanda@asu.edu	258.	Huber	Claudia	Technical University of Munich
209.	Ghosh	Souvik	Indian Institute of Science Education and Research (IISER), Kolkata	India	ghoshsouvik2197@gmail.com	259.	Huck	Wilhelm	Radboud University
210.	Ghosh	Rikhia	Max Planck Institute of Colloids and Interfaces	Germany	rikhia.ghosh@mpikg.mpg.de	260.	Hud	Nicholas V.	Georgia Institute of Technology
211.	Ghosh	Chandranath		India	chandranath244@gmail.com	261.	Hüttl	Lucas	Ludwig Maximilian University, Munich
212.	Giagnisi	Eleftheria		Germany	ritagiagnisi@gmail.com	262.	Hyodo	Ayumu	University of Tokyo
213.	Gianni	Edoardo	MRC Laboratory of Molecular Biology	United Kingdom	egianni@mrc-Imb.cam.ac.uk	263.	Hysi	Martina	Ludwig Maximilians University, Munich
214.	Gil Gutierrez	Nicolas		Colombia	nicolasgilgutierrez@hotmail.com	264.	Ianeselli	Alan	Ludwig Maximilian University, Munich
215.	Goeppel	Tobias	Technical University of Munich	Germany	goeppel.tobias@gmail.com	265.	Ibáñez-Costa	Alejandro	Maimónides Institute for Biomedical Research at Córdob
216.	Goh	Kahmin	University of Cologne	Germany	kmgoh1995@gmail.com	266.	Ichihashi	Norikazu	University of Tokyo
217.	Golden	Barbara	Purdue University	United States	barbgolden@purdue.edu	267.	Ichimura	Ryota	Tohoku University
218.	Gonzalez	Laura Elena	Universidad Autónoma de Nuevo León	Mexico	lau.ghdz91@gmail.com	268.	Ingalls	Miquela	Pennsylvania State University
219.	González	Sarah	Astrobiology Institute of Colombia	Colombia	sarah.gh988@gmail.com	269.	Ishida	Satoshi	University of Tokyo
220.	Green	Nicholas	MRC Laboratory of Molecular Biology	United Kingdom	ngreen@mrc-Imb.cam.ac.uk	270.	Islam	Saidul	University College London
221.	Greenwald	Jason	ETH Zurich	Switzerland	gjason@ethz.ch	271.	Isnard	Robin	ISIS
222.	Gruenberg	Raik	KAUST	Saudi Arabia	raik.gruenberg@gmail.com	272.	Izuhara	Yuichi	University of Tokyo
223.	Guerrero Cruz	Ariadna Alexandra	CDMX	Mexico	ari.guerrero07@gmail.com	273.	Jacobson	Sofia	University of Arizona
224.	Guillemin	Jean-Claude	ISCR-UMR CNRS 6226	France	jean-claude.guillemin@ensc-rennes.fr	274.	Jain	Sanjay	University of Delhi
225.	Guillén Soto	Rosa Guadalupe	Universidad de Guadalajara	Mexico	luguillens@gmail.com	275.	Jäkel	Anna	Technical University of Munich
226.	Guillou	Herve	Université Grenoble Alpes and CNRS	France	herve.guillou@neel.cnrs.fr	276.	Jandu	Harkabir	
227.	Güllülü	Ömer	University Hospital Frankfurt	Germany	oemer.guelluelue@kgu.de	277.	Janicki	Mikołaj	Wrocław University of Science and Technology
228.	Gunes	Yagmur	Istanbul Technical University	Turkey	gunesya@itu.edu.tr	278.	Janzen	Evan	University of California, Santa Barbara
229.	Guo	Wei	University of Hong Kong	China	wguohku@connect.hku.hk	279.	Jäschke	Andres	Heidelberg University
230.	Gupta	Kashish	Govt. College for Girls, Ludhiana, Panjab University	India	kashish.nath16@gmail.com	280.	Jasso	Reguinaldo	Birmex
231.	Guzmán	Isaac	Universidad Nacional Autónoma de México	Mexico	isaac-2702@hotmail.com	281.	Jens	Peter	AND BioPharma
232.	Haas	Maren	Ludwig Maximilian University, Munich	Germany	maren.haas@cup.uni-muenchen.de	282.	Jhandai	Prince	Indian Institute of Science Education and Research (IISEF
233.	Haesner	Melanie		Germany	m.haesner@tum.de	283.	Jia	Tony Z	Earth-Life Science Institute
234.	Haggmark	Michael	University of California, Santa Barbara	USA	haggmark@ucsb.edu	284.	Jia	Haiyang	Max Planck Institute of Biochemistry
235.	Halpern	Aaron	University College London	United Kingdom	zcqshal@ucl.ac.uk	285.	Jimenez del Rio	Luis Alberto	
236.	Hara	Ellie	University of Colorado	United States	ellie.hara@colorado.edu	286.	Jimeno Montiel	Edgar Fredy	SECTEI
237.	Harth-Kitzerow	Johannes	Max Planck Institute for Astrophysics	Germany	jharthki@mpa-garching.mpg.de	287.	Jin	Yulong	Chinese Academy of Sciences, Institute of Chemistry
238.	Hashimoto	Hinata	Tokyo Institute of Technology	Japan	hashimoto.h.aj@m.titech.ac.jp	288.	Joseph	Anton	DWI - Leibniz Institute for Interactive Materials e.V.
239.	Hayafune	Masahiro	Co-Lab.	Japan	co.lab.hayafune@gmail.com	289.	Joshi	Priyanka	University of California, Berkeley
240.	Heckl	Wolfgang	Deutsches Museum	Germany	generaldirektor@deutsches-museum.de	290.	Jung	Kirsten	Ludwig Maximilian University, Munich
241.	Hendrikse	Simone	University of Melbourne	Australia	shendriksela@unimelb.edu.au	291.	Kacar	Betul	University of Arizona
242.	Henkes	Gregory	Stony Brook University	United States	gregory.henkes@stonybrook.edu	292.	Kaila	Ville	Stockholm University
243.	Hennig	Susanne	Ludwig Maximilian University, Munich	Germany	hennig@cens.de	293.	Kamimura	Atsushi	University of Tokyo
244.	Henning	Thomas	Max Planck Institute for Astronomy	Germany	henning@mpia.de	294.	Kantarci	Ilayda	Ludwig Maximilian University, Munich
245.	Herbst	Konstantin	Christian-Albrechts-Universität zu Kiel	Germany	herbst@physik.uni-kiel.de	295.	Kapdan	Cansu	Ludwig Maximilian University, Munich
246.	Hernandez	Mario	IPN	Mexico	malberthortiz@gmail.com	296.	Kardeş	Gülce	University of Leipzig
247.	Hernandez	Erick	Universidad Autónoma de Nuevo León	Mexico	erick_419@live.com.mx	297.	Kardile	Vaishnavi	Indian Institute of Science Education and Research (IISEF
248.	Hernández	José G.		Germany	gregoriojose10@gmail.com	298.	Karne	Anagha	MIT World Peace University
249.	Hertkorn	Norbert	Helmholtz Zentrum München	Germany	hertkorn@helmholtz-muenchen.de	299.	Karr	Leonie	Ludwig Maximilian University, Munich
250.	Hirakawa	Yuta	Tohoku university	Japan	yuta.hirakawa.s2@dc.tohoku.ac.jp	300.	Kastsiusheuskaya	Aryna	
			-	-	2 N			-	

	NS:		-
		1	-
10 10			
2			

Country

Email Address

Czech Republic	klara.hlouchova@natur.cuni.cz
Switzerland	jens.hoeijmakers@space.unibe.ch
United States	m9238hogan@hotmail.com
United Kingdom	ph1@mrc-Imb.cam.ac.uk
Austria	philipp.honegger@univie.ac.at
Japan	monique.honsa@physik.uni-muenchen.de
Germany	lahuch@cup.uni-muenchen.de
Germany	claudia.huber@mytum.de
Netherlands	w.huck@science.ru.nl
United States	hud@chemistry.gatech.edu
Germany	lucas@huettl.de
Japan	531aym@g.ecc.u-tokyo.ac.jp
Germany	m.hysi@physik.uni-muenchen.de
Germany	a.ianeselli@physik.uni-muenchen.de
Spain	alejandro.ibanez@imibic.org
Japan	ichihashi@bio.c.u-tokyo.ac.jp
Japan	ryota.ichimura.r2@dc.tohoku.ac.jp
United States	ingalls@psu.edu
Japan	isatoshi@chem.s.u-tokyo.ac.jp
United Kingdom	saidul.islam@ucl.ac.uk
France	risnard@unistra.fr
Japan	izuhara-yuichi190@g.ecc.u-tokyo.ac.jp
United States	jacobsons1@email.arizona.edu
India	jain_physics@yahoo.co.in
Germany	anna.jaekel@tum.de
India	harkabir@gmail.com
India Poland	harkabir@gmail.com mikolaj.janicki@pwr.edu.pl
Poland	mikolaj.janicki@pwr.edu.pl
Poland United States	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu
Poland United States Germany	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de
Poland United States Germany Mexico	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com
Poland United States Germany Mexico Belgium	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl
Poland United States Germany Mexico Belgium India	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com
Poland United States Germany Mexico Belgium India Japan	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com tzjia@elsi.jp
Poland United States Germany Mexico Belgium India Japan Germany	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com tzjia@elsi.jp jia@biochem.mpg.de
Poland United States Germany Mexico Belgium India Japan Germany Mexico	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com tzjia@elsi.jp jia@biochem.mpg.de luis.jdelrio@alumnos.udg.mx
Poland United States Germany Mexico Belgium India Japan Germany Mexico Mexico	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com tzjia@elsi.jp jia@biochem.mpg.de luis.jdelrio@alumnos.udg.mx edgar.jimeno@educacion.cdmx.gob.mx jinyulong2008@gmail.com joseph@dwi.rwth-aachen.de
Poland United States Germany Mexico Belgium India Japan Germany Mexico China Germany United States	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com tzjia@elsi.jp jia@biochem.mpg.de luis.jdelrio@alumnos.udg.mx edgar.jimeno@educacion.cdmx.gob.mx jinyulong2008@gmail.com joseph@dwi.rwth-aachen.de prijoshi@berkeley.edu
Poland United States Germany Mexico Belgium India Japan Germany Mexico China Germany United States Germany	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com tzjia@elsi.jp jia@biochem.mpg.de luis.jdelrio@alumnos.udg.mx edgar.jimeno@educacion.cdmx.gob.mx jinyulong2008@gmail.com joseph@dwi.rwth-aachen.de prijoshi@berkeley.edu jung@lmu.de
Poland United States Germany Mexico Belgium India Japan Germany Mexico China Germany United States Germany United States	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com tzjia@elsi.jp jia@biochem.mpg.de luis.jdelrio@alumnos.udg.mx edgar.jimeno@educacion.cdmx.gob.mx jinyulong2008@gmail.com joseph@dwi.rwth-aachen.de prijoshi@berkeley.edu jung@lmu.de
Poland United States Germany Mexico Belgium India Japan Germany Mexico China Germany United States Germany United States Sweden	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com tzjia@elsi.jp jia@biochem.mpg.de luis.jdelrio@alumnos.udg.mx edgar.jimeno@educacion.cdmx.gob.mx jinyulong2008@gmail.com joseph@dwi.rwth-aachen.de prijoshi@berkeley.edu jung@lmu.de betul@arizona.edu ville.kaila@dbb.su.se
Poland United States Germany Mexico Belgium India Japan Germany Mexico China Germany United States Germany United States Sweden Japan	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com tzjia@elsi.jp jia@biochem.mpg.de luis.jdelrio@alumnos.udg.mx edgar.jimeno@educacion.cdmx.gob.mx jinyulong2008@gmail.com joseph@dwi.rwth-aachen.de prijoshi@berkeley.edu jung@lmu.de betul@arizona.edu ville.kaila@dbb.su.se kamimura@complex.c.u-tokyo.ac.jp
Poland United States Germany Mexico Belgium India Japan Germany Mexico China Germany United States Germany United States Sweden Japan Germany	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com tzjia@elsi.jp jia@biochem.mpg.de luis.jdelrio@alumnos.udg.mx edgar.jimeno@educacion.cdmx.gob.mx jinyulong2008@gmail.com joseph@dwi.rwth-aachen.de prijoshi@berkeley.edu jung@lmu.de betul@arizona.edu ville.kaila@dbb.su.se kamimura@complex.c.u-tokyo.ac.jp ilayda.kantarci@campus.lmu.de
Poland United States Germany Mexico Belgium India Japan Germany Mexico China Germany United States Germany United States Sweden Japan Germany	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com tzjia@elsi.jp jia@biochem.mpg.de luis.jdelrio@alumnos.udg.mx edgar.jimeno@educacion.cdmx.gob.mx jinyulong2008@gmail.com joseph@dwi.rwth-aachen.de prijoshi@berkeley.edu jung@lmu.de betul@arizona.edu ville.kaila@dbb.su.se kamimura@complex.c.u-tokyo.ac.jp ilayda.kantarci@campus.lmu.de cansukapdann@gmail.com
Poland United States Germany Mexico Belgium India Japan Germany Mexico China Germany United States Germany United States Sweden Japan Germany Germany	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com tzjia@elsi.jp jia@biochem.mpg.de luis.jdelrio@alumnos.udg.mx edgar.jimeno@educacion.cdmx.gob.mx jinyulong2008@gmail.com joseph@dwi.rwth-aachen.de prijoshi@berkeley.edu jung@lmu.de betul@arizona.edu ville.kaila@dbb.su.se kamimura@complex.c.u-tokyo.ac.jp ilayda.kantarci@campus.lmu.de cansukapdann@gmail.com
Poland United States Germany Mexico Belgium India Japan Germany Mexico China Germany United States Germany United States Sweden Japan Germany Germany	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com tzjia@elsi.jp jia@biochem.mpg.de luis.jdelrio@alumnos.udg.mx edgar.jimeno@educacion.cdmx.gob.mx jinyulong2008@gmail.com joseph@dwi.rwth-aachen.de prijoshi@berkeley.edu jung@lmu.de betul@arizona.edu ville.kaila@dbb.su.se kamimura@complex.c.u-tokyo.ac.jp ilayda.kantarci@campus.lmu.de cansukapdann@gmail.com gulcekardes@gmail.com
PolandUnited StatesGermanyMexicoBelgiumIndiaJapanGermanyMexicoChinaGermanyUnited StatesGermanyUnited StatesSwedenJapanGermanyGermanyIndiaGermanyBermanyIndiaIndia	mikolaj janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com tzjia@elsi.jp jia@biochem.mpg.de luis.jdelrio@alumnos.udg.mx edgar.jimeno@educacion.cdmx.gob.mx jinyulong2008@gmail.com joseph@dwi.rwth-aachen.de prijoshi@berkeley.edu jung@lmu.de betul@arizona.edu ville.kaila@dbb.su.se kamimura@complex.c.u-tokyo.ac.jp ilayda.kantarci@campus.lmu.de cansukapdann@gmail.com gulcekardes@gmail.com
Poland United States Germany Mexico Belgium India Japan Germany Mexico China Germany United States Germany United States Sweden Japan Germany Germany	mikolaj.janicki@pwr.edu.pl evanjanzen@ucsb.edu jaeschke@uni-hd.de rjassoa@hotmail.com pjens@koppert.nl princwjhandai@gmail.com tzjia@elsi.jp jia@biochem.mpg.de luis.jdelrio@alumnos.udg.mx edgar.jimeno@educacion.cdmx.gob.mx jinyulong2008@gmail.com joseph@dwi.rwth-aachen.de prijoshi@berkeley.edu jung@lmu.de betul@arizona.edu ville.kaila@dbb.su.se kamimura@complex.c.u-tokyo.ac.jp ilayda.kantarci@campus.lmu.de cansukapdann@gmail.com gulcekardes@gmail.com

Biological Chemistry for Molecular Science

Córdoba

ch (IISER), Mohali

ch (IISER), Pune

S.No	Last Name	First Name	Affiliations	Country	Email Address	S.No	Last Name	First Name	Affiliations
301.	Kaur	Harpreet	University of Strasbourg	France	harpreet.23_purewal@yahoo.it	351.	Le Vay	Kristian	Max Planck Institute of Biochemistry
302.	Кау	Alan	University of Iowa	United States	alan-kay@uiowa.edu	352.	Lee	Kyeong Ro	Ludwig Maximilian University, Munich
303.	Keating	Christine	Pennsylvania State University	United States	cmd8@psu.edu	353.	Lee	Si Young	University of California, Santa Barbara
304.	Kedzior	Mateusz	University of Arizona	United States	mati@arizona.edu	354.	Leman	Luke	Scripps Research
305.	Kempf	Felix	Ludwig Maximilian University, Munich	Germany	felix.kempf@physik.uni-muenchen.de	355.	Leung	Ching Yee	Ludwig Maximilian University, Munich
306.	Kenchel	Josh	University of California, Santa Barbara	United States	jkenchel@ucsb.edu	356.	Leveau	Gabrielle	University of Stuttgart
307.	Khaiwal	Sakshi	Université Cote d'Azur	India	sakshikhaiwal@gmail.com	357.	Li	Yongda	Dublin City University
308.	Khanna	Manavika	Indian Institute of Science Education and Research (IISER), Bhopal	India	manavika18@iiserb.ac.in	358.	Li	Yamei	Earth-Life Science Institute, Tokyo Institute of Te
309.	Khawaja	Nozair	Freie Universität Berlin	Germany	nozair.khawaja@fu-berlin.de	359.	Li	Bingsheng	Ludwig Maximilian University, Munich
310.	Kiani	Armin	University of Groningen	Netherlands	a.kiani@rug.nl	360.	Li	Carmen	University of Calgary
311.	Kilian	Lara	Heidelberg University	Germany	lara.kilian@stud.uni-heidelberg.de	361.	Liang	Dehai	Peking University
312.	Kilicarslan	Dilara	İnha University	Korea, Republic of	dilarakilicarslanscience@gmail.com	362.	Lichtenberg	Tim	University of Oxford
313.	Kimura	Hiroyuki	University of Tokyo	Japan	hkimura@chem.s.u-tokyo.ac.jp	363.	Liedl	Tim	Ludwig Maximilians University, Munich
314.	King	Ashley	Open University	United Kingdom	ashley.king@open.ac.uk	364.	Liu	Ziwei	MRC Laboratory of Molecular Biology
315.	Klenner	Fabian	Freie Universität Berlin	Germany	f.klenner@fu-berlin.de	365.	Liu	Wei	University of Freiburg
316.	Klumpe	Sven	Max Planck Institute of Biochemistry	Germany	klumpe@biochem.mpg.de	366.	Liu	Kai	University of Groningen
317.	Knipe	Peter	Queen's University Belfast	United Kingdom	p.knipe@qub.ac.uk	367.	Löffler	Philipp	University of Southern Denmark
318.	Koeglmayr	Daniel		Germany	da.koeg@gmail.com	368.	Loftus	Kaitlyn	Harvard University
319.	Kohler	Kai	Ludwig Maximilian University, Munich	Germany	kai.kohler@cup.uni-muenchen.de	369.	Longo	Savino	Università degli studi di Bari Aldo Moro
320.	Kohman	Richie	Wyss Institute	United States	richie.kohman@wyss.harvard.edu	370.	López Martínez	Patricia	University of Murcia
321.	Koksal	Elif	University of Oslo, Centre for Molecular Medicine	Norway	elif.koksal@ncmm.uio.no	371.	Lozoya Colinas	Adriana	Georgia Institute of Technology
322.	Komatsu	Yamato	University of Tokyo	Japan	ykomatsu@chem.s.u-tokyo.ac.jp	372.	Lunshof	Jeantine	Harvard Wyss Institute for Biologically Inspired
323.	Konnyu	Balazs	Eotvos Lorand University	Hungary	balazs.konnyu@gmail.com	373.	Lupas	Andrei	Max Planck Institute for Developmental Biology
324.	Kosikova	Tamara	Northwestern University	United States	tamara.kosikova@northwestern.edu	374.	Macdonald	Emyr	Cardiff University
325.	Kostina	Nina	DWI - Leibniz Institute for Interactive Materials e.V.	Germany	kostina@dwi.rwth-aachen.de	375.	Macey	Michael	Open University
326.	Kraemmer	Ulrike	Vienna University of Technology	Austria	ullikraemmer@gmail.com	376.	MacLeod	Rebecca	Illumina Inc.
327.	Kriebisch	Brigitte	Technical University of Munich	Germany	brigitte.kriebisch@tum.de	377.	Madhekar	Mukul	Fergusson College Pune
328.	Kriebisch	Christine	Technical University of Munich	Germany	christine.kriebisch@tum.de	378.	Madrigal	David	Universidad Nacional Autónoma de México
329.	Kucukturhan	Aysu	Centre for Molecular Medicine Norway	Norway	aysu.kucukturhan@ncmm.uio.no	379.	Madrigal Luna	Patricio	ITNM
330.	Kudella	Patrick	Ludwig Maximilian University, Munich	Germany	patrick.kudella@physik.uni-muenchen.de	380.	Magalhaes	Alvaro	University College London
331.	Kufner	Corinna	Harvard University	United States	corinna.kufner@cfa.harvard.edu	381.	Maguire	Oliver	Radboud University
332.	Kühnlein	Alexandra	Ludwig Maximilian University, Munich	Germany	alexandra.kuehnlein@physik.uni-muenchen.de	382.	Mann	Stephen	University of Bristol
333.	Kulkarni	Chandrashekhar V.	University of Central Lancashire	United Kingdom	cvkulkarni@uclan.ac.uk	383.	Marchand	Jorge	Harvard Medical School
334.	Kumar	Ravi		India	ravi.chemcdri@gmail.com	384.	Maria	Giraldez	Institute of BioMedicine of Seville
335.	Kundu	Subhradip		India	kundusubhradip@gmail.com	385.	Martin	Nora	University of Cambridge
			Vamaguchi University	United States		386.	Martínez Martín		onversity of cambridge
336. 337	Kurisu	Kanta	Yamaguchi University Tohoku University		kanta.k1997@gmail.com	386.		Carlos	Ludwig Maximilian University Munich
337.	Kurisu	Minoru	·	Japan	kurisu@bio.phys.tohoku.ac.jp		Maryshev	Ivan	Ludwig Maximilian University, Munich
338.	Kuster	David	Max Planck Institute for Molecular Cell Biology and Genetics	Germany	kuster@mpi-cbg.de	388.	Maslov	Sergei	University of Illinois
339.	Kuzmanka	Miriam	Max Planck School Matter to Life	Germany	miriam.kuzman@gmx.de	389.	Mast	Christof	Ludwig Maximilian University, Munich
340.	Kuzmenka	Aliaksei	Lund University	Sweden	kozjavka2000@gmail.com	390.	Mateos Salmon	Carolina	Universidad de Guadalajara
341.	Lacinbala	Ozan	Institut des Sciences Moléculaires d'Orsay	France	ozan.lacinbala@u-psud.fr	391.	Matreux	Thomas	Ludwig Maximilian University, Munich
342.	Lacoste	David	CNRS	France	david.lacoste@gmail.com	392.	Matsubara	Yoshiya	National Centre for Biological Sciences, TIFR
343.	Lagki	Ntea	Democritus University of Thrace	Greece	alex.lagki8@gmail.com	393.	Matsumoto	Mio	Tokyo University of Science
344.	Laha 	Sudarshana	Max Planck Institute for the Physics of Complex Systems	Germany	sudarshana@pks.mpg.de	394.	Matsuo	Muneyuki	Hiroshima University
345.	Laimer	Tobias	Ludwig Maximilian University, Munich	Germany	t.laimer@physik.uni-muenchen.de	395.	Matsuura	Tomoaki	Osaka University
346.	Landim	Ricardo	Technical University of Munich	Germany	ricardo.landim@tum.de	396.	Matzka	Marco	Helmholtz Zentrum München
347.	Langer	Michael	Technical University of Munich	Germany	michl.langer@tum.de	397.	McCalder	Janine	University of Calgary
348.	Langlais	Juliette	Ludwig Maximilian University, Munich	Germany	juliette.langlais@physik.uni-muenchen.de	398.	McGeoch	Julie	Harvard University
349.	Lau	Graham	Blue Marble Space	United States	grahamlau@bmsis.org	399.	Meierhenrich	Uwe	Université Côte d'Azur
350.	Lauber	Nino	University of the Basque Country	Spain	nino.lauber@ehu.eus	400.	Meinert	Cornelia	CNRS - Université Côte d'Azur

. + M		
Se se		
6	/	

Country

Email Address

Germany levay@biochem.mpg.de kyeong.lee@physik.uni-muenchen.de Germany United States siyounglee@ucsb.edu United States lleman@scripps.edu c.leung@physik.uni-muenchen.de Germany gabrielle.leveau@oc.uni-stuttgart.de Germany Ireland yongda.li26@mail.dcu.ie yamei.li@elsi.jp Japan Germany lbseeu@outlook.com Canada carmen.li@ucalgary.ca China dliang@pku.edu.cn United Kingdom tim.lichtenberg@physics.ox.ac.uk Germany tim.liedl@physik.lmu.de United Kingdom zliu@mrc-lmb.cam.ac.uk Germany wei.liu@makro.uni-freiburg.de Netherlands k.liu@rug.nl pmgl@sdu.dk Denmark United States kloftus@g.harvard.edu Italy savino.longo@uniba.it patricia.lopez16@um.es Spain alc6@gatech.edu United States United States jeantine.lunshof@wyss.harvard.edu andrei.lupas@tuebingen.mpg.de Germany United Kingdom macdonald@cardiff.ac.uk United Kingdom michael.macey@open.ac.uk United States rebecca.m.turk@gmail.com India madhekarmukul@gmail.com damt@ciencias.unam.mx Mexico patoml07@hotmail.com Mexico United Kingdom a.magalhaes@ucl.ac.uk Netherlands o.r.maguire@gmail.com United Kingdom S.Mann@bristol.ac.uk United States marchand@hms.harvard.edu giraldezjimenez@hotmail.com Spain United Kingdom nsm36@cam.ac.uk Spain carlos.biochem@gmail.com ivan.maryshev@physik.uni-muenchen.de Germany United States maslov@illinois.edu Germany christof.mast@physik.uni-muenchen.de Mexico carolina.mateos@alumnos.udg.mx th.matreux@physik.lmu.de Germany India yoshiyam@ncbs.res.in miomatsumoto110@gmail.com Japan muneyuki@hiroshima-u.ac.jp Japan Japan matsuura\_tomoaki@bio.eng.osaka-u.ac.jp Germany Canada jamccald@ucalgary.ca United States mcgeoch@fas.harvard.edu France uwe.meierhenrich@unice.fr France cornelia.meinert@univ-cotedazur.fr

of Technology

ired Engineering

S.No	Last Name	First Name	Affiliations	Country	Email Address	S.No	Last Name	First Name	Affiliations	Country	Email Address
401.	Mendoza	Hanna	Cinvestav MTY	Mexico	hanna.mendoza@cinvestav.mx	451.	Nichols	Claire	МІТ	United States	cion2@mit.edu
402.	Meringer	Markus	DLR - German Aerospace Center	Germany	markus.meringer@dlr.de	452.	Nishikawa	Shota	Tokyo Institute of Technology	Japan	nishikawa.s.ae@m.titech.ac.jp
403.	Meselson	Matthew	Harvard University	United States	msmeselson@gmail.com	453.	Nishimura	Hiroki	University of Tokyo	Japan	hiroki-gemini@g.ecc.u-tokyo.ac.jp
404.	Meyer	McCauley	Pennsylvania State University	United States	mxm1357@psu.edu	454.	Nishino	Mayu	Tokyo Metropolitan University	Japan	nishino-mayu@ed.tmu.ac.jp
405.	Micca Longo	Gaia	Università degli Studi di Bari Aldo Moro	Italy	gaia.miccalongo@uniba.it	455.	Novais	Aline	Federal University of Rio de Janeiro	Brazil	aline12@astro.ufrj.br
406.	Mierzejewski	Veronica	Skidmore College	United States	vmierzej@skidmore.edu	456.	Novoa Vásquez	Jose	Sociedad Científica de Astrobiología del Perú - Filial Trujillo	Peru	jnovoa2001@gmail.com
407.	Miladi	Milad	University of Freiburg	Germany	miladim@cs.uni-freiburg.de	457.	Nozaki	Shunsuke	Tohoku University	Japan	syunsuke.nozaki.q8@dc.tohoku.ac.jp
408.	Miranda Rosete	Arturo	Universidad Nacional Autónoma de México, ICN	Mexico	arturo.miranda@correo.nucleares.unam.mx	458.	Nutman	Allen	University of Wollongong	Australia	anutman@uow.edu.au
409.	Miranda-Perez	Ingrid	Universidad Nacional Autónoma de México	Mexico	ingridmipe@ciencias.unam.mx	459.	Oberg	Karin	Harvard University	United States	koberg@cfa.harvard.edu
410.	Mislik	Franz	Ludwig Maximilian University, Munich	Germany	f.mislik@campus.lmu.de	460.	Ochsenfeld	Christian	Ludwig Maximilian University, Munich	Germany	christian.ochsenfeld@uni-muenchen.de
411.	Mitra	Saibal		Netherlands	smitra00@gmail.com	461.	Olivi	Lorenzo	Wageningen University and Research	Netherlands	lorenzo.olivi@wur.nl
412.	Mizukami	Yusuke	Tokyo Institute of Technology	Japan	yukke.0616.m@gmail.com	462.	Olmedo Aguilar	José Manuel	INAOE	Mexico	cosmocaos@hotmail.com
413.	Mizuno	Yuta	Kanazawa Institute of Technology	Japan	curiosity0529@gmail.com	463.	Olmez	Tolga	Yale University	United States	tolga.olmez@yale.edu
414.	Mizuuchi	Ryo	University of Tokyo, Komaba Institute for Science	Japan	mizuuchi@bio.c.u-tokyo.ac.jp	464.	Omarova	Diana	Sabanci University	Turkey	dianaomarova@sabanciuniv.edu
415.	Mohanty	Anurup	SRM Institute of Science & Technology	India	1anurupmohanty@gmail.com	465.	Ornelas Guzmán	Erandhi Claudel	Universidad Nacional Autónoma de México	Mexico	erandhi@ciencias.unam.mx
416.	Mohit	Farhad	University of Sannio	Italy	farhadm@gmail.com	466.	Orsi	William	Ludwig Maximilian University, Munich	United States	w.orsi@lrz.uni-muenchen.de
417.	Mohite	Atul Tanaji	Ludwig Maximilian University, Munich	Germany	atul.mohite@physik.uni-muenchen.de	467.	Ortega	Julian	Universidad de Sonora	Mexico	jbotello690@gmail.com
418.	Möhler	Jasper	ETH Zurich	Switzerland	jasper.moehler@org.chem.ethz.ch	468.	Ortega Cano	Manuel Eduardo	Universidad Autónoma de Campeche	Mexico	al039513@hotmail.com
419.	Mojzsis	Stephen	University of Colorado	United States	mojzsis@colorado.edu	469.	Ortiz	Jorge	Universidad Autónoma de Chiapas	Mexico	jorge052ortiz@gmail.com
420.	Molaverdikhani	Karan	Max Planck Institute for Astronomy	Germany	karan@mpia.de	470.	Otieno	Kepher	KEMRI	Kenya	kepher.otieno98@gmail.com
421.	Morales	Abigail	CDMX	Mexico	abi.brmc@gmail.com	471.	Ouldridge	Tom	Imperial College London	United Kingdom	t.ouldridge@imperial.ac.uk
422.	Moran	Joseph	University of Strasbourg & CNRS	France	moran@unistra.fr	472.	Ozcan	Umut	Hacettepe University	Turkey	u.onurozcan@gmail.com
423.	Moran Tovar	Roberto	University of Cologne	Germany	r.morantovar@uni-koeln.de	473.	Öztürk	Zeynep	University of Cambridge	United Kingdom	zo216@cam.ac.uk
424.	Moura	Carina	University of Göttingen	Germany	carinamoura@uni-goettingen.de	474.	Pachiadaki	Maria	Woods Hole Oceanographic Institution	United States	mpachiadaki@whoi.edu
425.	Muchowska	Kamila	University of Strasbourg	France	muchowska@unistra.fr	475.	Pahuja	Anushka	Delhi University	India	pahuja4342@gmail.com
426.	Mueckl	Andrea	Technical University of Munich	Germany	andrea.mueckl@tum.de	476.	Panda	Debiprasad	Indian Institute of Science Education and Research (IISER), Pune	India	debiprasadpanda235@gmail.com
427.	Mukhtar	Usama		New Zealand	usamamukhtar90@gmail.com	477.	Paneque	David	Max Planck Institute for Physics	Germany	dpaneque@mppmu.mpg.de
428.	Mulewar	Sahil	Indian Institute of Science Education and Research (IISER), Pune	India	sahil.mulewar@students.iiserpune.ac.in	478.	Papanikolaou	Nikos	ETH Zurich	Switzerland	nikos-papanikolaou@hotmail.com
429.	Müller	Felix	Ludwig Maximilian University, Munich	Germany	felix.mueller@cup.uni-muenchen.de	479.	Parejo Vidal	Ana Delia	University of California, Santa Barbara	USA	aparejovidal@ucsb.edu
430.	Mumcuoglu	Ecenaz	Istanbul University	Turkey	ecenazmumcuoglu@gmail.com	480.	Pariona Velarde	Carlos Daniel	SCAP	Peru	jodanielbs@gmail.com
431.	Munera	Amalia		Colombia	amaliamunerab@gmail.com	481.	Partida	Eira Donaji	IBT	Mexico	eira_par@yahoo.com
432.	Munguia	Akari	IPN	Mexico	akarimr17@gmail.com	482.	Patki	Gauri	Indian Institute of Science Education and Research (IISER), Pune	India	patki.gauri@students.iiserpune.ac.in
433.	Muñoz Basagoiti	Maitane	ESPCI Paris, Gulliver Lab	France	maitane.munoz-basagoiti@espci.fr	483.	Patra	Satyajit	JNCASR	India	satyajit@jncasr.ac.in
434.	Muramatsu	David	Ludwig Maximilian University, Munich	Germany	d.muramatsu@physik.uni-muenchen.de	484.	Paul	Christian		Germany	ichpaul@gmx.net
435.	Murayama	Hanako	University of Tokyo	Japan	hmurayama0425@g.ecc.u-tokyo.ac.jp	485.	Pazienza	Lydia	Harvard University	United States	lpazienza@g.harvard.edu
436.	Mutlu	Ozal	Marmara University	Turkey	ozal.mutlu@marmara.edu.tr	486.	Peñaloza Mendoza	Dante Enrique		Mexico	union7_@hotmail.com
437.	Mutschler	Hannes	Max Planck Institute of Biochemistry	Germany	mutschler@biochem.mpg.de	487.	Penning	Alexander	Heidelberg University	Germany	a.penning@stud.uni-heidelberg.de
438.	Ν	SREESHMA	IARI	India	sreeshma09n@gmail.com	488.	Penzinger	Julian	University of Vienna, Institute of Astrophysics	Austria	julian.penzinger@aon.at
439.	Ν	Vikas		Canada	vikas.nath@gmail.com	489.	Pereira	Joana	Max Planck Institute for Developmental Biology	Germany	joana.pereira@tuebingen.mpg.de
440.	Nader	Serge	University of Alberta	Canada	serge.nader@ualberta.ca	490.	Pérez	Gilberto	Cinvestav	Mexico	gilberto.perez@cinvestav.mx
441.	Nakamura	Keisuke	Kyoto University	Japan	nakamura.keisuke.43n@st.kyoto-u.ac.jp	491.	Peter	Benedikt	Max Planck Institute of Biochemistry	Germany	peter@biochem.mpg.de
442.	Nakano	Taiki	Tohoku University	Japan	taikinakano0717@gmail.com	492.	Philp	Douglas	University of St Andrews	United Kingdom	d.philp@st-andrews.ac.uk
443.	Namani	Trishool	University of Akron	United States	tnamani@uakron.edu	493.	Pimentel	Karen	Universidad Autónoma de Chiapas	Mexico	moyapim03@gmail.com
444.	Nanda	Jayanta	Indian Institute of Engineering Science and Technology (IIEST), Shibpur	India	drjayantananda16@gmail.com	494.	Pipilos	Atakan		Turkey	atakanpipilos03@gmail.com
445.	Narasimhan	Kamesh	Harvard Medical School	United States	kamesh_narasimhan@hms.harvard.edu	495.	Pir Cakmak	Fatma	MIT	United States	fatmapir@mit.edu
446.	Nasufovska	Atida	Georg-August-Universität Göttingen	Germany	anasufo@gwdg.de	496.	Pirzer	Tobias	Technical University of Munich	Germany	pirzer@tum.de
447.	Nath	Artash		Canada	artash.nath@gmail.com	497.	Pitts	J. Brian	University of Cambridge	United Kingdom	jbp25@cam.ac.uk
448.	Neubauer	Сај	California Institution of Technology, Hanse-Wissenschaftskolleg	Germany	123caj@gmail.com	498.	Pogodaev	Aleksandr	Radboud University	Netherlands	aleksandr@science.ru.nl
449.	Neupane	Chetanath	Astrobiology Research in Nepal (ARiN)	Nepal	scholar.chetanath@gmail.com	499.	Poldsalu	Inga	University of Oslo	Norway	inga.poldsalu@ncmm.uio.no
450.	Ni	Ziqin	University of Maryland	United States	zni@umd.edu	500.	Poros-Tarcali	Eszter	Harvard University	United States	eporostarcali@fas.harvard.edu

S.No	Last Name	First Name	Affiliations	Country	Email Address	S.No	Last Na
501.	Poudyal	Raghav	Pennsylvania State University	United States	rup34@psu.edu	551.	Romero Ma
502.	Powner	Matt	University College London	United Kingdom	matthew.powner@ucl.ac.uk	552.	Rosenberg
503.	Pramanik	Bapan		Israel	bapanpramanik92@gmail.com	553.	Rossetto
504.	Preiner	Martina	Heinrich Heine University	Germany	preiner@hhu.de	554.	Rout
505.	Prondzinsky	Paula	Earth-Life Science Institute, Tokyo Institute of Technology	Japan	prondzinsky@elsi.jp	555.	Rubén de J
506.	Pudritz	Ralph	McMaster University	Canada	pudritz@mcmaster.ca	556.	Ruf
507.	Radakovic	Aleksandar	Harvard Medical School/Massachusetts General Hospital	United States	aradakovic@g.harvard.edu	557.	S
508.	Rahimi	Khosrow	DWI - Leibniz Institute for Interactive Materials e.V.	Germany		558.	Saad
509.	Rahimzadeh	Mehrnoush	DWI - Leibniz Institute for Interactive Materials e.V.	Germany	rahimzadeh@dwi.rwth-aachen.de	559.	Sahai
510.	Rai	Tina	University of Chinese Academy of Sciences	China	dimmalikrisha123@gmail.com	560.	Sakamoto
511.	Raith	Johannes	Technical University of Munich	Germany	johannes.raith@tum.de	561.	Salaveri
512.	Raja	Anand	Delft University of Technology	Netherlands	a.raja@student.tudelft.nl	562.	Salditt
513.	Rajamani	Sudha	Indian Institute of Science Education and Research (IISER), Pune	India	srajamani@iiserpune.ac.in	563.	Saldivar
514.	Ramírez	Alejandra	Preparatoria Federal Lázaro Cárdenas	Mexico	alerammmz@gmail.com	564.	Saleski
515.	Ramírez	Luisa	Universidad Nacional Autónoma de México, Ciencias de la Tierra	Mexico	luisalawliet@gmail.com	565.	Salibi
516.	Rani Malta	Fernanda	Federal University of São Carlos (UFSCar)	Brazil	fernanda.maltacd@gmail.com	566.	Samanta
517.	Rao	Anish	Indian Institute of Science Education and Research (IISER), Pune	India	anishrao95@gmail.com	567.	Samanta
518.	Rasheed	Rabeea	BMSIS	Pakistan	rabeea.rasheed1@gmail.com	568.	Samoilova
519.	Rattray	Jayne	University of Calgary	Canada	jayne.rattray@ucalgary.ca	569.	Sanchez O
520.	Rau	Maximilian	Ludwig Maximilian University, Munich	Germany	maximilian.rau@physik.uni-muenchen.de	570.	Santiago
521.	Raulin	Jeanine		France	raulinj@free.fr	571.	Santos
522.	Rauscher	Sophia	University of Strasbourg	France	srauscher@unistra.fr	572.	Sarkar
523.	Reginato	Paul	MIT	United States	reginato@mit.edu	573.	Sasselov
524.	Reichert	Michael		Germany	mmichaelreichert@gmail.com	574.	Schanke
525.	Rein	Christina	Ludwig Maximilian University, Munich	Germany	c.rein@physik.uni-muenchen.de	575.	Scheu
526.	Reitner	Joachim	University of Göttingen	Germany	jreitne@gwdg.de	576.	Schmidt
527.	Remsing	Rick	Rutgers University	United States	rick.remsing@rutgers.edu	577.	Schmitt
528.	Renard	Andre	University of Liege, Montefiore Institute	Belgium	arenard@uliege.be	578.	Schmitt-Ko
529.	Restrepo Sierra	Ana Maria	Delft University of Technology	Netherlands	a.m.restreposierra@tudelft.nl	579.	Schnitter
530.	Reyes	Rosa	Universidad Nacional de Colombia	Colombia	rareyesq@unal.edu.co	580.	Schnitzer
531.	Richert	Clemens	University of Stuttgart	Germany		581.	Schoenma
532.	Rieß	Benedikt	Technical University of Munich	Germany	riess.benedikt@gmail.com	582.	Schreiner
533.	Rimmer	Paul Brandon	University of Cambridge	United Kingdom	pbr27@cam.ac.uk	583.	Schulte-Kra
534.	Ritson	Dougal	MRC Laboratory of Molecular Biology	United Kingdom	dritson@mrc-Imb.cam.ac.uk	584.	Schwille
535.	Rivas Medrano	Mario	NASA, Astrobiology Institute - University of Houston	United States	mrivasmedrano@uh.edu	585.	Schwintek
536.	Roa	Diana	Universidad Autónoma de Chiapas	Mexico	diana.roa.dra@gmail.com	586.	Sciortino
537.	Robidillo	Christopher Jay	University of Alberta	Canada	robidill@ualberta.ca	587.	Sedaghatp
538.	Robles Hernández	Tania Maria	Mexican Space Agency	Mexico	taniarblsh@gmail.com	588.	Seitz
539.	Robu	Irina	Radboud University	Netherlands	i.robu@student.ru.nl	589.	Selcuk
540.	Rocha	Carolina	Charles University	Czechia	sanchezalm@natur.cuni.cz	590.	Serrao
541.	Rodon Fores	Jennifer	Technical University of Munich	Germany	jennifer.rodon-fores@tum.de	591.	Sevgen
542.	Rodriguez	Ignacio	Universidad Nacional Autónoma de México	Mexico	jirobiolexp@gmail.com	592.	Sevgen
543.	Rodriguez	David	Universidad Nacional de Colombia	Colombia	dftovarr@unal.edu.co	593.	Shah
544.	Rodríguez Arteaga	Alfredo	Universidad Nacional Autónoma de México	Mexico	alfredo.r.arteaga@ciencias.unam.mx	594.	Shaikh
545.	Rodriguez Ramirez	Paula Catalina	ETH Zurich	Switzerland	pramirez@ethz.ch	595.	Shapiro
546.	Rodriguez Robles	Emilio	ETH Zurich	Switzerland	emilio.rodriguez@bsse.ethz.ch	596.	Sharma
547.	Rodríguez Torres	Luis Miguel	Benemérita Universidad Autónoma de Puebla	Mexico	luis_mrt1995@hotmail.com	597.	Sharma
548.	Rodriguez-Emmenegger		DWI - Leibniz Institute for Interactive Materials e.V.	Germany	rodriguez@dwi.rwth-aachen.de	598.	Shirai
549.	Rodríguez-Román	Eduardo	Venezuelan Institute for Scientific Research	Venezuela	erodriguezroman@gmail.com	599.	Shorttle
550.	Roias	Tatihana	Universidad Industrial de Santander	Colombia	tatihanaroiasg@gmail.com	600.	Siedler

Colombia

tatihanarojasg@gmail.com

600. Siedler

Frank

Max Planck Institute of Biochemistry

550. Rojas

Tatihana

Universidad Industrial de Santander

10	Last Name	First Name	Affiliations
51.	Romero Martin	Juan Manuel	iHM Solutions
52.	Rosenberger	Joachim	Technical University of Munich
53.	Rossetto	Daniele	University of Trento
54.	Rout	Saroj Kumar	ETH Zurich
55.	Rubén de Jesús	Tovilla Quesada	Instituto Politécnico Nacional
56.	Ruf	Alexander	Université Aix-Marseille
57.	S	Prashanth	National Centre for Biological Sciences
58.	Saad	Noy	Ben-Gurion University of the Negev
59.	Sahai	Nita	University of Akron
60.	Sakamoto	Ryota	London Centre for Nanotechnology
61.	Salaveri	Anna	University of Edinburgh
62.	Salditt	Annalena	Ludwig Maximilian University, Munich
63.	Saldivar	Alexis	Universidad Autónoma Metropolitana
64.	Saleski	Tatyana	ESPCI Paris
65.	Salibi	Elia	Max Planck Institute of Biochemistry
66.	Samanta	Mousumi	Ben-Gurion University of the Negev
67.	Samanta	Avik	University of Freiburg - Humboldt Research Fellow
68.	Samoilova	Ekaterina	Federal Research and Clinical Center
69.	Sanchez Ospina	Cristhian Mateo	Universidad Nacional de Colombia
70.	Santiago	Ibon	Technical University of Munich
71.	Santos	Madeleine	University of the Philippines
72.	Sarkar	Susovan	Indian Institute of Science Education and Research (IISEF
73.	Sasselov	Dimitar	Harvard University
74.	Schanke	Ingrid Jin	University of Oslo
75.	Scheu	Bettina	Ludwig Maximilian University, Munich
76.	Schmidt	Fabian	University of Bremen
77.	Schmitt	Christian	Heidelberg University
78.	Schmitt-Kopplin	Philippe	Helmholtz Zentrum München
79.	Schnitter	Fabian	Technical University of Munich
80.	Schnitzer	Tobias	Eindhoven University of Technology
81.	Schoenmakers	Ludo	Radboud University
82.	Schreiner	Peter Richard	Justus Liebig University, Institute of Organic Chemistry
83.	Schulte-Krauss	Andrea	CRC 235 - Emergence of Life
84.	Schwille	Petra	Max Planck Institute of Biochemistry
85.	Schwintek	Philipp	Ludwig Maximilian University, Munich
86.	Sciortino	Alfredo	Technical University of Munich
87.	Sedaghatpour	Fatemeh	Harvard University
88.	Seitz	Christian	Technical University of Munich
89.	Selcuk	Zeynep Su	Ludwig Maximilian University, Munich
90.	Serrao	Adriana	Ludwig Maximilian University, Munich
91.	Sevgen	Sılanur	Middle East Technical University
92.	Sevgen	Serhat	Middle East Technical University, Institute of Marine Scien
93.	Shah	Dhruvi	Maharaja Sayajirao University of Baroda, India
94.	Shaikh	Aaqifa	
95.	Shapiro	Irwin	Center for Astrophysics Harvard Smithsonian
96.	Sharma	Sunanda	MIT
97.	Sharma	Siddhant	University of Delhi
98.	Shirai	Mizuho	Kyushu University
99.	Shorttle	Oliver	University of Cambridge
00.	Siedler	Frank	Max Planck Institute of Biochemistry

Country

Email Address

Spain	romero.martin.jm@gmail.com
Germany	joachim.h.rosenberger@tum.de
Italy	daniele.rossetto@unitn.it
Switzerland	saroj.rout@phys.chem.ethz.ch
Mexico	investigacionmb2@gmail.com
France	rufalexan@gmail.com
India	prash231097@gmail.com
Israel	nsa@post.bgu.ac.il
United States	sahai@uakron.edu
United Kingdom	ryota.sakamoto@ucl.ac.uk
United Kingdom	s1720560@ed.ac.uk
Germany	a.salditt@physik.uni-muenchen.de
Mexico	asaldivarg93@gmail.com
France	tatyana.saleski@espci.fr
Germany	salibi@biochem.mpg.de
Israel	mousumi.wb@gmail.com
Germany	avik.samanta@makro.uni-freiburg.de
Russian Federation	samoyket@gmail.com
Colombia	cmsanchezos@unal.edu.co
Germany	ibon.santiago@tum.de
Philippines	mcsantos7@up.edu.ph
India	susovan.sarkar@students.iiserpune.ac.in
United States	dsasselov@cfa.harvard.edu
Norway	i.j.schanke@kjemi.uio.no
Germany	b.scheu@lmu.de
Germany	f.schmidt@uni-bremen.de
Germany	sh401@uni-heidelberg.de
Germany	schmitt-kopplin@helmholtz-muenchen.de
Germany	fabian.schnitter@tum.de
Netherlands	t.schnitzer@tue.nl
Netherlands	ludo.schoenmakers@ru.nl
Germany	prs@uni-giessen.de
Germany	a.schulte-krauss@physik.uni-muenchen.de
Germany	schwille@biochem.mpg.de
Germany	philipp.schwintek@physik.uni-muenchen.de
Germany	alfredo.sciortino@tum.de
United States	fsedaghatpour@fas.harvard.edu
Germany	c.seitz@tum.de
Germany	zeynepsuselcuk@hotmail.com
Germany	a.serrao@physik.uni-muenchen.de
Turkey	sila.sevgen@gmail.com
Turkey	sevgenserhat@gmail.com
India	dhruvishah2501@gmail.com
India	aaqifa2000@gmail.com
United States	ishapiro@cfa.harvard.edu
United States	sunanda.sharma.92@gmail.com
India	siddhaantsharma.ss@gmail.com
Japan	shirai.mizuho@phys.kyushu-u.ac.jp
United Kingdom	shorttle@ast.cam.ac.uk
Germany	siedler@biochem.mpg.de

earch (IISER), Pune

arine Sciences

S.No	Last Name	First Name	Affiliations	Country	Email Address	S.No	Last Name	First Name	Affiliations
601.	Silva-Flores	Adan	Centro de Investigaciones Biológicas del Noroeste	Mexico	asilva@pg.cibnor.mx	651.	Todisco	Marco	Università degli Studi di Milano
602.	Simmel	Friedrich	Technical University of Munich	Germany	simmel@tum.de	652.	Tomar	Anju	University of Trento
603.	Simonis	Philippe	EPS	Belgium	philippejustin.simonis@gmail.com	653.	Tomohara	Kanji	University of Tokyo
604.	Singh	Akash	University of Delhi	India	incurablewriter7@gmail.com	654.	Torres Ramírez	Victoria	Universidad Nacional Autónoma de México
605.	Singh	Jyoti	University of Freiburg	Germany	singhjyoti103@gmail.com	655.	Toru (Shay)	Hannah	University of Illinois at Urbana-Champaign
606.	Singh	Mohnish	University of Lucknow	India	mohnishsingh1@gmail.com	656.	Toussaint	Pablo	Ludwig Maximilian University, Munich
607.	Sireci	Matteo	University of Granada	Spain	msireci@onsager.ugr.es	657.	Tower	David	Rockport Public Schools
608.	Sivagnanam	Ulaganathan	Indian Institute of Technology Madras	India	ulags.iitm@gmail.com	658.	Trapp	Oliver	Ludwig Maximilian University, Munich
609.	Skaf	Nour	PSL - Paris Observatory	France	nour.skaf@gmail.com	659.	Tröger	Lucas	Ludwig Maximilian University, Munich
610.	Smokers	Iris	Radboud University	Netherlands	i.smokers@student.ru.nl	660.	Tsanakopoulou	Maria	
611.	Söder	Dominik	DWI - Leibniz Institute for Interactive Materials e.V.	Germany	soeder@dwi.rwth-aachen.de	661.	Turpin	Victor	Université Paris Saclay
612.	Song	Emilie	Max Planck Institute of Biochemistry	Germany	ysong@biochem.mpg.de	662.	Tych	Katarzyna	University of Groningen
613.	Soriano López	Carlos Alberto	Universidad Autónoma del Estado de México	Mexico	carlos.sorianol@uaem.edu.mx	663.	Ucan	Deniz Davide	Goethe University Frankfurt am Main
614.	Spezzano	Silvia	Max Planck Institute for Extraterrestrial Physics	Germany	spezzano@mpe.mpg.de	664.	Ucar	Fatih	Koc University
615.	Spourita	Maria		Greece	marispou02@gmail.com	665.	Upadhyay	Abhishek	Institute for Theoretical Biology
616.	Springsklee	Christina	Ludwig Maximilian University, Munich	Germany	christina.springsklee@min.uni-muenchen.de	666.	Urazov	Aman	Radboud University
617.	Spustova	Karolina	Centre for Molecular Medicine Norway	Norway	karolina.spustova@ncmm.uio.no	667.	Vahedian Movahed	Hanif	Harvard University
618.	Stadler	Rob		United States	stadlerrw@gmail.com	668.	Valencia Guzmán	Jasiel Alan	Universidad Nacional Autónoma de México
619.	Stein	Julian	Ludwig Maximilian University, Munich	Germany	julian.stein@physik.uni-muenchen.de	669.	Valencia-Esquivel	Israel	Universidad Autónoma del Estado de Morelos
620.	Steiner	Thomas	Technical University of Munich	Germany	thomas.steiner@tum.de	670.	Valer	Luca	University of Trento
621.	Steller	Luke	University of New South Wales	Australia	l.steller@unsw.edu.au	671.	Valle	Scarlett	Unison
622.	Stolar	Tomislav	Ruđer Bošković Institute	Croatia	tomislav.stolar@irb.hr	672.	van Duppen	Peer	Radboud University
623.	Su	Meng	MRC Laboratory of Molecular Biology	United Kingdom	mengsu@mrc-Imb.cam.ac.uk	673.	van Esch	Jan H.	Delft University of Technology
624.	Subramanian	Hemachander	National Institute of Technology Durgapur	India	hemachander@gmail.com	674.	Vázquez-Salazar	Alberto	University of California, Los Angeles
625.	Sugiyama	Hironori	University of Tokyo	Japan	h-suqiyama@q.ecc.u-tokyo.ac.jp	675.	Vergara Ovando	Cindel	Universidad Nacional Autónoma de México
626.	Sun	Daxiao	Max Planck Institute for Molecular Cell Biology and Genetics	Germany	dsun@mpi-cbq.de	676.	Vijendravarma	Roshan Kumar	Institute Curie
627.	Sun	Tsu-Wang	National Chung Hsing University	Taiwan	stsuwang@smail.nchu.edu.tw	677.	Villarreal Gómez	María José	Universidad Industrial de Santander
628.	Sun	Jing	University of Ulm	Germany	jing.sun@uni-ulm.de	678.	Vishwakarma	Harsh Satyaindra	Government Holkar Science College, Indore
629.	Suranse	Sonali	Prestige Public School Indore Madhya Pradesh	India	sonalisuranse18@gmail.com	679.	Vogel	Julian	Ulm University
630.	Surman	Andrew	King's College, London	United Kingdom	andrew.surman@kcl.ac.uk	680.	Vogele	Kilian	Technical University of Munich
631.	Sydow	Constanze	Ludwig Maximilian University, Munich	Germany	constanze.sydow@cup.uni-muenchen.de	681.	Vollenhofer-Schrumpf	Sabine	SY-Lab Geraete GmbH
632.	Szabla	Rafał	University of Edinburgh	United Kingdom	rafal.szabla@ed.ac.uk	682.	von der Esch	Beatriz	Ludwig Maximilian University, Munich
633.	Szentgyorgyi	Andrew	Center for Astrophysics Harvard Smithsonian	United States	saint@cfa.harvard.edu	683.	Vorobii	Mariia	DWI - Leibniz Institute for Interactive Materials e.V., RW
634.	Szokoli	Deni	Max Planck Institute of Biochemistry	Germany	szokoli@biochem.mpg.de	684.	Vyborna	Yuliia	Sorbonne University
635.	Tablas Alcázar	Laila	Universidad Nacional Autónoma de México	Mexico	sladlai1394@gmail.com	685.	Vybornyi	Mykhailo	ESPCI Paris
636.	Tabone	Benoît	Leiden University	Netherlands	tabone@strw.leidenuniv.nl	686.	Vyle	Joseph	Queen's University Belfast
637.	Tachsin	Achmet	Gebze Technical University	Turkey	tachsinachmet@gmail.com	687.	Wagner	Anna Maria	DWI - Leibniz Institute for Interactive Materials e.V.
638.	Takeuchi	Nobuto	University of Auckland	New Zealand	nobuto.takeuchi@auckland.ac.nz	688.	Waldenmaier	Stefan	ORIGINS
					-				
639.	Tang	Dora	Max Planck Institute for Molecular Cell Biology and Genetics	Germany	tang@mpi-cbg.de	689.	Walton	Craig	University of Cambridge
640.	Tatton	Ben	Open University	United Kingdom	ben.tatton@open.ac.uk	690.	Wang	Anna	University of New South Wales
641.	Tebcharani	Laura	Technical University of Munich	Germany	I.tebcharani@gmx.de	691.	Wanner	Barry	Harvard Medical School
642.	Teders	Michael	Radboud University	Netherlands	michael.teders@ru.nl	692.	Weber	Christoph A.	Max Planck Institute for the Physics of Complex System
643.	Tenbusch	Jan	DWI - Leibniz Institute for Interactive Materials e.V.	Germany	tenbusch@dwi.rwth-aachen.de	693.	Weidmann	Laura	Max Planck Institute for Developmental Biology
644.	Terrazas Esparza	Diana Laura		Mexico	dianalaura.te@gmail.com	694.	Weightman	Peter	University of Liverpool
645.	Thimaradka	Vikram	Kyoto University	Japan	thimaradka.vikram.38z@st.kyoto-u.ac.jp	695.	Weingart	Maximilian	Ludwig Maximilian University, Munich
646.	Thole	Esther	NEMO Kennislink	Netherlands	thole@nemokennislink.nl	696.	Weise	Laura	Max Planck Institute of Biochemistry
647.	Thoma	Benjamin	University College London	United Kingdom	uccabth@ucl.ac.uk	697.	Weller	Daniel	Ludwig Maximilian University, Munich
648.	Thordarson	Pall	University of New South Wales	Australia	p.thordarson@unsw.edu.au	698.	Wenzel	Eric	
649.	Thutupalli	Shashi	National Centre for Biological Sciences	India	shashi@ncbs.res.in	699.	Wieczorek	Tom	University of Bristol
650.	Tikhomirov	Grigory	California Institute of Technology	United States	dnano@caltech.edu	700.	Wiedemann	Stefan	Ludwig Maximilian University, Munich



Italy	marco.todisco@unimi.it
Italy	anju.tomar@unitn.it
Japan	tomohara.k18@smb.t.u-tokyo.ac.jp
Mexico	torres.victoria@ciencias.unam.mx
United States	hshay2@illinois.edu
Germany	p.toussaint@campus.lmu.de
United States	dtower51147@gmail.com
Germany	oliver.trapp@cup.uni-muenchen.de
Germany	lucas.troeger@physik.uni-muenchen.de
United Kingdom	mtsanako@gmail.com
France	victor.turpin@universite-paris-saclay.fr
Netherlands	k.m.tych@rug.nl
Germany	ucandenizdavide@gmail.com
Turkey	fucar14@ku.edu.tr
Germany	abhibiotechnologist@gmail.com
Netherlands	atomolecule357@gmail.com
United States	
Mexico	hanif@hms.harvard.edu
	jazhiel@ciencias.unam.mx
Mexico	biologoisrael@gmail.com
Italy	luca.valer@unitn.it
Mexico	domi.06.07.13@gmail.com
Netherlands	peer.vanduppen@ru.nl
Netherlands	J.H.vanEsch@tudelft.nl
United States	albertovazquez@ucla.edu
Mexico	cindelvergara@ciencias.unam.mx
France	roshan-kumar.vijendravarma@curie.fr
Colombia	majo8020@gmail.com
India	vishwakarmah68@gmail.com
Germany	julian.vogel@uni-ulm.de
Germany	kilian.vogele@tum.de
Austria	s.vollenhofer-schrumpf@gmx.at
Germany	beespc@cup.uni-muenchen.de
Germany	vorobii@dwi.rwth-aachen.de
France	yuliia.vyborna@upmc.fr
France	mykhailo.vybornyi@espci.fr
United Kingdom	j.vyle@qub.ac.uk
Germany	awagner@dwi.rwth-aachen.de
Germany	Stefan.waldenmaier@origins-cluster.de
United Kingdom	crw59@cam.ac.uk
Australia	anna.wang@unsw.edu.au
United States	barry_wanner@hms.harvard.edu
Germany	weber@pks.mpg.de
Germany	lweidmann@tuebingen.mpg.de
United Kingdom	peterw@liverpool.ac.uk
Germany	m.weingart@physik.uni-muenchen.de
Germany	lweise@biochem.mpg.de
Germany	daniel.weller@min.uni-muenchen.de
Germany	ericwenzel@gmx.net

tom.257@hotmail.co.uk

stefan.wiedemann@cup.lmu.de

s e.V., RWTH Aachen University Ger

United Kingdom

Germany

ex Systems

S.No	Last Name	First Name	Affiliations	Country	Email Address	S.No	Last Name	First Name	Affiliations
701.	Wienand	Karl	Deutsches Museum	Germany	k.wienand@deutsches-museum.de	751.		Anonymous	Harvard Medical School
702.	Williams	Ann	University of California, Santa Barbara	United States	annwilliams@ucsb.edu	752.		Anonymous	Harvard University
703.	Winkler	Max	Max Planck Institute for Extraterrestrial Physics	Germany	winkler@mpe.mpg.de	753.		Anonymous	Harvard University
704.	Winkler	Wade	University of Maryland	United States	wwinkler@umd.edu	754.		Anonymous	Heidelberg University
705.	Witzdam	Lena	DWI - Leibniz Institute for Interactive Materials e.V.	Germany	witzdam@dwi.rwth-aachen.de	755.		Anonymous	Heidelberg University
706.	Wong	EB	Sultan Idris Education University	Malaysia	eeb109@gmail.com	756.		Anonymous	Heidelberg University
707.	Wong	Michael L	University of Washington	United States	miquai@uw.edu	757.		Anonymous	IIT Guwahati
708.	Wu	Yi-Lin	Cardiff University	United Kingdom	wuyl@cardiff.ac.uk	758.		Anonymous	Indian Institute of Science Education and Research (I
709.	Wu	Longfei	MRC Laboratory of Molecular Biology	United Kingdom		759.		Anonymous	Indian Institute of Science Education and Research (I
710.	Wunnava	Sreekar	Ludwig Maximilian University, Munich	Germany	s.wunnava@physik.uni-muenchen.de	760.		Anonymous	Istanbul Sehir University
711.	Würbser	Michaela	Technical University of Munich	Germany	michaela.wuerbser@tum.de	761.		Anonymous	ISTP - CNR
712.	Xing	Yanyan	LR pharmaceuticals	United States	xing.yyan@icloud.com	762.		Anonymous	Kyoto University
713.	Xu	Jianfeng	MRC Laboratory of Molecular Biology	United Kingdom	jxu@mrc-Imb.cam.ac.uk	763.		Anonymous	Ludwig Maximilian University, Munich
714.	Xue	Lin	Center for Molecular Medicine Norway	Norway	linxu@ncmm.uio.no	764.		Anonymous	Ludwig Maximilian University, Munich
715.	Yadav	Naveen	Max Planck Institute for Astrophysics & ORIGINS	Germany	ny@mpa-garching.mpg.de	765.		Anonymous	Ludwig Maximilian University, Munich
716.	Yay	Cansu	Marmara University	Turkey	cansuyay@marun.edu.tr	766.		Anonymous	Ludwig Maximilian University, Munich
717.	Yeh	Aaron	INIFAP	Mexico	ayega17@gmail.com	767.		Anonymous	Ludwig Maximilian University, Munich
718.	Yeh Martin	Noel	Ludwig Maximilian University, Munich	Germany	noel.martin@physik.uni-muenchen.de	768.		Anonymous	Ludwig Maximilian University, Munich
719.	Yewdall	Amy	Eindhoven University of Technology	Netherlands	n.a.yewdall@tue.nl	769.		Anonymous	Ludwig Maximilian University, Munich
720.	Yi	Jing	Institut de Science et d'Ingénierie Supramoléculaires	France	jing.yi@unistra.fr	770.		Anonymous	Ludwig Maximilian University, Munich
721.	Yomo	Tetsuya	East China Normal University	China	tetsuyayomo@gmail.com	771.		Anonymous	Marmara University
722.	Yüksel	ilke		Turkey	ilkeeyuksell@gmail.com	772.		Anonymous	Max Planck Institue for Dynamics and Self-Organizat
723.	Yuvraj	Angad	University of Delhi	India	yuvi9292@gmail.com	773.		Anonymous	Max Planck Institute for Developmental Biology
724.	Zambrano	Pablo	Max Planck Institute of Biochemistry	Germany	zambranolobos@biochem.mpg.de	774.		Anonymous	Max Planck Institute for Molecular Cell Biology and G
725.	Zanchetta	Giuliano	Università degli Studi di Milano	Italy	giuliano.zanchetta@unimi.it	775.		Anonymous	Max Planck Institute for Terrestrial Microbiology
726.	Zeilinger	Carsten	Leibniz University Hannover	Germany	zeilinger@cell.uni-hannover.de	776.		Anonymous	Max Planck Institute of Colloids and Interfaces
727.	Zeravcic	Zorana	ESPCI Paris	France	zorana.zeravcic@espci.fr	777.		Anonymous	Max Planck Institute of Colloids and Interfaces
728.	Zetterlind	Bobby		Sweden	zetterlind.bobby@gmail.com	778.		Anonymous	MIT
729.	Zetterlind	Alexandra		Sweden	alexandra.zetterlind@gmail.com	779.		Anonymous	NASA
730.	Zhang	Wen	Indiana University School of Medicine	United States	wz15@iu.edu	780.		Anonymous	Ninguno
731.	Zhao	Lei	Chinese Academy of Sciences	China	zhaolei@mail.iggcas.ac.cn	781.		Anonymous	Open University
732.	Zhao	Jianguo	Duke University	United States	jianguo.zhao@duke.edu	782.		Anonymous	Rensselaer Polytechnic Institute
733.	Zhao	Hang	LCPO, UBx-CNRS-BxINP	France	hang.zhao@enscbp.fr	783.		Anonymous	Technical University of Munich
734.	Zhou	Lijun	нни	United States	Izhou@molbio.mgh.harvard.edu	784.		Anonymous	Tohoku University
734.	2100	Anonymous	Bilkent University	Turkey	121104@httpiblo.htgh.harvard.edu	785.		Anonymous	Universidad Nacional Autónoma de México
736.		Anonymous	Bruker Corporation	Turkey		786.		Anonymous	Université Grenoble Alpes
730.			Bruker Corporation	United States		787.			University College London
		Anonymous						Anonymous	
738.		Anonymous	Chuo University	Japan		788.		Anonymous	University of Bristol
739.		Anonymous	DTU - Technical University of Denmark	Denmark		789.		Anonymous	University of Calgary
740.		Anonymous	Earth-Life Science Institute, Tokyo Institute of Technology	Japan		790.		Anonymous	University of Calgary
741.		Anonymous	Eindhoven University of Technology	Netherlands		791.		Anonymous	University of Calgary
742.		Anonymous	Erasmus University Rotterdam	Netherlands -		792.		Anonymous	University of California, Los Angeles
743.		Anonymous	ESPCI Paris	France		793.		Anonymous	University of California, Los Angeles
744.		Anonymous	ESPCI Paris	France		794.		Anonymous	University of Edinburgh
745.		Anonymous	ESPCI Paris - PSL	France		795.		Anonymous	University of Florida
746.		Anonymous	ETH Zurich	Switzerland		796.		Anonymous	University of Groningen
747.		Anonymous	ETH Zurich	Switzerland		797.		Anonymous	University of Kiel
748.		Anonymous	Freie Universität Berlin	Germany		798.		Anonymous	University of Regensburg
749.		Anonymous	Georgia Institute of Technology	United States		799.		Anonymous	University of Regensburg
750.		Anonymous	Ghent University	Belgium		800.		Anonymous	University of Stuttgart









United States United States United States Germany Germany Germany India India India Turkey Italy Japan Germany Germany Germany Germany Germany Germany Germany Germany Turkey Germany Germany Germany Germany Germany Germany United States United States Chile United Kingdom United States Germany Japan Mexico France United Kingdom United Kingdom Canada Canada Canada United States United States United Kingdom United States Netherlands Germany Germany Germany Germany

ch (IISER), Mohali ch (IISER), Mohali

nization

nd Genetics

S.No	Last Name	First Name	Affiliations	Country
801.		Anonymous	University of Stuttgart	Germany
802.		Anonymous	University of Tokyo	Japan
803.		Anonymous	University of Tokyo	Japan
804.		Anonymous	University of Tokyo	Japan
805.		Anonymous	University of Tokyo	Japan
806.		Anonymous		Germany
807.		Anonymous		India
808.		Anonymous		India
809.		Anonymous		Mexico

CREDITS

**Bookcover Illustration Design** Priyanka Kaur Oberoi priyankakauroberoi@gmail.com • https://cargocollective.com/priyankaoberoi

**Publication Design** Roopali Sood designbytesinfo@gmail.com



Email Address







MOLECULAR ORIGINS OF LIFE, MUNICH Literaturhaus Munchen